

DOCUMENT-IDENTIFIER: US 6328885 B1
TITLE: Current-efficient suppressors

DEPR:

The term "packing" refers to stationary flow-through solid material disposed in a flow channel of the suppressor. It can be a screen or a porous monolithic matrix, a resin particle bed or other form. It can be strongly charged, weakly charged or of neutral charge, as will be explained. The term packing is alternatively called "bridging means."

DEPR:

In the above system, one way to increase current efficiency is leave the sample stream flow channel open without packing or to use packing which is of neutral charge or of low capacity relative to the packing of high capacity ion exchange material in the ion receiving flow channel and, for a two membrane suppressor, in the ion source channel. While the above description refers to the stationary flow-through packing of ion exchange material in the form of a high capacity charged screen, other forms of packing may also be employed as described above. Such other packing forms of ion exchange material include packed beds of ion exchange resin or monolithic materials of charged material with sufficient porosity for the flow of an aqueous liquid stream through them. The packing in the ion receiving channel has a substantially higher capacity than ion exchange packing in the sample flow channel, if present. Thus, if a charged packing is used in the sample stream flow channel, it preferably is of low capacity, with a capacity of substantially less than that of the packing in the ion receiving flow channel. Suitably, the ratio of total capacities of the packing in the sample stream flow channel to that in the ion receiving stream

flow channel is no greater than about 0.9, and preferably no greater than about 0.7 to 0.5, and more preferably no greater than about 0.1.

CCOR:

210/198.2

DOCUMENT-IDENTIFIER: US 6325976 B1
TITLE: Continuous electrolytically regenerated packed bed
suppressor for ion
chromatography

CCXR:
210/198.2

ORFL:
Petro et al., "Molded Monolithic Rod of Macroporous
Poly(styrene-co-divinylbenzene) as a Separation Medium for HPLC of
Synthetic
Polymers: "On-Column" Precipitation -Redissolution Chromatography
as an
Alternative to Size Exclusion Chromatography of Styrene Oligomers
and dPolymers"
Analytical Chemistry, 68(2):315-321 (Jan. 15, 1996).

DOCUMENT-IDENTIFIER: US 6309549 B1

TITLE: Polynucleotide separations on polymeric separation

ABPL:

Non-polar polymeric separation media, such as beads or monoliths, are suitable for chromatographic separation of mixtures of polynucleotides when the surfaces of the media are unsubstituted or substituted with a hydrocarbon group having from one to 1,000,000 carbons and when the surfaces are substantially free from multivalent cation contamination. The polymeric media provide efficient separation of polynucleotides using Matched Ion Polynucleotide Chromatography. Methods for maintaining and storing the polymeric media include treatment with multivalent cation binding agents.

ESPR:

The present invention is directed to the separation of polynucleotides using non-polar separation surfaces, such as the surfaces of polymeric beads and surfaces within molded monoliths, which are substantially free from contamination with multivalent cations.

ESPR:

Another object of the present invention is to provide a method for separating polynucleotides using nonporous polymer separation media, such as beads or monoliths (e.g., rods), having non-reactive, non-polar surfaces.

ESPR:

In one aspect, the invention is a method for separating a mixture of polynucleotides by applying a mixture of polynucleotides having up to 1500 base pairs to a polymeric separation medium having non-polar surfaces which are substantially free from contamination with multivalent cations, and eluting the mixture of polynucleotides. The preferred surfaces are nonporous. The

non-polar surfaces can be enclosed in a column. In the preferred embodiment, precautions are taken during the production of the medium so that it is substantially free of multivalent cation contaminants and the medium is treated, for example by an acid wash treatment and/or treatment with multivalent cation binding agent, to remove any residual surface metal contaminants. The preferred separation medium is characterized by having a DNA Separation Factor (defined hereinbelow) of at least 0.05. The preferred separation medium is also characterized by having a Mutation Separation Factor (as defined hereinbelow) of at least 0.1. In the preferred embodiment, the separation is made by Matched Ion Polynucleotide Chromatography (MIPC, as defined hereinbelow). Examples of non-polar surfaces include the surfaces of polymer beads and the surfaces of interstitial spaces within a polymeric **monolith**. The elution step preferably uses a mobile phase containing a counterion agent and a water-soluble organic solvent. Examples of a suitable organic solvent include alcohol, nitrile, dimethylformamide, tetrahydrofuran, ester, ether, and mixtures of one or more thereof, e.g., methanol, ethanol, 1-propanol, 2-propanol, tetrahydrofuran, ethyl acetate, acetonitrile. The most preferred organic solvent is acetonitrile. The counterion agent is preferably selected from the group consisting of lower alkyl primary amine, lower alkyl secondary amine, lower alkyl tertiary amine, lower trialkylammonium salt, quaternary ammonium salt, and mixtures of one or more thereof. Non-limiting examples of counterion agents include butylammonium acetate, triethylammonium acetate, dodecylammonium acetate, pyridiniumammonium acetate, cyclonexylammonium acetate, diethylammonium acetate, propylethylammonium acetate, propyldiethylammonium acetate, butylethylammonium acetate, methylhexylammonium acetate,

tetramethylammonium
acetate, tetrapropylammonium acetate, tetrabutylammonium acetate,
dimethyldiethylammonium acetate, triethylammonium acetate,
tripropylammonium
acetate, tributylammonium acetate, tetraethylammonium acetate,
tetrapropylammonium acetate, tetrabutylammonium acetate, and
mixtures of any
one or more of the above. The counterion agent includes an
anion, e.g.,
acetate, carbonate, bicarbonate, phosphate, sulfate, nitrate,
propionate,
formate, chloride, perchlorate, or bromide. The most preferred
counterion
agent is triethylammonium acetate or triethylammonium
hexafluoroisopropyl
alcohol.

BSFR:

In yet another embodiment, the invention is a method for
separating a mixture
of polynucleotides comprising flowing a mixture of
polynucleotides having up to
1500 base pairs through a polymeric monolith, and separating the
mixture of
polynucleotides using MIPC. In this embodiment, the non-polar
separation
surfaces are the surfaces of interstitial spaces of a polymeric
monolith. An
example of such a monolith is a polymeric rod prepared within the
confines of a
chromatographic column. The monolith of the invention is
characterized by
having a DNA Separation Factor of at least 0.05. In a preferred
embodiment,
the monolith is characterized by having a DNA Separation Factor
of at least
0.5. The monolith is preferably characterized by having a
Mutation Separation
Factor of at least 0.1. The mobile phase used in the separation
preferably
includes an organic solvent as exemplified by alcohol, nitrite,
dimethylformamide, tetrahydrofuran, ester, ether, and mixtures
thereof.
Examples of suitable solvents include methanol, ethanol,
2-propanol,
1-propanol, tetrahydrofuran, ethyl acetate, acetonitrile, and
mixtures thereof.
The most preferred organic solvent is acetonitrile. The mobile
phase
preferably includes a counterion agent such as lower primary,

secondary and tertiary amines, and lower trialkylammonium salts, or quaternary ammonium salts. More specifically, the counterion agent can be octylammonium acetate, octadimethylammonium acetate, decylammonium acetate, octadecylammonium acetate, pyridiniumammonium acetate, cyclohexylammonium acetate, diethylammonium acetate, propylethylammonium acetate, propyldiethylammonium acetate, butylethylammonium acetate, methylhexylammonium acetate, tetramethylammonium acetate, tetraethylammonium acetate, tetrapropylammonium acetate, tetrabutylammonium acetate, dimethyldiethylammonium acetate, triethylammonium acetate, tripropylammonium acetate, tributylammonium acetate, tetrapropylammonium acetate, tetrabutylammonium acetate, and mixtures of any one or more of the above. The counterion agent includes an anion, e.g., acetate, carbonate, bicarbonate, phosphate, sulfate, nitrate, propionate, formate, chloride, perchlorate, and bromide. However, the most preferred counterion agent is triethylammonium acetate.

ESPE:

In the preferred embodiment, precautions are taken during the production of the polymeric monolith so that it is substantially free of multivalent cation contaminants and the monolith is treated, for example, by an acid wash treatment, to remove any residual surface metal contaminants. In the embodiment, the monolith is characterized by having a DNA Separation Factor of at least 6.65. In a preferred embodiment, the monolith is characterized by having a DNA separation factor of at least 9.5. Also in a preferred embodiment, the monolith is characterized by having a Mutation Separation Factor of at least 0.1.

ESPE:

In another aspect, the present invention is a method for treating the non-polar surface of a polymeric medium used for separating polynucleotides

such as the surface of beads in a MIPC column or the interstitial spaces in a polymeric monolith, in order to improve the resolution of polynucleotides, such as dsDNA, separated on said surface. This treatment includes contacting the surface with a solution containing a multivalent cation binding agent. In a preferred embodiment, the solution has a temperature of about 50.degree. C. to 90.degree. C. An example of this treatment includes flowing a solution containing a multivalent cation binding agent through a MIPC column, wherein the solution has a temperature of about 50.degree. C. to 90.degree. C. The preferred temperature is about 70.degree. C. to 80.degree. C. In a preferred embodiment, the multivalent cation binding agent is a coordination compound, examples of which include water-soluble chelating agents and crown ethers. Specific examples include acetylacetone, alizarin, aluminon, chloranilic acid, kojic acid, morin, rhodizonic acid, thionalide, thiourea, .alpha.-furildioxime, niexime, salicylaldoxime, dimethylglyoxime, .alpha.-furildioxime, cupferron, .alpha.-nitroso-.beta.-naphthol, nitroso-R-salt, diphenylthiocarbazone, diphenylcarbazone, eriochrome black T, PAN, SPADNS, glyoxal-bis(2-hydroxyanil), murexide, .alpha.-benzoinoxime, mandelic acid, anthranilic acid, ethylenediamine, glycine, triaminotriethylamine, thionalide, triethylenetetramine, ethylenediaminetetraacetic acid (EDTA), metalphthalein, arsenic acids, .alpha.,.alpha.'-bipyridine, 4-hydroxybenzothiazole, 8-hydroxyquinoline, 8-hydroxyquinoline, 1,10-phenanthroline, picolinic acid, salicylic acid, .alpha.,.alpha.',.alpha."-terpyridyl, 2,2'-bipyridyl, 2,2'-bipyridyl-5,5'-dicarboxylic acid, pyrocatechol, salicylic acid, tiron, 1,2-dimercaptobenzene, dithiol, mercaptobenzothiazole, maleic acid, oxalic acid, sodium diethyldithiocarbamate, and zinc dibenzylidithiocarbamate. However, the most preferred chelating agent is EDTA. In this aspect of the invention, the solution preferably includes

an organic solvent as exemplified by alcohol, triole, dimethylformamide, tetrahydrofuran, ester, ether, and mixtures thereof. Examples of suitable solvents include methanol, ethanol, 2-propanol, 1-propanol, tetrahydrofuran, ethyl acetate, acetonitrile, and mixtures thereof. The most preferred organic solvent is acetonitrile. In one embodiment, the solution can include a counterion agent such as lower primary, secondary and tertiary amines, and lower trialkylammonium salts, or quaternary ammonium salts. More specifically, the counterion agent can be octylammonium acetate, octadimethylammonium acetate, decylammonium acetate, octadecylammonium acetate, pyridiniumammonium acetate, cyclohexylammonium acetate, diethylammonium acetate, propylethylammonium acetate, propyldiethylammonium acetate, butylethylammonium acetate, methylhexylammonium acetate, tetramethylammonium acetate, tetraethylammonium acetate, tetrapropylammonium acetate, tetrabutylammonium acetate, dimethyldiethylammonium acetate, triethylammonium acetate, tripropylammonium acetate, tributylammonium acetate, tetrapropylammonium acetate, tetrabutylammonium acetate, and mixtures of any one or more of the above. The counterion agent includes an anion, e.g., acetate, carbonate, bicarbonate, phosphate, sulfate, nitrate, propionate, formate, chloride, perchlorate, and bromide. However, the most preferred counterion agent is triethylammonium acetate.

ESPR:

In yet a further aspect, the invention provides a method for storing a medium used for separating polynucleotides, e.g., the beads of a MIP column. The polymer is monolith, in order to improve the resolution of double stranded DNA fragments separated using the medium. In the case of a MIP column, the preferred method includes flowing a solution containing a multivalent cation binding agent through the column prior to storing the column. In

a preferred embodiment, the multivalent cation binding agent is a coordination compound, examples of which include water-soluble chelating agents and crown ethers. Specific examples include acetylacetone, alizarin, aluminon, chloranilic acid, kojic acid, morin, rhodizonic acid, thionalide, thiourea, .alpha.-furildioxime, nioxime, salicylaldoxime, dimethylglyoxime, .alpha.-furildioxime, cupferron, .alpha.-nitroso-.beta.-naphthol, nitroso-R-salt, diphenylthiocarbazone, diphenylcarbazone, eriochrome black T, PAN, SPADNS, glyoxal-bis(2-hydroxyanil), murexide, a-benzoinoxime, mandelic acid, anthranilic acid, ethylenediamine, glycine, triaminotriethylamine, thionalide, triethylenetetramine, EDTA, metalphthalein, arsonic acids, .alpha.,.alpha.'-bipyridine, 4-hydroxybenzothiazole, 8-hydroxyquinoline, 8-hydroxyquinoline, 1,10-phenanthroline, picolinic acid, quinaldic acid, .alpha.,.alpha.',.alpha."-terpyridyl, 6-methyl-2,3,7-trihydroxy-6-fluorone, pyricatechol, salicylic acid, tiron, 4-chloro-1,2-dimercaptobenzene, dithiol, mercaptobenzothiazole, rubeanic acid, oxalic acid, sodium diethyldithiocarbamate, and zinc dibenzylthiocarbamate. However, the most preferred chelating agent is EDTA. In this aspect of the invention, the solution preferably includes an organic solvent as exemplified by alcohols, nitriles, dimethylformamide, tetrahydrofuran, esters, and ethers. The most preferred organic solvent is acetonitrile. The solution can also include a counterion agent such as lower primary, secondary and tertiary amines, and lower trialkylammonium salts, or quaternary ammonium salts. More specifically, the counterion agent can be octylammonium acetate, decylammonium acetate, dodecylammonium acetate, pyridinium acetate, cyclohexylammonium acetate, diethylammonium acetate, propylethylammonium acetate, propyldiethylammonium acetate, butylethylammonium acetate, methylhexylammonium acetate, tetramethylammonium acetate, tetraethylammonium acetate, tetrapropylammonium acetate,

tetrabutylammonium
acetate, dimethyldiethylammonium acetate, triethylammonium
acetate,
tripropylammonium acetate, tributylammonium acetate,
tetrapropylammonium
acetate, tetrabutylammonium acetate, and mixtures of any one or
more of the
above. The counterion agent includes an anion, e.g., acetate,
carbonate,
bicarbonate, phosphate, sulfate, nitrate, propionate, formate,
chloride,
perchlorate, and bromide. However, the most preferred counterion
agent is
triethylammonium acetate.

DEBB:

The medium can be enclosed in a column. In one embodiment, the
non-polar
surfaces comprise the surfaces of polymeric beads. In an
alternative
embodiment, the surfaces comprise the surfaces of interstitial
spaces in a
molded polymeric monolith. For purposes of simplifying the
description of the
invention and not by way of limitation, the separation of
polynucleotides using
nonporous beads, and the preparation of such beads, will be
primarily described
herein, it being understood that other separation surfaces, such
as the
interstitial surfaces of polymeric monoliths, are intended to be
included
within the scope of this invention. Monoliths such as rods
contain polymer
separation media which have been formed inside a column as a
unitary structure
having through pores or interstitial spaces which allow eluting
solvent and
analyte to pass through and which provide the non-polar
separation surface.

DEBB:

In another embodiment of the present invention, the separation
medium can be in
the form of a polymeric monolith such as a rod like monolithic
column. The
monolithic column is polymerized or formed as a single unit
inside of a tube as
described in the Examples hereinbelow. The through pore or
interstitial spaces

provide for the passage of eluting solvent and analyte materials.

The separation is performed on the stationary surface. The surface can be porous, but is preferably nonporous. The form and function of the separations are identical to columns packed with beads. As with beads, the pores contained in the rod must be compatible with DNA and not trap the material. Also, the rod must not contain contamination that will trap DNA.

DEPE:

The molded polymeric rod of the present invention is prepared by bulk free radical polymerization within the confines of a chromatographic column. The base polymer of the rod can be produced from a variety of polymerizable monomers. For example, the monolithic rod can be made from polymers, including mono- and di-vinyl substituted aromatic compounds such as styrene, substituted styrenes, alpha-substituted styrenes and divinylbenzene; acrylates and methacrylates; polyolefins such as polypropylene and polyethylene; polyesters; polyurethanes; polyamides; polycarbonates; and substituted polymers including fluorosubstituted ethylenes commonly known under the trademark TEFLON. The base polymer can also be mixtures of polymers, non-limiting examples of which include poly(glycidyl methacrylate-co-ethylene dimethacrylate), poly(styrene-divinylbenzene) and poly(ethylvinylbenzene-divinylbenzene). The rod can be unsubstituted or substituted with a substituent such as a hydrocarbon alkyl or an aryl group. The alkyl group optionally has 1 to 1,000,000 carbons inclusive in a straight or branched chain, and includes straight chained, branched, cyclic, saturated, unsaturated, aromatic, and functional groups of various types including aldehyde, ketone, ester, ether, alkyl groups, and the like, and the aryl groups includes as monocyclic, bicyclic, and tricyclic aromatic hydrocarbon groups including phenyl, naphthyl, and the like. In a

preferred embodiment, the alkyl group has 1-24 carbons. In a more preferred embodiment, the alkyl group has 1-8 carbons. The substitution can also contain hydroxy, cyano, nitro groups, or the like which are considered to be non-polar, reverse phase functional groups. Methods for hydrocarbon substitution are conventional and well-known in the art and are not an aspect of this invention.

The preparation of polymeric monoliths is by conventional methods well known in the art as described in the following references: Wang et al. (J Chromatog. A 699:230 (1994)), Petro et al. (Ana. Chem. 68:315 (1996)), and the following U.S. Pat. Nos. 5,334,310; 5,453,185; 5,522,994 (to Frechet).

Monolith or rod columns are commercially available from Merck & Co (Darmstadt, Germany).

DEPR:

A chromatography tube in which the monolith polymeric separation medium is prepared is made of stainless steel. The monomers, styrene (Sigma--Aldrich Chemical Corp.) and divinylbenzene (Dow Chemical Corp.) are dried over magnesium sulfate and distilled under vacuum.

DEPR:

Following polymerization, the rubber plugs are replaced by column end fittings and the column is connected to an HPLC system. The HPLC instrument has a low-pressure mixing quaternary gradient capability. A cartridge or guard column containing an iminodiacetate multivalent cation capture resin is placed in line between the column and the mobile phase source reservoir.

The column is then washed by flowing 100 mL of tetrahydrofuran (THF) at 1 mL/min. through the column to remove the dodecyl alcohol and toluene, thereby creating through-pores in the otherwise solid polymer monolith.

DEPR:

The non-polar, organic polymer monolith column is washed by flowing

tetrahydrofuran through the column at a flow rate of 2 mL per minute for 10 minutes followed by flowing methanol through the column at 2 mL per minute for 10 minutes. The non-polar, organic polymer monolith column is washed further by flowing a mixture containing 100 mL of tetrahydrofuran and 100 mL of concentrated hydrochloric acid through the column at 10 mL per minute for minutes. Following this acid treatment, the non-polar, organic polymer monolith column is washed by flowing tetrahydrofuran/water (1:1) through the column at 2 mL per minute until neutral (pH 7).

DEPF:

Any double bonds remaining on the surface of the monolith column prepared in Example 9 are reacted with bromine as described in Example 7.

IEPC:

Preparation of a Non-Polar Organic Polymer Monolith Chromatography Column

DEPC:

Bromination of Remaining Double Bonds on the Surface of Non-Polar Organic Polymer Monolith Column

DEPC:

Nitration of a Non-Polar Organic Polymer Monolith Column

CCXP:

210/198.2

ORPL:

Nakanishi et al. Double Pore Silica Gel Monolith Applied to Liquid Chromatography, In: Sol-Gel Science & Technology, vol. 8, pp. 547-552, 1997.

ORPL:

Petro et al, Modified Monolithid Rod of Macroporous Poly(Sytrene-Co-Divinylbenzene) as a Separation Medium for HPLC of Synthetic Polymer . . . , Analytical Chemistry, 68: 315-321 (1996).

DOCUMENT-IDENTIFIER: US 6296771 B1

TITLE: Parallel high-performance liquid chromatography with serial injection

BSFR:

Gel permeation chromatography (GPC), a well-known form of size exclusion

chromatography (SEC), is a frequently-employed chromatographic technique for

separation of samples generally, and for polymer size determination particular.

Another chromatographic separation approach is illustrated by U.S. Pat. No.

5,334,310 to Frechet et al. and involves the use of a porous monolithic

stationary-phase as a separation medium within the

chromatographic column,

combined with a mobile-phase composition gradient. Other

separation approaches

are also known in the art, including for example, normal-phase (e.g.,

adsorption) chromatography and reverse-phase chromatography, hydrophobic

interaction chromatography, hydrophilic interaction

chromatography,

ion-exchange chromatography, affinity chromatography, among others.

DEPR:

The chromatographic column 102 (or a series of columns in one or more of the

chromatographic channels) further comprises a separation medium having a

stationary-phase within the separation cavity. The separation medium can

consist essentially of a stationary-phase or can also include, in addition,

thereto, an inert support for the stationary phase. The column 102 can also

comprise one or more filters, frits (for separation medium retention and/or for

filtering), and various fittings and features appropriate for preparing and/or

maintaining the column for its intended application. The particular separation

medium to be employed as the stationary-phase is not critical, and will

typically depend on the separation strategy for the particular chemistry of the polymer samples of interest, as well as on the desired detection, sample-throughput and/or information quality. Typical stationary-phase media can be a bed of packed beads, fibers, irregular or other shaped-particles, or a monolithic medium (typically greater than about 5 mm in thickness), each of which can be characterized and optimized for a particular separation strategy with respect to the material, size, shape, pore size, pore size distribution, surface area, solvent regain, bed homogeneity (for packed shaped-particles), inertness, polarity, hydrophobicity, chemical stability, mechanical stability and solvent permeability, among other factors. Generally preferred stationary-phase include porous media (e.g., porous beads, porous monoliths), such as are suitable for gel permeation chromatography (GPC), precipitation-redissolution chromatography, normal-phase (e.g., adsorption) chromatography and reverse-phase chromatography, hydrophobic interaction chromatography, hydrophilic interaction chromatography, ion-exchange chromatography, affinity chromatography, among others. Non-porous particles or empty columns and/or capillaries with adsorptive walls can be used as well. If beads are employed, spherical beads are preferred over other shapes. Particularly preferred stationary-phase media for polymer characterization applications are disclosed in greater detail below, but can generally include silica, cross-linked polymeric resins (e.g., poly(2-3-dihydroxypropylmethacrylate), poly(hydroxyethyl methacrylate), and polystyrenic polymers such as poly(styrene-divinylbenzene)).

END:

210/198.2

DOCUMENT-IDENTIFIER: US 6290909 B1

TITLE: Sample injector for high pressure liquid chromatography

DEFR:

A sample injector A is also connected to common junction 335 between

hydrostatic pressure source 310 and HPLC column 320. The injector is comprised

of at least two elements such as 340 and 350. Each element comprises a

container having an inlet and outlet end and filled with a dielectric material

to form a porous bed within the container. Containers can include any

geometric configuration capable of containing the porous bed of dielectric

material, such as capillary tubes, and capable of withstanding pressures of up

to about 40,000 psi. Also included are microchannels fabricated on a substrate

such as those described by Paul et al. in U.S. Pat. Nos. 6,913,164 and

6,919,882 and by Arnold in prior co-pending U.S. patent application Ser. No.

09/404,945, filed Sep. 9, 1999, now U.S. Pat. No. 6,210,986, entitled

"Microfluidic Channel Fabrication Method" assigned to the same assignee.

Elements 340 and 350 are connected together in series configuration with a

common junction 335. The dielectric material filling each container is

selected so as to minimize any chromatographic separation of the sample and can

be any non-porous material, known to those skilled in the art, used to form a

porous, packed bed. By way of example, the dielectric material can be

comprised of coated and nonporous silica, glass, or polymer beads or is a

monolithic polymer material. Further, the dielectric material is selected to

resist pressure-driven flow but to allow

electroosmotically-driven flow. Thus,

additionally it is preferred that the pore diameter of the porous bed be in the

range of about 25 to 300 nm. One of elements 340/350 serves as a

sample inlet
and is in communication with a sample container 360 and the other
is connected
to a waste reservoir 370. A power supply 375 is connected across
both elements
of sample injector A. Placing one of the electrodes of power
supply 375 in the
sample container and the other in the waste reservoir can make
this connection.
Alternatively, power supply 375 can be connected to the elements
of the sample
injector by means of salt bridges, as discussed above.

CCXR:

210/198.2

DOCUMENT-IDENTIFIER: US 6287822 B1
TITLE: Mutation detection method

DEPR:

MIPC uses unique non-polar separation media which comprises organic polymers, silica media having a non-polar surface comprising coated or covalently bound organic polymers or covalently bound alkyl and/or aryl groups, continuous non-polar separation media, so called monolith or rod columns, comprising non-polar silica gel and organic polymer. The separation media used in MIPC can be porous or non-porous. A detailed description of the MIPC separation process, MIPC separation media, and MIPC systems is found in U.S. Pat. No. 5,772,669 (1998) to Gjerde and in co-pending U.S. patent application Ser. No. 09/066,560 filed Apr. 10, 1998; Ser. No. 09/058,337 filed Apr. 10, 1998 now abandoned; Ser. No. 09/065,913 filed Apr. 24, 1998; now U.S. Pat. No. 5,986,085 Ser. No. 09/081,040 filed May 18, 1998; now U.S. Pat. No. 5,997,742 Ser. No. 09/081,039 filed May 18, 1998; now U.S. Pat. No. 5,972,122 and Ser. No. 09/080,547 filed May 18, 1998 now U.S. Pat. No. 6,017,457. MIPC systems and separation media are commercially available (Transgenomic, Inc. San Jose, Calif.)

CCXR:

210/198.2

ORPL:

Retro et al., Molded Monolithid Rod of Macrophorous Polyethylene - ~~Monolithid~~ as a Separation Medium for HPLC of Synthetic Polymers . . . , Analytical Chemistry, 61:115-121 (1990).

TITLE: High-temperature characterization of polymers

Liquid chromatography is well known in the art for characterizing a polymer

components of a polymer sample from other components thereof by flow through a

a flow-through detector. Approaches for liquid chromatography can vary.

form of size. Gel permeation chromatography (GPC), a well-known

technique for polymer size determination. In GPC, the polymer sample is

component in solution. More specifically, a polymer sample is injected into a

more chromatographic columns packed with porous beads. Molecules with

remain therein for longer periods, and therefore exit the column after

characterize one or more separated components of the polymer sample.

separation approach is illustrated by G.S. Ant. III, 3, 4, 5
 11. Krogher et al.

separation medium within the chromatographic column, combined with a

12/18/2001, EAST Version: 1.02.0008

Molded Monolithic

Rod of Macroporous Poly(styrene-co-divinylbenzene) as a Separation Medium for HPLC Synthetic Polymers: "On-Column" Precipitation-Redissolution Chromatography as an Alternative to Size Exclusion Chromatography of Styrene Oligomers and Polymers, Anal. Chem., 68, 315-321 (1996); and Petro et al, Immobilization of Trypsin onto "Molded" Macroporous Poly (Glycidyl Methacrylate-co-Ethylene Dimethacrylate) Rods and Use of the Conjugates as Bioreactors and for Affinity Chromatography, Biotechnology and Bioengineering, Vol. 49, pp. 355-363 (1996)). Chromatography involving the porous monolith is reportedly based on a precipitation/redissolution phenomenon that separates the polymer according to size--with the precipitated polymer molecules selectively redissolving as the solvent composition is varied. The monolith provides the surface area and permeation properties needed for proper separation. Other separation approaches are also known in the art, including for example, normal-phase adsorption chromatography (with separation of polymer components being based on preferential adsorption between interactive functionalities of repeating units and an adsorbing stationary-phase) and reverse-phase chromatography (with separation of polymer components being based on hydrophobic interactions between a polymer and a non-polar stationary-phase). After separation, a detector can measure a property of the polymer or of a polymer component--from which one or more characterizing properties, such as molecular weight can be determined as a function of time. Specifically, a number of molecular-weight related parameters can be determined, including for example: the weight-average molecular weight ($M_{sub.w}$), the number-average molecular weight ($M_{sub.n}$), the molecular-weight distribution shape, and an index of the breadth of the molecular-weight distribution ($M_{sub.w} / M_{sub.n}$), known as the

polydispersity index (PDI). Other characterizing properties, such as mass, particle size, composition or conversion can likewise be determined.

BSPF:

Aspects of polymer characterization, such as sample preparation and polymer separation, have been individually and separately investigated. For example, Poche et al. report a system and approach for automated high-temperature dissolution of polymer samples. See Poche et al., Use of Laboratory Robotics for Gel Permeation Chromatography Sample Preparation: Automation of High-Temperature Polymer Dissolution, J. Appl. Polym. Sci., 64(8), 1613-1623 (1997). Stationary-phase media that reduce chromatographic separation times of individual polymer samples have also been reported. See, for example, Petro et al., Molded continuous poly(styrene-co-divinylbenzene) rod as a separation medium for the very fast separation of polymers; Comparison of the chromatographic properties of the monolithic rod with columns packed with porous and non-porous beads in high-performance liquid chromatography., Journal of Chromatography A, 752, 59-66 (1996); and Petro et al., Monodisperse Hydrolyzed Poly(glycidyl methacrylate-co-ethylene dimethacrylate) Beads as a Stationary Phase for Normal-Phase HPLC, Anal. Chem., 69, 1131 (1997). However, such approaches have not contemplated nor been incorporated into protocols and systems suitable for large-scale, or even moderate-scale, combinatorial chemistry research, and particularly, for combinatorial material science research directed at the characterization of polymers.

DEPF:

The chromatographic column 102 further comprises a separation medium having a stationary-phase within the separation cavity. The separation medium can consist essentially of a stationary-phase or can also include, in

addition thereto, an inert support for the stationary phase. The column 102 can also comprise one or more fillers, frits (for separation medium retention and/or for filtering), and various fittings and features appropriate for preparing and/or maintaining the column for its intended application. The particular separation medium to be employed as the stationary-phase is not critical, and will typically depend on the separation strategy for the particular chemistry of the polymer samples of interest, as well as on the desired detection, sample-throughput and/or information quality. Typical stationary-phase media can be a bed of packed beads, rods or other shaped-particles, or a monolithic medium (typically greater than about 5 mm in thickness), each of which can be characterized and optimized for a particular separation strategy with respect to the material, size, shape, pore size, pore size distribution, surface area, solvent regain, bed homogeneity (for packed shaped-particles), inertness, polarity, hydrophobicity, chemical stability, mechanical stability and solvent permeability, among other factors. Generally preferred stationary-phase include porous media (e.g., porous beads, porous monoliths), such as are suitable for gel permeation chromatography (GPC), and media suitable for precipitation-redissolution chromatography, adsorption chromatography, and/or reverse-phase chromatography. Non-porous particles or empty columns and/or capillaries with adsorptive walls can be used as well. If beads are employed, spherical beads are preferred over other shapes. Particularly preferred stationary-phase materials for liquid chromatization applications are disclosed in greater detail below, but can generally include silica, cross-linked resins, hydroxylated polyglycidyl methacrylates, (e.g., poly(2- β -dihydroxypropylmethacrylate)), poly(hydroxyethyl methacrylate), and polystyrenic polymers such as poly(styrene-divinylbenzene).

DEPF:

In other variations, the short column may comprise column stationary-phase packing other than is typically used for GPC, such as normal-phase or reverse-phase silica particles, polymer monoliths, inorganic monoliths, and other well-known column stationary-phase materials or filter media. For example, short columns containing adsorption chromatography stationary-phase can be used to remove components either more polar or less polar than the polymer sample of interest, such as water or solvents initially introduced with the sample. Also in a preferred aspect of this embodiment, more than one short column may be used in series, for example a short GPC column in combination with a short normal-phase adsorption chromatography column, such that polymer is separated from low-molecular-weight components, which are then further separated by polarity. (See Ex. 20). This can be particularly useful for rapidly separating polymer from residual monomer or solvent in a polymerization reaction, and then further quantifying the type and amount of monomer or solvent within a single, rapid analysis.

DEPF:

While some aspects of the following description refer to "beads", such reference is to be considered exemplary; other stationary-phase media (e.g., rods, monoliths, etc.) can be readily employed instead of such beads.

DEPF:

Pre-precipitation-resolution chromatography involves the use of mobile-phase having a solvent gradient in conjunction with an insoluble stationary-phase (i.e., a polymer monolith). The polymer sample is injected into a mobile-phase solvent that is a "poor" solvent for the polymer being characterized (sometimes called a "non-solvent"), thereby causing precipitation of the

polymer sample.

The precipitated polymer sample then adsorbs onto the stationary-phase (e.g., monolith) surface. Gradually, a better solvent for the polymer being characterized is introduced into the mobile phase. When the better solvent contacts the precipitated polymer sample, the smaller particles of the polymer sample redissolve first. As more of the better solvent contacts the precipitated polymer sample, larger particles of the polymer sample redissolve, until the entire polymer sample has been redissolved. In this fashion, the polymer sample is separated by size (with the smaller particles corresponding to smaller size molecules). Solvent choices depend on the solubility characteristics of the polymer samples being characterized. For a typical hydrophobic polymer such as polystyrene, "good" solvents include tetrahydrofuran, toluene, dichloromethane, etc., while "poor" non-solvents include methanol, ethanol, water, or hexane. It is generally preferred that the good solvent and the poor solvent used for any particular separation be miscible.

DEPR:

The precipitation-redissolution chromatography approaches described herein--particularly employing monolithic columns such as those disclosed by Petro et al., vide supra., generally lead to high-speed characterization with good quality of information.

DEPR:

The stationary phase can be selected according to the type of polymer to be analyzed. Materials recommended for this approach include porous monoliths and beads. Since or hydrophilic polymer beads are used for adsorption of polar polymers or for removing of highly polar components of the samples, such as water, which would otherwise interfere with the analysis of

compounds of interest, such as monomers and polymers. Polymeric beads with diol functionalities are preferred for this purpose since they have higher adsorptivity than silica with minimized non-specific interactions with the characterized polymers (See M. Petro, et al., Anal. Chem., 1997, 69 3131; M. Petro, et al., J. Polym. Sci. A: Polym. Chem., 1997, 35, 1173; J. M. J. Frechet, et. al., Polym. Mater. Sci. Eng. 1997, 77, 38.).

DEPR:

The typical mobile phase (e.g., solvent) used for this adsorption chromatography is tetrahydrofuran, either alone or in mixtures with hexane (to enhance adsorption) or water (to enhance elution). Octadecyl-silica beads (commonly used in conventional reverse-phase HPLC) and polystyrene-based monoliths are used for a separation of compounds of medium polarity under the conditions typical of reversed-phase chromatography, usually in combination with a mixture of water and tetrahydrofuran. Optionally, gradients in connection with this technique can be employed, changing either the composition, temperature or flow rate of the mobile phase.

DEPR:

The robotic auto-sampler and injection valve set-up as in Example 1 was fitted with two sample loops (each having 50 microliter volume) in combination with a high-pressure liquid chromatographic (HPLC) apparatus comprising a two-pump gradient chromatography system, primed with methanol and tetrahydrofuran (THF) solvent. A porous crosslinked polystyrene monolithic column was utilized, prepared as described in Frechet et al., Journal of Chromatography, 1996, 69-66 and Frechet et al., Anal. Chem. 1996, 68, 327. The HPLC system was configured such that the combined flow of the pump system passed through the valve, the column, and then to a UV chromatographic detector. The entire system, including pump control and data acquisition from

the detector
was computer-controlled.

DEPR:

Adsorption chromatography was used for separation of various components of the reaction mixtures that contained the comonomers, (co)polymers, solvents and catalyst components. Good separation was achieved in 60 seconds per sample using a short, high-aspect ratio reversed-phase column and gradient of THF in water with a concave profile. The specific gradient profile allows to separate small molecules with similar retention behavior from each other as well as elute a highly retained polymer in a very short time. Columns of various sizes, porosities and chemistries were used for this purpose including polystyrene-based monoliths and silica-based porous beads.

DEPR:

A single, short, high-aspect ratio column (0.8 cm.times.5 cm) contained a polystyrene monolith as the separation medium and resided in a PE-210 HT-GPC oven maintained at 140.degree. C. The system was configured substantially as shown in FIG. 6 and described in connection therewith and as follows. Two mobile-phase reservoirs 114, 120 were provided and equipped with two Waters 515 pumps 116, 118. A "mobile-phase A" reservoir 114 feeding pump 116 (hereinafter "pump A") comprised trichlorobenzene (TCB) and, in operation, was configured to pump mobile-phase A through the injection valve 210 (100) and through the oven, whereby the mobile-phase A was heated to become the hot mobile phase (i.e., hot TCB). A "mobile-phase B" reservoir 120 feeding pump 118 (hereinafter "pump B") comprised trichlorobenzene, and in operation, was configured to pump mobile-phase B to bypass most of the heated environment, and to enter the oven immediately prior to the column 102 as an essentially ambient-temperature mobile phase (i.e., cold TCB). Detection was effected with a PD

2000
light-scattering detector (90.degree.).

CCXP:
210/198.2

ORPL:
Petro et al., 1996, I.J. Chromotography A, 752: 59-66 Molded
continuous poly
(styrene-co-divinylbenzene) rod as a separation medium for the
very fast
separation of polymers Comparison of the chromatographic
properties of the
monolithic rod with columns packed with porous and non-porous
beads in
high-performance liquid chromatography of polystyrenes.

ORPL:
Petro et al., Analytical Chemistry, 1996, vol. 68: 315-321 Molded
monolithic
rod of macroporous Poly(styrene-co-divinylbenzene) as a
Separation Medium for
HPLC of Synthetic Polymers: "on-Column"
Precipitation-Redissolution
Chromatography as an Alternative to Size Exclusion Chromatography
of Styrene
Oligomers and Polymers.

DOCUMENT-IDENTIFIER: US 6258264 B1

TITLE: Non-polar media for polynucleotide separations

BSPP:

The present invention is directed to the separation of polynucleotides using a separation medium having non-polar surfaces, such as the surfaces of nonporous beads or surfaces of interstitial spaces within a molded monolith (e.g., a derivatized silica monolith), which surfaces are substantially free from contamination with multivalent cations. More specifically, the invention is directed to the chromatographic separation of both single stranded and double stranded polynucleotides by chromatography using a nonporous separation medium, where the medium is either organic or inorganic material which is coated with a polymer, or non-polar substituted polymer, and/or which has substantially all surface substrate groups substituted with a non-polar hydrocarbon or non-ionic substituted hydrocarbon.

ESPP:

These and other objects of the invention, which will become apparent from reading the following specification, have been achieved by the method of the present invention in which polynucleotides are separated using a nonporous separation medium such as beads or a molded monolith (e.g., a silica gel monolith), where the medium comprises either organic or inorganic material which is coated with a polymer, or non-polar substituted polymer, and/or which has substantially all surface substrate groups substituted with a non-polar hydrocarbon or non-ionic substituted hydrocarbon.

ESPP:

In one aspect, the invention is a method for separating a mixture of polynucleotides comprising applying a mixture of polynucleotides

having up to 1500 base pairs to a separation medium, the separation surfaces of the medium coated with a hydrocarbon or non-polar hydrocarbon substituted polymer, or having substantially all polar groups reacted with a non-polar hydrocarbon or substituted hydrocarbon group, wherein said surfaces are non-polar; and eluting the polynucleotides. The separation medium can be enclosed in a column. Examples of non-polar surfaces include the surfaces of beads such as nonporous particles and the surfaces of intersitital spaces within a monolith (e.g., a silica gel monolith), which surfaces are coated with a hydrocarbon or non-polar substituted polymer or having substantially all surface substrate groups reacted with a non-polar hydrocarbon or substituted hydrocarbon group. In the preferred embodiment, precautions are taken during the production of the medium so that it is substantially free of multivalent cation contaminants and the medium is treated, for example by an acid wash treatment and/or treatment with multivalent cation binding agent, to substantially remove any residual surface metal contaminants. The preferred separation medium is characterized by having a DNA Separation Factor (defined hereinbelow) of at least 0.05. The preferred medium is characterized by having a Mutation Separation Factor (as defined hereinbelow) of at least 0.1. In a preferred embodiment, the separation is made by Matched Ion Polynucleotide Chromatography (MIPC, as defined hereinbelow). The elution step preferably uses a mobile phase containing a counterion agent and a water-soluble organic solvent. Examples of a water-soluble organic solvent include alcohol, nitrile, dimethylformamide, tetrahydrofuran, ester, ether, and mixtures of one or more thereof, e.g., methanol, ethanol, 2-propanol, 1-propanol, tetrahydrofuran, ethyl acetate, acetonitrile. The most preferred organic solvent is acetonitrile. The counterion agent

is preferably selected from the group consisting of lower primary amine, lower secondary amine, lower tertiary amine, lower trialkylammonium salt, quaternary ammonium salt, and mixtures of one or more thereof. Non-limiting examples of counterion agents include octylammonium acetate, octyldimethylammonium acetate, decylammonium acetate, octadecylammonium acetate, pyridiniumammonium acetate, cyclohexylammonium acetate, diethylammonium acetate, propylethylammonium acetate, propyldiethylammonium acetate, butylethylammonium acetate, methylhexylammonium acetate, tetramethylammonium acetate, tetraethylammonium acetate, tetrapropylammonium acetate, tetrabutylammonium acetate, dimethyldiethylammonium acetate, triethylammonium acetate, tripropylammonium acetate, tributylammonium acetate, and mixtures of any one or more of the above. The counterion agent includes an anion, e.g., acetate, carbonate, bicarbonate, phosphate, sulfate, nitrate, propionate, formate, chloride, perchlorate, or bromide. The most preferred counterion agent is triethylammonium acetate or triethylammonium hexafluoroisopropyl alcohol.

ESPE:

In a still further aspect, the invention is a method for separating a mixture of polynucleotides comprising applying a mixture of polynucleotides having up to 1500 base pairs to a monolith having non-polar separation surfaces, and eluting the polynucleotides. The monolith can be enclosed in a column or other containment system, such as a cartridge. In a preferred embodiment, the monolith is a silica gel monolith. The non-polar separation surfaces are the surfaces of intersitital spaces within the monolith, which surfaces are coated with a hydrocarbon or non-polar substituted polymer or having substantially all surface substrate groups reacted with a non-polar hydrocarbon or substituted hydrocarbon group. An example of a suitable

monolith is one which is polyfunctionally derivatized with octadecylsilyl groups. In the preferred embodiment, precautions are taken during the production of the monolith so that it is substantially free of multivalent cation contaminants and the monolith is treated, for example by an acid wash treatment and/or treatment with multivalent cation binding agent, to substantially remove any residual surface metal contaminants. The preferred monolith is characterized by having a DNA Separation Factor of at least 0.05. The preferred monolith is characterized by having a Mutation Separation Factor of at least 0.1. In a preferred embodiment, the separation is made by Matched Ion Polynucleotide Chromatography. The elution step preferably uses a mobile phase containing a counterion agent and a water-soluble organic solvent. Examples of a suitable organic solvent include alcohol, nitrile, dimethylformamide, tetrahydrofuran, ester, ether, and mixtures of one or more thereof, e.g., methanol, ethanol, 2-propanol, 1-propanol, tetrahydrofuran, ethyl acetate, acetonitrile. The most preferred organic solvent is acetonitrile. The counterion agent is preferably selected from the group consisting of lower primary amine, lower secondary amine, lower tertiary amine, lower trialkylammonium salt, quaternary ammonium salt, and mixtures of one or more thereof. Non-limiting examples of counterion agents include octylammonium acetate, octyldimethylammonium acetate, decylammonium acetate, octadecylammonium acetate, pyridiniumammonium acetate, dodecylammonium acetate, diethylammonium acetate, propylethylammonium acetate, propyldiethylammonium acetate, butylethylammonium acetate, methylhexylammonium acetate, tetramethylammonium acetate, tetraethylammonium acetate, tetrapropylammonium acetate, tetrabutylammonium acetate, dimethyldiethylammonium acetate, triethylammonium acetate, tripropylammonium

acetate, tributylammonium acetate, and mixtures of any one or more of the above. The counterion agent includes an anion, e.g., acetate, carbonate, bicarbonate, phosphate, sulfate, nitrate, propionate, formate, chloride, perchlorate, or bromide. The most preferred counterion agent is triethylammonium acetate or triethylammonium hexafluoroisopropyl alcohol.

ESPR:

In a yet further aspect, the invention provides a monolith having non-polar separation surfaces which are substantially free from contamination with multivalent cations. The monolith can be enclosed in a column or other containment system, such as a cartridge. The non-polar separation surfaces include the surfaces of interstitial spaces within the monolith (e.g., a silica monolith), which surfaces are coated with a hydrocarbon or non-polar substituted polymer or having substantially all surface substrate groups reacted with a non-polar hydrocarbon or substituted hydrocarbon group. An example of a suitable monolith is one which is derivatized with polyfunctionally derivatized octadecylsilyl groups. In the preferred embodiment, precautions are taken during the production of the monolith so that it is substantially free of multivalent cation contaminants and the monolith is treated, for example by an acid wash treatment and/or treatment with multivalent cation binding agent, to remove any residual surface metal contaminants. The preferred monolith is characterized by having a DNA separation factor of at least 1.5. The preferred monolith is characterized by having a nucleic acid separation factor of at least 1.1.

ESPR:

In another aspect, the present invention is a method for treating the non-polar surfaces of a medium used for separating polynucleotides, such as the surfaces of beads in a MIFC column or the surfaces of interstitial spaces



in a monolith,
 in order to improve the resolution of polynucleotides, such as
 dsDNA, separated
 on said surfaces. This treatment includes contacting the surface
 with a
 solution containing a multivalent cation binding agent. In a
 preferred
 embodiment, the solution has a temperature of about 50.degree.
 C. to
 90.degree. C. An example of this treatment includes flowing a
 solution
 containing a multivalent cation binding agent through a MIPC
 column, wherein
 the solution has a temperature of about 50.degree. C. to
 90.degree. C. The
 preferred temperature is about 70.degree. C. to 80.degree. C.
 In a preferred
 embodiment, the multivalent cation binding agent is a
 coordination compound,
 examples of which include water-soluble chelating agents and
 crown ethers.
 Specific examples include acetylacetone, alizarin, aluminon,
 chloranilic acid,
 kojic acid, morin, rhodizonic acid, thionalide, thiourea,
 .alpha.-furyldioxime,
 niexime, salicylaldehyde, dimethylglyoxime, .alpha.-furyldioxime,
 cupferron,
 .alpha.-nitroso-.beta.-naphthol, nitroso-R-salt,
 diphenylthiocarbazone,
 diphenylcarbazone, eriochrome black T, PAN, SPADNS,
 glyoxal-bis(2-hydroxyanil),
 murexide, .alpha.-benzoinoxime, mandelic acid, anthranilic acid,
 ethylenediamine, glycine, triamino triethylamine, thionalide,
 triethylenetetramine, ethylenediaminetetraacetic acid (EDTA),
 metalphthalein,
 arsenic acids, .alpha., .alpha.'-bipyridine,
 4-hydroxybenzothiazole,
 6-hydroxyquinoline, 1,10-phenanthroline,
 picolinic acid,
 quinaldic acid, .alpha., .alpha.', .alpha."-terpyridyl,
 2,4,6-trihydroxy-5-fluorine, pyrocatechol, salicylic
 acid, tiron,
 2,2',4,4'-thiobis(5-fluorine), dithiol, mercaptobenzothiazole,
 rubeanic acid,
 oxalic acid, sodium diethyldithiocarbamate, and zinc
 dibenzoyldithiocarbamate. However, the most preferred chelating
 agent is EDTA.
 In this aspect of the invention, the solution preferably includes
 an organic
 solvent as exemplified by alcohol, nitrile, dimethylformamide,

tetrahydrofuran, ester, ether, and mixtures thereof. Examples of suitable solvents include methanol, ethanol, 2-propanol, 1-propanol, tetrahydrofuran, ethyl acetate, acetonitrile, and mixtures thereof. The most preferred organic solvent is acetonitrile. In one embodiment, the solution can include a counterion agent such as lower primary, secondary and tertiary amines, and lower trialkylammonium salts, or quaternary ammonium salts. More specifically, the counterion agent can be octylammonium acetate, octadecylammonium acetate, decylammonium acetate, octadecylammonium acetate, pyridiniumammonium acetate, cyclohexylammonium acetate, diethylammonium acetate, propylethylammonium acetate, propyldiethylammonium acetate, butylethylammonium acetate, methylhexylammonium acetate, tetramethylammonium acetate, tetraethylammonium acetate, tetrapropylammonium acetate, tetrabutylammonium acetate, dimethyldiethylammonium acetate, triethylammonium acetate, tripropylammonium acetate, tributylammonium acetate, tetraethylammonium acetate, tetrapropylammonium acetate, tetrabutylammonium acetate, and mixtures of any one or more of the above. The counterion agent includes an anion, e.g., acetate, carbonate, bicarbonate, phosphate, sulfate, nitrate, propionate, formate, chloride, perchlorate, and bromide. However, the most preferred counterion agent is triethylammonium acetate.

ESPE:

In yet a further aspect, the invention provides a method for storing a medium used for separating polynucleotides, e.g., the beads of a MIPC column.

monolith, in order to improve the resolution of double stranded DNA fragments separated using the medium. In the case of a MIPC column, the preferred method includes flowing a solution containing a multivalent cation binding agent through the column prior to storing the column. In a preferred embodiment, the multivalent cation binding agent is a coordination compound,

examples of which include water-soluble chelating agents and crown ethers. Specific examples include acetylacetone, alizarin, aluminon, chloranilic acid, kojic acid, morin, rhodizonic acid, thionalide, thiocurea, .alpha.-furildioxime, nioxime, salicylalldoxime, dimethylglyoxime, .alpha.-furildioxime, cupferron, .alpha.-nitroso-.alpha.-naphthol, nitroso-R-salt, diphenylthiocarbazone, diphenylcarbazone, eriochrome black T, PAN, SPADNS, glyoxal-bis(2-hydroxyanil), murexide, .alpha.-benzoinoxime, mandelic acid, anthranilic acid, ethylenediamine, glycine, triaminetriethylamine, thionalide, triethylenetetramine, EDTA, metalphthalein, arsonic acids, .alpha.,.alpha.'-bipyridine, 4-hydroxybenzothiazole, 8-hydroxyquinaldine, 8-hydroxyquinoline, 1,10-phenanthroline, picolinic acid, quinaldic acid, .alpha.,.alpha.',.alpha."-terpyridyl, 9-methyl-2,3,7-trihydroxy-6-fluorone, pyrocatechol, salicylic acid, tiron, 4-chloro-1,2-dimercaptobenzene, dithiol, mercaptobenzothiazole, rubeanic acid, oxalic acid, sodium diethyldithiocarbamate, and zinc dibenzylidithiocarbamate. However, the most preferred chelating agent is EDTA. In this aspect of the invention, the solution preferably includes an organic solvent as exemplified by alcohols, nitriles, dimethylformamide, tetrahydrofuran, esters, and ethers. The most preferred organic solvent is acetonitrile. The solution can also include a counterion agent such as lower primary, secondary and tertiary amines, and lower trialkylammonium salts, or quaternary ammonium salts. More specifically, the counterion agent can be octylammonium acetate, tetradecylammonium acetate, decylammonium acetate, octadecylammonium acetate, dodecylammonium acetate, cyclohexylammonium acetate, diethylammonium acetate, propyldiethylammonium acetate, butyldiethylammonium acetate, methylnonylammonium acetate, tetramethylammonium acetate, tetraethylammonium acetate, tetrapropylammonium acetate, tetrabutylammonium acetate, dimethyldiethylammonium acetate, triethylammonium

acetate, tripropylammonium acetate, tributylammonium acetate, tetraethylammonium acetate, tetrapropylammonium acetate, tetrabutylammonium acetate, and mixtures of any one or more of the above. The counterion agent includes an anion, e.g., acetate, carbonate, bicarbonate, phosphate, sulfate, nitrate, propionate, formate, chloride, perchlorate, and bromide. However, the most preferred counterion agent is triethylammonium acetate.

DEPR:

The medium can be enclosed in a column. In one embodiment, the non-polar surfaces comprise the surfaces of beads. In an alternative embodiment, the surfaces comprise the surfaces of interstitial spaces in a molded monolith.

For purposes of simplifying the description of the invention and not by way of limitation, the separation of polynucleotides using nonporous beads, and the preparation of such beads, will be primarily described herein, it being

understood that other separation surfaces, such as the interstitial surfaces of monoliths, are intended to be included within the scope of this invention.

Monoliths such as derivatized silica gel rods contain separation media which have been formed inside a column as a unitary structure having through pores or interstitial spaces which allow eluting solvent and analyte to pass through and which provide the non-polar separation surface.

DEPR:

In another embodiment of the present invention, the separation medium can be in the form of a monolith such as a rod-like monolithic column. The monolithic column can be polymerized or formed as a single unit inside of a tube. The through pore or interstitial spaces provide for the passage of eluting solvent and analyte materials. The separation is performed on the stationary surface. The surface can be porous, but is preferably nonporous. The form

and function of the separations are identical to columns packed with beads. As with beads, the pores contained in the rod must be compatible with DNA and not trap the material. Also, the rod must not contain contamination that will trap DNA.

DEPE:

In one embodiment of the present invention, the separation medium is continuous

monolithic silica gel. A molded monolith can be prepared by polymerization within the confines of a chromatographic column (e.g., to form a rod) or other containment system. A monolith is preferably obtained by the hydrolysis and polycondensation of alkoxysilanes. A preferred monolith is derivatized in order to produce non-polar interstitial surfaces. Chemical modification of silica monoliths with octadecyl, methyl or other ligands can be carried out.

An example of a preferred derivatized monolith is one which is polyfunctionally derivatized with octadecylsilyl groups. The preparation of derivatized silica monoliths is by conventional methods well known in the art as described in

Example 15 and in the following references which are hereby incorporated in their entirety herein: Nakanishi, et al., J. Sol-Gel Sci. Technol. 8:547 (1997); Nakanishi, et al., Bull. Chem. Soc. Jpn. 67:1327 (1994); Cabrera, et al., Trends Analytical Chem. 17:50 (1998); Jinno, et al., Chromatographia 27:198 (1989).

DEPE:

The non-polar, derivatized silica monolith column is washed by flowing acetonitrile through the column at a flow rate of 2 mL per minute for 10 minutes followed by flowing methanol through the column at 2 mL per minute for 10 minutes. The non-polar monolith column is washed further by flowing a mixture containing 100 mL of tetrahydrofuran and 100 mL of concentrated

hydrochloric acid through the column at 10 mL per minute for 20 minutes.

Following this acid treatment, the monolith column is washed by flowing tetrahydrofuran/water (1:1) through the column at 2 mL per minute until neutral (pH 7).

DEPC:

Preparation of a Silica Monolith

CCOR:

210/198.2

OEPL:

Nakanishi et al. Double Pore Silica Gel Monolith Applied to Liquid Chromatography, J. Sol-Gel Science & Technology, vol. 8, pp. 547-552, 1997.

OEPL:

Petro et al, Molded Monolithid Rod of Macrophrous Poly(Styrene-CO-Divinylbenzene) as a Separation Medium for PHLC of Synthetic Polymers . . . , Analytical Chemistry, 68: 315-321 (1996).

DOCUMENT-IDENTIFIER: US 6245227 B1

TITLE: Integrated monolithic microfabricated electrospray and liquid chromatography system and method

TTL:

Integrated monolithic microfabricated electrospray and liquid chromatography system and method

ABPL:

An electrospray device, a liquid chromatography device and an electrospray-liquid chromatography system are disclosed. The electrospray device comprises a substrate defining a channel between an entrance orifice on an injection surface and an exit orifice on an ejection surface, a nozzle defined by a portion recessed from the ejection surface surrounding the exit orifice, and an electrode for application of an electric potential to the substrate to optimize and generate an electrospray; and, optionally, additional electrode(s) to further modify the electrospray. The liquid chromatography device comprises a separation substrate defining an introduction channel between an entrance orifice and a reservoir and a separation channel between the reservoir and an exit orifice, the separation channel being populated with separation posts perpendicular to the fluid flow; a cover substrate bonded to the separation substrate to enclose the reservoir and the separation channel adjacent the cover substrate; and, optionally, electrode(s) for application of an electric potential to the fluid. The exit orifice of the chromatography device may be homogeneously interfaced with the entrance orifice of the electrospray device to form an integrated single system. An array of multiple systems may be fabricated in a single monolithic chip for rapid sequential fluid processing and generation of electrospray for

subsequent analysis, such as by positioning the exit orifices of the electrospray devices near the sampling orifice of a mass spectrometer.

PCPR:

This application is related to copending U.S. application Ser. No.

09/156,507, entitled INTEGRATED MONOLITHIC MICROFABRICATED ELECTROSPRAY AND LIQUID CHROMATOGRAPHY SYSTEM AND METHOD, filed Sep. 17, 1998.

BSFR:

The present invention relates generally to an integrated miniaturized chemical analysis system fabricated using microelectromechanical systems (MEMS) technology. In particular, the present invention relates to an integrated monolithic microfabricated electrospray and liquid chromatography device. This achieves a significant advantage in terms of high-throughput analysis by mass spectrometry, as used, for example, in drug discovery, in comparison to a conventional system.

BSFR:

In all of the above-described devices, edge-spraying from a monolithic chip is a poorly controlled process due to the inability to rigorously and repeatably determine the physical form of the chip's edge. In another embodiment of edge-spraying, ejection nozzles, such as small segments of drawn capillaries, are separately and individually attached to the chip's edge. This process is inherently cost-inefficient and unreliable, imposes space constraints in chip design, and is therefore undesirable for manufacturing.

The fabrication of the electrospray device 100 will now be explained with reference to FIGS. 9A-20B. The electrospray device 100 is preferably fabricated as a monolithic silicon integrated circuit utilizing established, well-controlled thin-film silicon processing techniques such as

thermal oxidation, photolithography, reactive-ion etching (RIE), ion implantation, and metal deposition. Fabrication using such silicon processing techniques facilitates massively parallel processing of similar devices, is time- and cost-efficient, allows for tighter control of critical dimensions, is easily reproducible, and results in a wholly integral device, thereby eliminating any assembly requirements. Further, the fabrication sequence may be easily extended to create physical aspects or features on the injection surface and/or ejection surface of the electrospray device to facilitate interfacing and connection to a fluid delivery system or to facilitate integration with a fluid delivery sub-system to create a single integrated system.

DEPR:

The above described fabrication sequence for the electrospray device 100 can be easily adapted to and is applicable for the simultaneous fabrication of a single monolithic system comprising multiple electrospray devices including multiple channels and/or multiple ejection nozzles embodied in a single monolithic substrate. Further, the processing steps may be modified to fabricate similar or different electrospray devices merely by, for example, modifying the layout design and/or by changing the polarity of the photomask and utilizing negative-working photoresist rather than utilizing positive-working photoresist.

DEPR:

The upstream fluid delivery device 310 may be a monolithic integrated circuit chip or a fluid channel through which a fluid sample can pass directly or indirectly to the entrance orifice of the electrospray device 100. The upstream fluid delivery device 310 may be a silicon microchip-based liquid separation device capable of, for example, capillary electrophoresis, capillary

electrochromatography, affinity chromatography, liquid chromatography (LC) or any other condensed-phase separation methods. Further, the upstream fluid delivery device 318 may be a silicon, glass, plastic and/or polymer based device such that the electrospray device 100 may be chip-to-chip or wafer-to-wafer bonded thereto by any suitable method. An example of a monolithic liquid chromatography device for utilization in, for example, the single integrated system 316, is described below.

DEPR:

The silicon-based liquid chromatography device 400 reduces the size of a typical liquid chromatography device by nearly two orders of magnitude. The dimensional scaling may provide the advantage of significantly reducing the mass of the analyte and/or the volume of the fluid sample required for accurate analysis. Further, by reducing a macroscopic separation column and its packaging materials to a monolithic device, the liquid chromatography device 400 can be a component of an on-chip integrated system.

DEPR:

Referring now to FIGS. 30-35, although the liquid chromatography device has been described as comprising a single reservoir and a single separation channel, the monolithic liquid chromatography device may be easily adapted and modified to comprise multiples of the liquid chromatography device and/or multiple entrance orifices, exit orifices, reservoirs and/or separation channels. In each of the variations, any or all of the reservoir(s), separation channel(s), and separation orifice(s) may have different dimensions and/or shapes.

DEPR:

The fabrication of the liquid chromatography device of the present invention will now be explained with reference to FIGS. 36A-46C. The

liquid chromatography device is preferably fabricated as a monolithic silicon micro device utilizing established, well-controlled thin-film silicon processing techniques such as thermal oxidation, photolithography, reactive-ion etching (RIE), ion implantation, and metal deposition. Fabrication using such silicon processing techniques facilitates massively parallel processing of similar devices, is time- and cost-efficient, allows for tighter control of critical dimensions, is easily reproducible, and results in a wholly integral device, thereby eliminating any assembly requirements. Manipulation of separate components and/or sub-assemblies to build an liquid chromatography device with high reliability and yield is not desirable and may not be possible at the micrometer dimensions required for efficient separation.

DEPR:

The above described fabrication sequence for the liquid chromatography device may be easily adapted to and is applicable for the simultaneous fabrication of a monolithic system comprising multiple liquid chromatography devices including multiple reservoirs and/or multiple separation channels as described above embodied in a single monolithic substrate.

CCDR:

210/198.2

DOCUMENT-IDENTIFIER: US 6238565 B1

TITLE: Monolithic matrix for separating bio-organic molecules

TTL:

Monolithic matrix for separating bio-organic molecules

ABFL:

The present invention provides monolithic polymer matrices for the separation of bio-organic molecules by liquid chromatography. In one embodiment, the matrix is formed from a polymerization mixture including (i) a hydrophobic monomer, (ii) a crosslinking agent, and (iii) a porogenic solvent or mixture of porogenic components. The monolithic matrices of the invention are particularly useful for resolving polynucleotides (e.g., DNA and/or RNA) in samples by way of reversed-phase ion-pairing chromatography.

BBFL:

The present invention relates to the separation of bio-organic molecules. In particular, the invention provides a monolithic polymer matrix for the separation of polynucleotides by reversed-phase ion-pairing chromatography.

ESFL:

The above theory relating to beds of packed particles is not particularly useful for predicting the behavior of macromolecules in continuous monolithic beds, where mass transport may be a combination of diffusive and convective processes.

EPFL:

The present invention, relating to monolithic beds for separating mixtures containing polynucleotides, is based in part on the discovery that monoliths provide reduced pressure drops corresponding to the use of large particle stationary phases while maintaining the separation resolution of columns packed

with small spherical particles. More particularly, it has been discovered that reversed-phase monolithic matrices can provide an improved method for the high speed separation of DNA molecules and that such separations can be performed with high resolution at greatly reduced operating pressures compared to previously available methods. This surprising finding now permits the high-resolution separation of polynucleotides under conditions not possible with preexisting technology. Moreover, the monolithic columns of the present invention can be constructed with stationary phase geometries significantly different than those available with packed beds. The effects of such novel geometries on the separation of macromolecules have not been predicted so far by current chromatographic theory.

ESPR:

The monolithic columns of the present invention provide all of the advantages of the previous best technology for polynucleotide separations (i.e., packed beds of alkylated nonporous polymer beads), without the need to tediously prepare beads and pack them into efficient columns. The columns produced by the current invention are easily prepared using simple processes and once prepared, cannot fail through shifting within a packed bed because there are no individual beads to shift position.

ESPR:

In addition to the improved ease of manufacturing of the new columns and lack of bead shifting, the monolithic columns described herein provide a surprising advantage in that they are capable of providing at least 50% better resolution than that expected for a column of packed spheres when normalized for operating pressures.

ESPR:

One aspect of the present invention provides a method for

resolving a mixture containing at least one polynucleotide (e.g., DNA and/or RNA). According to the method, the mixture is passed through a monolithic polymer matrix held in a stationary fashion by a support. The mixture is separated by ion-pair reverse-phase chromatography.

BSPP:

In one embodiment, the monolithic polymer matrix has hydrophobic surface groups. The monolithic polymer matrix can be comprised, at least in part, of a polymer selected from the group consisting of polymethacrylates and polystyrenes. In one embodiment, the polymer is a polymethacrylate.

PSPP:

According to one embodiment, the monolithic polymer matrix is formed from a polymerization mixture including a hydrophobic monomer, a crosslinking agent, and a porogen. The porogen may be (i) a porogenic solvent, (ii) a mixture of porogenic solvents, or (iii) one or more porogenic solvents containing at least one polymeric additive that contributes to pore formation. The hydrophobic monomer can be an alkyl methacrylate.

ESPP:

In one embodiment, the method further includes the step of passing an eluant containing an ion-pairing agent through the monolithic matrix. The separation is carried out under the driving force of a reasonable pressure (e.g., less than about 5,000 psi).

ASPP:

An additional embodiment of the invention provides a plate in which the monolithic polymer matrix may fill a channel formed in the plate, or it may take the form of a thin film on the plate.

BSPP:

Another aspect of the invention provides a chromatographic

apparatus comprising
a support holding a macroporous monolithic methacrylate-based
polymer matrix.
In one embodiment, the polymer matrix has a hydrophobic surface
capable of
interacting with hydrophobic groups of an ion-pairing agent for
resolving
polynucleotides by reverse-phase ion-pair chromatography. The
polymer matrix
may have a relatively high void fraction (e.g., greater than
about 0.6).

ESPE:

In another embodiment, the kit includes: (i) a monolithic polymer
matrix having
hydrophobic surface groups held in a stationary fashion by a
support; and (ii)
an ion-pairing agent capable of interacting with negatively
charged phosphate
groups of the polynucleotides and also with the hydrophobic
surface groups of
the monomer.

DEFE:

FIG. 1 is a chromatogram showing the separation of
oligothymidylic acids
between 12 to 18 units in length on a C6 monolithic column
constructed in
accordance with an embodiment of the present invention.

DEFE:

FIG. 2A is a chromatogram showing the separation of
double-stranded DNA
fragments on a C12 monolithic column constructed in accordance
with an
embodiment of the present invention.

DEFE:

FIG. 3A is a chromatogram showing the separation of homoduplex
DNA fragments
of 4 base pairs in length on a porous monolithic C12 column
constructed in
accordance with an embodiment of the present invention.

DEFE:

FIG. 3B is a chromatogram showing the separation of a mixture
containing the
same type of homoduplex DNA fragments of FIG. 3A as well as a
variant sequence
containing a single base pair substitution on a porous monolithic

C12 column
constructed in accordance with an embodiment of the present
invention.

DEPP:

The present invention provides a monolithic bed, and method of
making and using
the same, for resolving bio-organic molecules.

DEPP:

Any suitable crosslinking agents known to those skilled in art
may be employed
in forming the monolithic matrix of the invention. Preferred
crosslinking
monomers contain at least two carbon-carbon double bonds capable
of
polymerization in the presence of an initiator. Exemplary
crosslinking
monomers include divinyl benzene, butadiene, trimethylolpropane
trimethacrylate
(TRIM), etc.

DEPI:

For TRIM-based polymerization mixtures, highly permeable
monoliths will result,
for example, from the use of pure isooctane as a porogenic
solvent. Lower
permeability will result, for example, from addition of 5-20% of
other
solvents, such as 2-octanone, toluene, and/or ethyl propionate,
to the
isooctane. Permeability may also be reduced by increasing the
ratio of
hydrophobic monomer to crosslinker, and by decreasing the
proportion of
porogenic solvent in the mixture. Additionally, the choice of
initiator may be
used as a means to control the pore distribution.

DEPP:

In an exemplary embodiment, the monolithic matrix is prepared
from a
polymerization mixture of crosslinker, alkyl methacrylate, or alkane
porogenic
solvent or mixture of porogenic components, and a free radical
initiator. For
example, a polymerization mixture was prepared using
trimethylolpropane
trimethacrylate, an alkyl methacrylate, isooctane, and
azobisisobutyronitrile.

The mixture was poured into an empty HPLC column tube, capped, and polymerized at 65-70.degree. C. for 8-24 hours. The polymerized matrix was then equipped with endfittings and flushed with solvent to remove unreacted components.

DEPR:

A typical monolithic column constructed in accordance with the present invention will have pores in the less than 5,000 nm range, down to about 10 nm. The size of larger pores can be determined using very large DNA and/or microscopic exam. For example, the columns of the invention have been used to separate DNA products of 2,072, 2,647, and 3,147 base pairs in length. The largest of these would correspond to a molecular size of at least 1 micron. Accordingly, the monolithic column used to separate this DNA would be expected to include pores having a diameter of at least 1 micron. This is in agreement with permeability calculations that indicate an apparent particle diameter of 6 microns, or a pore size of about 2 microns. Microscopic exam revealed that a typical matrix of the invention includes small globules of about 1-2 microns in diameter that are fused into continuous structures with pores in the 1-3 micron size range. This is consistent with the estimated pore size based on permeability and DNA fragment chromatography.

DEPR:

Once prepared, the monolithic matrix of the invention is useful for resolving bio-organic molecules, for example, by gradient or isocratic liquid chromatography. In an exemplary use, a column of the invention, prepared as described above, is employed to carry out a separation of polynucleotides contained in a sample. In one embodiment, water and acetonitrile, both containing an ion-pairing agent, are used in gradient liquid chromatography to elute polynucleotides injected into the column.

DEPR:

A variety of support structures may be used with the monolithic matrix of the invention. For example, one embodiment contemplates the use of an elongated column tube. In another embodiment, the monolithic matrix is held in the lumen of a capillary tube. In these embodiments, the matrix extends across the entire cross-sectional area of the tube. The tube may be of any desired cross-section, e.g., circular, polygonal, or other shape. The monolithic matrix may be polymerized in situ within the tube, or it may be formed outside of the tube and then inserted by any suitable means.

DEPR:

In alternative embodiments, the monolithic matrix is (i) held in a channel or groove on a plate, or (ii) applied as a thin film across a plate.

While the monoliths of the invention may be provided on a substrate or supporting plate of virtually any size and composed of any suitable material, the embodiment of the present invention contemplates using a supporting plate of a standard size, which can be constructed using conventional materials and means. Thus, in this embodiment, an injection molded rectangular plastic plate, the length and width of which conform to the commonly used standard of 8.810".times.6.365" (127.76 mm and 65.47 mm), is preferred. Similarly, while a single plate may support any reasonable number of sites, constructions corresponding to the commonly used "96-well" microtiter plate format are preferred. Thus, one preferred

construction includes an 8.times.12 array of sites situated on a standard sized support plate. Each site may support one or more separate monoliths. The position and spacing of the sites on the support plate may be standard, as well (e.g., spaced about 9 mm center-to-center). Utilization of standard outside dimensions for the plate frame, as well as standard spacing for

the sites on the plate, facilitates use of the plates with existing equipment, such as automated dispensers or optical readers, if desired. It should be appreciated that such an apparatus is capable of handling multiple simultaneous and parallel separations.

DEPR:

In a further embodiment, the monolithic matrix fills one or more channels formed on a microchip. For example, a plurality of channels can be formed on a glass microchip substrate using standard microfabrication techniques, e.g., photolithographic procedures and chemical wet etching. Optionally, a cover plate can be directly bonded to the substrate over the channels. A plurality of separations can be conducted on multiple samples in a substantially parallel fashion upon a single substrate chip.

DEPR:

Advantageously, the columns of this invention are not limited to the fixed surface areas, pressure drops and interstitial diffusional distances of a packed bed. Because the void fraction can be much higher in the monolithic columns taught herein, all of these factors can be manipulated for improved performance for a given application. For example, columns can be produced with the same void fraction and widely different pressure drops, or columns could be produced with the same pressure drop but different interstitial distances. In a particular example, some columns produced in this invention performing well for DNA separations have void fractions of about 0.7. A void fraction of 0.7 cannot be produced for nonporous packings by packing discrete particles.

DEPR:

FIG. 1 is a chromatogram showing the separation of single-stranded oligothymidylic acids on a C6 monolithic column constructed in

the manner just described. Specifically, the sample was 10 microliters of oligothymidylic adds between 12 and 18 units in length. At a flow rate of 2 ml/minute, a gradient of 9-12% acetonitrile in 100 mM aqueous triethylammonium acetate, 10 mM disodium EDTA, pH 7.0 was used to elute the oligonucleotides. Detection was performed using UV absorbance at 254 nm. An extremely low HPLC system pressure of only 150 psi was observed during the separation.

DEPR:

FIG. 2A is a chromatogram showing the separation of double-stranded DNA fragments on a monolithic C12 column constructed in the manner just described. Specifically, the sample was 3 microliters of pUC18 DNA digested by the MSP I restriction enzyme. At a flow rate of 1.0 ml/minute, a 10 minute gradient of 35-60% acetonitrile containing 2 mM tetrapropylammonium bromide in 20 mM aqueous tetrapropylammonium bromide, 2 mM disodium EDTA, pH 7.0 was used to elute the DNA fragments. Detection was performed using UV absorbance at 254 nm. At 0.5 ml/min an HPLC system pressure of 50 psi was observed. Resolution was calculated for the peaks indicated in FIG. 2A by arrows.

DEPR:

The chromatographic data from the above experiments are summarized in Table 1. The resolution of the marked peaks was found to be 4.04 for the monolithic

column of FIG. 2A and 5.83 for the column packed with nonporous spheres in FIG.

2B. Using standard permeability calculations (Introduction to Liquid Chromatography, 2nd edition, L. R. Snyder and J. J. Kirkland, Wiley, and Sons, New York, 1979, p. 37), it can be calculated that the

column of FIG. 2A has an operating pressure equivalent to that of a column

packed with 6.21

micron diameter spheres whereas that in FIG. 2B would be estimated as 1.10

micron. Based on these different effective particle sizes, the

column of FIG.

2A has a resolution 58% greater than expected. A critical figure of merit for chromatographic separations is the efficiency or resolution per unit pressure.

It can be seen from the data of Table 1 that the monolithic column offers an order of magnitude increase in resolution per unit pressure compared to the column packed with nonporous spheres.

DEPR:

The present invention also makes possible the separation of partially denatured double-stranded polynucleotides. As shown in FIGS. 3A and 3B, a porous

monolithic column was used to separate DNA fragments 304 base pairs in length.

In FIG. 3A, the sample contained a single type of DNA, whereas in FIG. 3B the

sample contained a mixture of the type seen in FIG. 3A and a variant sequence

containing a single base pair substitution. At the selected temperature, the

sample of FIG. 3B shows a second peak that represents partially denatured DNA

containing the single base pair substitution.

DEPR:

Specifically, with regard to FIG. 3A, homoduplex DNA with sequence 304 base

pairs in length was separated on a monolithic C12 column, prepared as described

above. In carrying out the separation, the conditions as set out in Table 2,

below, were observed:

DEPR:

For comparative purposes, columns were constructed in accordance with Examples

III and VI of the '310 patent. Briefly, these examples describe monolithic

columns which are prepared in situ. After the polymerizations of these

compositions, end fittings were attached and the columns flushed with methanol,

as described in the '310 patent. The composition produced by Example III of

the '310 patent was translucent in appearance and had a very high operating

pressure (greater than 2,000 psi at 0.25 ml/min with methanol), precluding its use in practical HPLC of DNA. The composition produced by Example VI of the '310 patent was white and had a relatively low operating pressure. However, this latter column did not provide useful separations of DNA restriction fragments under reversed-phase ion-pairing conditions. No typical pattern of peaks was observed when a separation was attempted under conditions that would provide a useful separation using the matrices of the present invention. These findings are in accord with previous suggestions in the literature that columns with unmodified polystyrene/divinylbenzene structures are not desirable for DNA separations.

DETL:

TABLE 1 Column with Nonporous Column with C12 Spherical Packing Monolithic
 Polymer Retention Time Peak 1 (min.) 6.80 9.36 Area Peak 1 (.mu.V*sec) 80678.00 84592.00 Height Peak 1 (.mu.V) 11370.00 7438.00
 Retention Time Peak 2 (min.) 7.52 10.10 Area Peak 2 (.mu.V*sec) 103115.00 103644.00
 Height Peak 2 (.mu.V) 13348.00 9779.00 Resolution 5.83 4.04 Column Pressure (psi) 1340.00
 100.00 Apparent Particle Size (.mu.m) 1.20 6.20 Expected Resolution 5.83 3.57
 Excess Resolution = 0.00 58.00 Resolution/1,000 psi 4.35 40.42

CLPR:

1. The method of claim 1, wherein said separating comprises passing an eluent containing an ion-pairing agent through said monolithic matrix, said ion-pairing agent being an alkylammonium salt.

III:

2. The method of claim 1, further comprising the step of passing an eluent containing an ion-pairing agent through said monolithic matrix, said ion-pairing agent being capable of interacting with negatively charged phosphate groups of said at least one polynucleotide and also

with said
hydrophobic surface of said monolithic polymer matrix.

CLPF:

9. The method of claim 8, wherein said monolithic polymer matrix is held in a channel formed in said plate.

CLPR:

10. The method of claim 8, wherein said monolithic polymer matrix takes the form of a thin film on said plate.

CLPV:

applying said mixture to a porous monolithic polymer matrix held in a stationary fashion by a support,

CLFV:

wherein said monolithic polymer matrix is formed from polymerization of a monomer, or combination of monomers, selected from C.sub.3 to C.sub.30 alkyl methacrylates, in the presence of a crosslinking agent and a porogenic solvent, and

CLFV:

separating said mixture by ion-pair reverse-phase chromatography, by passing an eluant containing an ion-pairing agent through the monolithic matrix.

CCXR:

210/198.2

CEPL:

Article by Viklund et al, entitled "'Molded' Macroporous Poly (glycidylmethacrylate-co-trimethylolpropane trimethacrylate), Materials with Fine Controlled Porous Properties: Preparation of Monoliths Using Free-Initiated Polymerization" published in Chem Mater. In 1997, 9, 361-371.

CEPL:

Article by Viklund et al., entitled "Monolithic, Molded , Porous Materials with High Flow Characteristics for Separations, Catalysis, or Solid-Phase

Chemistry: Control of Porous Properties during Polymerization,"
published in
Chem. Mater in 1996, in vol. 8, pp. 744-750.

DOCUMENT-IDENTIFIER: US 6218153 B1
TITLE: Target DNA amplification by MIPC and PCR

DEPR:

MIPC uses unique non-polar separation media which comprises organic polymers, silica media having a non-polar surface comprising coated or covalently bound organic polymers or covalently bound alkyl and/or aryl groups, and continuous non-polar separation media, i.e., monolith or rod columns such as non-polar silica gel or organic polymer. The separation media used in MIPC can be porous or non-porous. A detailed description of the MIPC separation process, MIPC separation media, and MIPC systems is found in U.S. Pat. No. 5,772,889 (1998) to Gjerde and in co-pending U.S. patent applications Ser. Nos. 09/058,580 filed Mar. 10, 1998; (abandoned); 09/058,337 filed Mar. 10, 1998; (abandoned); 09/061,040 filed May 13, 1998 (now U.S. Pat. No. 5,947,742); 09/080,547 filed May 18, 1998 (now U.S. Pat. No. 6,017,457); and in the U.S. patent application Ser. No. 09/169,440 filed Oct. 9, 1998. MIPC systems and separation media are commercially available (Transgenomic, Inc. San Jose, Calif.). The entire MIPC analysis can be automated by means of a desk top computer and a sample auto-injector. Analytical data for each sample can be analyzed in real time, or collected and stored in a computer memory device for analysis at a later time.

CLASS:

210/198.2

DOCUMENT-IDENTIFIER: US 6210385 B1

TITLE: Modifying double stranded DNA to enhance separations by matched ion polynucleotide chromatography

BSPR:

In one aspect, the invention provides a method for enhancing the detection of a polynucleotide separated by Matched Ion Polynucleotide Chromatography which includes (a) covalently attaching a chemical tag to the polynucleotide to form a tagged polynucleotide, (b) applying the tagged polynucleotide to a separation medium having a non-polar separation surface, the medium characterized by having a DNA Separation Factor of at least 0.05, (c) eluting the tagged polynucleotide from the surface with a mobile phase containing a counterion agent and an organic solvent, and (d) detecting the tagged polynucleotide. The chemical tag is preferably a fluorescent group, a chemical which absorbs at a wavelength different from the polynucleotide itself, or, less preferably, a group containing a radioactive atom (e.g., P-32, tritium, or S-35).

Non-limiting examples of fluorescent groups which absorb at a wavelength

different from the polynucleotide itself include

6-carboxyfluorescein,

3',7'-dimethoxy-4',5'-dichloro-6-carboxyfluorescein,

N,N,N',N'-tetramethyl-6-carboxyrhodamine, 6-carboxy-X-rhodamine,

fluorescein,

Rhodamine, BODIPY-TR-X, Cascade Blue, Alexa 350, and porphyrin

derivatives

(e.g., texaphyrin). Non-limiting examples of fluorescent groups include

6-carboxyfluorescein,

3',7'-dimethoxy-4',5'-dichloro-6-carboxyfluorescein,

N,N,N',N'-tetramethyl-6-carboxyrhodamine, 6-carboxy-X-rhodamine,

fluorescein,

Rhodamine, BODIPY-TR-X, Cascade Blue, and Alexa 350. The

preferred separation

medium is characterized by having a Mutation Separation Factor of at least 0.1.

The preferred medium is substantially free from contamination

with multivalent cations. In one embodiment, the separation medium consists of polymer beads having an average diameter of 0.5 to 100 microns and having a surface composition essentially completely substituted with a moiety selected from the group consisting of unsubstituted, methyl, ethyl, hydrocarbon, and hydrocarbon polymer, wherein the hydrocarbon polymer optionally has from 23 to 1,000,000 carbons, wherein the hydrocarbon includes alkyl and alkyl substituted aryl groups having from 23 to 1,000,000 carbons, the alkyl groups including straight chained, branch chained, cyclic, saturated, unsaturated nonionic functional groups of various types including aldehyde, ketone, ester, ether, alkyl groups, and the like, and the aryl groups including as monocyclic, bicyclic, and tricyclic aromatic hydrocarbon groups including phenyl, naphthyl, and the like.

In another embodiment, the separation medium consists of beads having an average diameter of 0.5 to 100 microns, the beads comprising nonporous particles coated with a hydrocarbon or non-polar hydrocarbon substituted polymer, wherein the hydrocarbon has optionally from 1 to 1,000,000 carbons, wherein the hydrocarbon polymer has optionally from 1 to 1,000,000 carbons, or particles having substantially all polar groups reacted with a non-polar hydrocarbon or substituted hydrocarbon group, wherein the particles are a member selected from the group consisting of silica, silica carbide, silica nitride, titanium oxide, aluminum oxide, zirconium oxide, carbon, insoluble polysaccharide, and diatomaceous earth. In other embodiments, the separation medium is a polymeric monolith or a derivatized silica gel monolith. The tagged polynucleotide can be a PCR amplification product obtained by providing a PCR primer having a covalently bound tag during a PCR amplification wherein the tag is incorporated into the PCR amplification product.

DEPP:

The medium can be enclosed in a column. In one embodiment, the non-polar surfaces comprise the surfaces of separation beads, such as polymeric beads or derivatized particles (e.g., silica particles). In an alternative embodiment, the surfaces comprise the surfaces of interstitial spaces in a molded monolith

such as a polymeric monolith or a silica gel monolith. For purposes of simplifying the description of the invention and not by way of limitation, the separation of polynucleotides using nonporous beads, and the preparation of such beads, will be primarily described herein, it being understood that other separation surfaces, such as the surfaces within interstitial spaces of monoliths, are intended to be included within the scope of this invention.

Examples of suitable monoliths include polymeric rods and derivatized silica gel rods which have been formed inside a column as a unitary structure having through pores or interstitial spaces which allow eluting solvent and analyte to pass through and which provide the non-polar separation surface.

DEPP:

In another embodiment of the present invention, the separation medium can be in the form of a polymeric monolith such as a rod-like monolithic column. The

monolithic column is polymerized or formed as a single unit inside of a tube.

The through pore or interstitial spaces provide for the passage of eluting solvent and analyte materials. The separation is performed on the stationary surface. The surface can be porous, but is preferably nonporous.

The function of the separations are identical to columns packed with beads. As with beads, the pores contained in the rod must be compatible with DNA and not trap the material. Also, the rod must not contain contamination that will trap DNA.

DEPR:

The molded polymeric rod of the present invention is prepared by bulk free

radical polymerization within the confines of a chromatographic column. The

base polymer of the rod can be produced from a variety of polymerizable

monomers. For example, the monolithic rod can be made from polymers, including

mono- and di-vinyl substituted aromatic compounds such as styrene, substituted

styrenes, alpha-substituted styrenes and divinylbenzene; acrylates and

methacrylates; polyolefins such as polypropylene and polyethylene; polyesters;

polyurethanes; polyamides; polycarbonates; and substituted polymers including

fluorosubstituted ethylenes commonly known under the trademark TEFLON. The

base polymer can also be mixtures of polymers, non-limiting examples of which

include poly(glycidyl methacrylate-co-ethylene dimethacrylate), poly(styrene-divinylbenzene) and

poly(ethylvinylbenzene-divinylbenzene). The

rod can be unsubstituted or substituted with a substituent such as a

hydrocarbon alkyl or an aryl group. The alkyl group optionally has 1 to

1,000,000 carbons inclusive in a straight or branched chain, and includes

straight chained, branch chained, cyclic, saturated, unsaturated nonionic

functional groups of various types including aldehyde, ketone, ester, ether,

alkyl groups, and the like, and the aryl groups includes as monocyclic,

bicyclic, and tricyclic aromatic hydrocarbon groups including phenyl, naphthyl,

and the like. In a preferred embodiment, the alkyl group has 1-24 carbons. In

a more preferred embodiment, the alkyl group has 1-8 carbons. The rod can also

contain hydroxy, cyano, nitro groups, or one or more which are considered

to be non-polar, reverse phase functional groups. Methods for hydrocarbon

substitution are conventional and well-known in the art and are not an aspect

of this invention. The preparation of polymeric monoliths is by

conventional methods well known in the art as described in the following references: Wang et al. (J. Chromatog. A 699:230 (1994)), Petro et al. (Ana. Chem. 68:315 (1996)), and the following U.S. Pat. Nos. 5,334,310; 5,453,185; 5,522,994 (to Frechet). Monolith or rod columns are commercially available from Merck & Co (Darmstadt, Germany).

DEPE:

In another embodiment of the present invention, the separation medium is continuous monolithic silica gel. A molded monolith can be prepared by polymerization within the confines of a chromatographic column (e.g., to form a rod) or other containment system. A monolith is preferably obtained by the hydrolysis and polycondensation of alkoxysilanes. A preferred monolith is derivatized in order to produce non-polar interstitial surfaces. Chemical modification of silica monoliths with octadecyl, methyl or other ligands can be carried out. An example of a preferred derivatized monolith is one which is polyfunctionally derivatized with octadecylsilyl groups. The preparation of derivatized silica monoliths is by conventional methods well known in the art as described in the following references which are hereby incorporated in their entirety herein: Nakanishi, et al., J. Sol-Gel Sci. Technol. 6:347 (1997); Nakanishi, et al., Bull. Chem. Soc. Jpn. 67:1327 (1994); Cabrera, et al., Trends Analytical Chem. 17:50 (1998); Jinno, et al., Chromatographia 27:288 (1990).

DEPE:

to detecting said tagged polynucleotide, wherein said medium is characterized by having a DNA Separation Factor of at least 0.5, wherein said medium comprises a polymeric monolith.

CLPV:

d) detecting said tagged polynucleotide, wherein said medium is characterized by having a DNA Separation Factor of at least 0.5, wherein said medium comprises a derivatized silica gel monolith.

CCXR:

210/198.2

DOCUMENT-IDENTIFIER: US 6210570 B1
TITLE: Monolithic silica column

TTL:
Monolithic silica column

ABPL:

The present invention relates to capillary columns including a monolith and a method for preparing a capillary column including a monolith. The monolith can be prepared by a sol gel method, and in the transformation from hydrosol to hydrogel, the monolith undergoes essentially no syneresis or volume shrinkage. Thus, deleterious effects of syneresis are avoided, such as the formation of channels having large dimensions that provide a pathway of least resistance for a mobile phase to effectively bypass portions of a stationary phase. The method for preparing a column having a monolith that undergoes essentially no syneresis involves a hydrogel solution that has a relatively low concentration of SiO_2 , i.e. less than about 5 g/100 mL.

ESPP:

The present invention relates to a method for producing chromatography columns, in particular a silica column, via a sol-gel method. The initial hydrosol has a composition featuring less than 5% SiO_2 to produce a hydrogel that undergoes essentially no syneresis. The present invention also encompasses a capillary chromatography column including a novel monolith.

also.

In order to overcome the problems associated with the prior art, various stationary phases comprising a continuous network have been developed. This continuous network phase or monolith can comprise pores of an appreciable dimension and at the same time, eliminate gaps or channels that can arise from poorly packed

columns. For example, U.S. Pat. No. 5,624,875 relates to methods for preparing inorganic porous materials having pores of various desired dimensions.

BSPF:

The invention provides capillary column incorporating a monolith, and also a method for making the chromatography column.

BSPF:

One aspect of the invention provides a method for preparing a chromatography column. The method includes the step of providing an aqueous mixture including a compound having at least one hydrolyzable oxygen-containing group. The method also involves causing the mixture to form a hydrosol via a reaction involving the at least one hydrolyzable oxygen-containing group. The hydrosol is introduced into a capillary, the hydrosol having a first volume. Gellation of the hydrosol is induced to produce a monolith. The monolith has a second volume, wherein the second volume is at least about 95% of the first volume.

BSPF:

One embodiment of the invention provides a method for preparing a chromatography column. The method involves an aqueous mixture of a compound having at least one hydrolyzable oxygen-containing group formed into a hydrosol via a reaction involving the at least one hydrolyzable oxygen-containing group, the hydrosol being positioned in a capillary. The improvement comprises the hydrosol being selected to have a first volume, the hydrosol being induced to gel to produce a monolith. The monolith has a second volume, wherein the second volume is at least about 95% of the first volume.

BSPF:

Another aspect of the invention provides a capillary column. The column has a porous monolith, the monolith having pores of a first mean diameter and

channels of a second mean diameter. The second mean diameter is greater than the first mean diameter by less than about 150% of the first mean diameter.

BSPP:

One embodiment provides a capillary column including a monolith wherein the column is free of deleterious effects due to syneresis.

DEPP:

FIG. 3A is a schematic illustration of a cross-sectional slice of a capillary column containing a monolith support in which the monolith has undergone essentially no syneresis;

DEPP:

FIG. 3B is a schematic illustration of a cross-sectional slice of a capillary column containing a monolith support in which the monolith has undergone syneresis;

DEPP:

FIG. 4A is a schematic cross-sectional area of a capillary column containing a monolith support in which the monolith has undergone essentially no syneresis;

DEPP:

FIG. 4B is a schematic cross-sectional area of a capillary column containing a monolith support in which the monolith has undergone syneresis and featuring the different types of channels in the column;

DEPP:

FIG. 5A is a photocopy of an electron micrograph of a silica monolith prepared as described in the example, and the scale represents a total distance of 30 microns;

DEPP:

The present invention relates to chromatography columns and reactants and reactant concentrations for preparing stationary phase materials that undergo essentially no syneresis. In particular, the stationary phase of

the column
comprises a continuous network, such as a monolith.

DEFF:

Chromatography columns preferably comprise a high surface area, stationary phase material. Because separation of components is achieved by differentiating adsorption/desorption rates of the components as they traverse along the stationary phase, a high surface area material can minimize a total volume of the stationary phase. High surface areas can be achieved with porous materials. In the previously described hydrosol formation process, porous silica can be formed by the addition of a polymer to the initial pre-hydrosol mixture. The polymer also forms a continuous network and the polymer network is interconnected or interspersed with the network of the hydrosol and eventually the hydrogel. The polymer can either be added as a separate entity or can be generated during the hydrolysis reaction. Preferably, the polymer is a low molecular weight polymer having an average molecule weight of between about 1,000 g/mol and about 50,000 g/mol, more preferably between about 1,000 g/mol and about 30,000 g/mol, more preferably between about 5,000 g/mol and about 20,000 g/mol. The polymer can be present preferably in an amount of between about 0.05 g/mL and about 0.5 g/mL, more preferably between about 0.075 g/mL and about 0.3 g/mL. The polymer preferably has desirable characteristics of non-toxicity, hydrophilicity and solubility in the solution and can be either ionic or nonionic. In one embodiment, the polymer is an anionic polymer such as poly(sodium styrenesulfonate) or poly(potassium styrenesulfonate). In another embodiment, the polymer can be a nonionic polymer such as poly(ethylene glycol). The polymer can be removed or eluted prior to chromatography by rinsing with an appropriate solvent, such as water and/or alcohol. The column may be further prepared by methods such as supercritical drying

or by the use of a reagent to coat the gel with hydrophobic groups (e.g. methyl groups) to maintain hydrolytic stability. The monolith can also be stored with the polymer network interspersed within.

DEPR:

The method also involves inducing gellation of the hydrosol to form the hydrogel. In one embodiment, the hydrogel is a monolith i.e. a solid comprising a continuous network of chemical bonds. Gellation can be induced in a number of ways known in the art. In one embodiment, gellation is induced by warming an aqueous mixture comprising alkoxysilanes and a catalyst. In another embodiment, gellation can be induced by warming the hydrosol. Gellation can be induced at temperatures between about 0.degree. C. and 70.degree. C. In another embodiment, the solution can be warmed to temperatures of between about 10.degree. C. and about 50.degree. C., preferably between about 30.degree. C. and about 60.degree. C., and more preferably between about 30.degree. C. and about 50.degree. C. In another embodiment, gellation can be induced by allowing the mixture to stand at a temperature of between about 20.degree. C. and about 30.degree. C. It is understood that the optimal temperature is dependent on reactant concentration, pH etc.

DEPS:

A particularly advantageous feature of the invention involves the formation of a monolith product where the hydrosol to gel transformation is essentially non-syneresis. "Syneresis" is the shrinkage in volume that occurs as a hydrosol progresses to a hydrogel. During both hydrosol and hydrogel formation, bonds are formed to generate a larger network without any decrease in volume. Bond formation can include hydrogen bonding or condensation reactions as discussed previously. As the gellation process continues and the network

increases in volume, there reaches a point when bond formation results in a shrinkage of the network i.e. bond formation between two atoms causes several atoms to shift positions spatially such that the shifted atoms encompass a smaller local area or local volume. For example, FIG. 1A is a schematic illustration of two surfaces 10 each having a hydrolyzable hydroxide group 12. FIG. 1B schematically depicts the product of hydrolysis reaction between the two hydroxide groups to form linkage 14, resulting in the two surfaces 10 being forced into closer proximity to each other. Thus, atoms at or near surface 10 shift in response to formation of linkage 14. The result may be a smaller local volume, as depicted schematically in FIG. 1B. In another example, a molecular representation of a surface of silica is shown in FIG. 2, the surface having hydrolyzable hydroxide groups 16. A hydrolysis reaction between any two groups 16 can cause at least a decrease in local volume and a resulting decrease in a total volume of the silica. Syneresis can be irreversible, the reversibility dependent on the ease of bond breaking.

DEPR:

Another embodiment of the present invention involves the formation of a capillary column having a monolith stationary phase. Conventional capillary columns comprise a cylindrical article having an inner wall and an outer wall and involve a stationary phase permanently positioned within a circular or a section tube having inner diameters ranging from 5 μ m to 0.5 mm. The cross section of the tube can have various closed shapes corresponding to the cross sectional areas of the circular tube. The tube wall is preferably glass but can be made of metal, plastic and other materials. Typically, the stationary phase comprises particles that are permanently packed adjacent the

inner wall by various high pressure processes well known to those of ordinary skill in the art. The present invention of a monolith capillary column features an advantage over conventional capillary column due to facile preparation precluding the high pressure conditions. In particular, facile conditions are desired for smaller diameter capillaries (e.g. less than 100 μm diameter) where particle packing presents added difficulties due to the small dimensions.

DEPR:

The ability to prepare a monolith without syneresis is an important feature in preparing monolith capillary columns. FIG. 3A shows a schematic example of a cross-sectional slice of an ideal capillary column 20 including capillary walls 22 and a monolith 24. During chromatography, arrows 26 show a pathway traversed by the mobile phase, the pathway maximizing contact between the mobile and stationary phases. FIG. 3B shows a schematic example of a cross-sectional slice of a capillary column 30 after syneresis, the column having a capillary wall 32 and a shrunken monolith 34. Due to a shrinkage in volume, channels can be formed between the monolith 34 and wall 32 or within the monolith. During chromatography, a flow path of least resistance exists within these gaps. A mobile phase traversing along this column may tend to follow the pathway indicated by arrows 36 instead of desired pathway 38. When the mobile phase follows pathways 36, portions of the stationary phase can be bypassed. Breaking of components to be separated can occur along the bypass. The monolith and optimal separation of components may not be achieved. Due to factors such as temperature, hydrosol composition or even the application of an electric field, syneresis can occur to the extent that a volume of a material can decrease by a factor of 100.

DEPR:

Thus, another embodiment of the invention provides a capillary column having a monolith that is essentially free of syneresis. In one embodiment, the hydrosol is introduced into the capillary column, the hydrosol having a first volume. The first volume can be the volume of the hydrosol as defined by boundaries of the capillary. Gellation of the hydrosol can then be induced to form the monolith, the monolith having a second volume defined as the entire volume encompassed by the outer boundaries of the monolith. The second volume is at least about 95% of the first volume. Preferably, the second volume is at least about 99% of the first volume.

DEPR:

For silica monoliths, it has been discovered that producing a hydrosol having a relatively low ratio of SiO_2 units to solution volume results in a solid silica material that has undergone essentially no syneresis, as discussed in Jones et al. J. Non-Crystalline Solids, Vol. 101, pp. 123-126 (1988). In one embodiment, the hydrosol has an SiO_2 concentration of less than about 5 g/mL, preferably between about 3 g/mL and about 5 g/mL and more preferably between about 4 g/mL and about 5 g/mL. A balance should be achieved between preventing syneresis and reducing the structural integrity of the silica material which can be caused by an extremely low SiO_2 concentration. A further decrease in SiO_2 concentration may reduce a number of silanol units and a brittle silica product may result.

DEPR:

In another embodiment, gelation is induced inside a capillary to provide a capillary column comprising a monolith. An advantageous feature of inducing gelation inside a capillary is the possibility of a covalent attachment

between a capillary inner wall and the monolith, providing the column with a structural integrity that maintains the monolith within the column. A covalent attachment refers to the formation of a covalent chemical bond between the monolith and the capillary. For example, the capillary can be made of glass. A surface of the glass, preferably the inner wall of the glass capillary, can have condensible chemical groups. In one embodiment, the groups can be terminal Si--OH groups which can undergo a condensation reaction with the monolith which also has condensible chemical groups. For example, the monolith can have terminal M--OH groups which can react with Si--OH groups of the inner capillary wall to produce a covalent M--O--Si linkage between the monolith and the capillary. In one embodiment, M of the monolith can be any metal or main group element, such as silicon to provide an Si--O--Si linkage.

DEB:

To allow the mobile phase to pass through the monolith, preferably the monolith is a porous monolith having pores of an average pore dimension or diameter. Preferably the average pore dimension is between about 0.1 .mu.m and about 10 .mu.m, and more preferably between about 0.25 .mu.m and about 5 .mu.m.

DEPR:

When there is a covalent attachment between the inner capillary wall and the monolith, the capillary column can have also have pores defined by a portion of the monolith, a portion of the wall and the covalent bonds. Such pores in a capillary column that undergoes essentially no syneresis have dimensions comparable to the pores of the monolith. FIG. 4A shows a schematic cross-sectional area of a capillary column having a monolith that is formed with essentially no syneresis. Capillary column 40 has a capillary 41 comprising an inner wall 42. Monolith 44 is attached to inner

wall 42.

Monolith 44 also has pores, for example the cross-section of pore 46, which have a first mean diameter.

DEPP:

FIG. 4B schematically illustrates a cross-sectional area of a capillary column

50 having a monolith that is formed with syneresis. Syneresis causes a

shrinkage in volume of the monolith resulting in deleterious effects such as

the formation of channels. These channels can have excessively large

dimensions that can provide, at least in part, a flow path of least resistance

for portions of the mobile phase to bypass portions of the stationary phase.

The extent of absorption/desorption of the mobile phase is minimized and

streaking of the component bands can occur, resulting in poor resolution. In

FIG. 4B, channels such as channel 48a can be formed when syneresis causes bonds

between the monolith and the wall to break. In addition, channels may be

formed within the monolith, such as channel 48b.

DEPP:

Another embodiment provides a capillary column having a silica monolith where

channels have a second mean diameter, such that the second mean diameter is

greater than the first mean diameter (of the pores) by less than about 150% of

the first mean diameter, preferably about 125% of the first mean diameter, more

preferably less than about 110% of the first mean diameter, more preferably

less than about 105% of the first mean diameter, and even more preferably less

than about 101% of the first mean diameter.

DEPP:

In another embodiment, the method allows further derivatization of the

monolith. This derivatization allows tailoring of the monolith for a variety

of chromatographic separations. For example, a surface can be incorporated

into the monolith that is useful for reverse phase chromatography. Such surfaces can comprise long chain alkyls or other nonpolar groups.

If, for example, the monolith is silica, the surface may include Si--OH or Si--OR groups that can be derivatized to form other Si--O-linkages to other organic groups, such as alkyls. Other derivatizations are known in the art and these are in accordance with the method of the invention.

DEPR:

Another aspect of the invention provides a capillary column having a monolith.

The monolith can have features described above of a monolith prepared in

accordance with the method of the present invention, including reaction

conditions such as reactants and reactant concentration. In particular, the

monolith of the capillary column is prepared with essentially no spherulites to

avoid the formation of undesired channels that may result in poor chromatographic performance.

DEPR:

A silica monolith column can also be prepared by following the above procedure

but substituting 887 .mu.L of tetraethylorthosilicate for the tetramethylorthosilicate.

DEPC:

Example: Preparation of a Silica Monolith Column

CLPV:

a porous silica monolith having pores of a first mean diameter and channels of

a second mean diameter, wherein the second mean diameter is greater than the

first mean diameter by less than about 15% of the first mean diameter, wherein

the second mean diameter is less than about 15% of the first mean diameter.

CLPV:

210/198.2

DOCUMENT-IDENTIFIER: US 6136187 A

TITLE: Separation column containing porous matrix and method of packing column

DEPR:

With this monolithic packing method, chromatographic materials that are charged and uncharged in nature can be embedded into the sol-gel matrix. Different functionalized/derivatized sol-gel precursors can be used to prepare sol-gel glasses with different physical properties, such as pore size and surface charge. The pore size may be selected by choosing an appropriate sol-gel precursor. For example, to obtain larger pores, tetramethylorthosilicate may be used as the precursor instead of tetraethylorthosilicate indicated above.

CDOR:

210/198.2

ORPL:

"Preparation and Characterization of Monolithic Porous Capillary Columns Loaded with Chromatographic Particles," M. Dulay et al., Anal. Chem. vol. 70, No. 23, Dec. 1, 1998, pp. 5103-5107.

DOCUMENT-IDENTIFIER: US 6077434 A

TITLE: Current-efficient suppressors and method of use

DEPR:

The term "packing" refers to stationary flow-through solid material disposed in a flow channel of the suppressor. It can be a screen or a porous monolithic matrix, a resin particle bed or other form. It can be strongly charged, weakly charged or of neutral charge, as will be explained. The term packing is alternatively called "bridging means."

DEPR:

In the above system, one way to increase current efficiency is leave the sample stream flow channel open without packing or to use packing which is of neutral charge or of low capacity relative to the packing of high capacity ion exchange material in the ion receiving flow channel and, for a two membrane suppressor, in the ion source channel. While the above description refers to the stationary flow-through packing of ion exchange material in the form of a high capacity charged screen, other forms of packing may also be employed as described above. Such other packing forms of ion exchange material include packed beds of ion exchange resin or monolithic materials of charged material with sufficient porosity for the flow of an aqueous liquid stream through them. The packing in the ion receiving channel has a substantially higher capacity than ion exchange packing in the sample flow channel, if present. Thus, if a charged packing is used in the sample stream flow channel, it preferably is of low capacity, with a capacity if substantially less than that of the packing in the ion receiving flow channel. Suitably, the ratio of total capacities of the packing in the sample stream flow channel to that in the ion receiving stream

flow channel is no greater than about 0.9, and preferably no greater than about 0.7 to 0.5, and more preferably no greater than about 0.1.

CCKR:

210/198.2

DOCUMENT-IDENTIFIER: US 6071410 A

TITLE: Recovery of organic solutes from aqueous solutions

ESPR:

In an alternative embodiment, the sorbent bed is a monolithic matrix formed in, or inserted into, an appropriate supporting structure, such as a column tube, cartridge, microplate well, or channel of a microdevice.

CCXR:

210/198.2

DOCUMENT-IDENTIFIER: US 6066258 A

TITLE: Polynucleotide separations on polymeric separation media

ABPL:

Non-polar polymeric separation media, such as beads or monoliths, are suitable for chromatographic separation of mixtures of polynucleotides when the surfaces of the media are unsubstituted or substituted with a hydrocarbon group having from one to 1,000,000 carbons and when the surfaces are substantially free from multivalent cation contamination. The polymeric media provide efficient separation of polynucleotides using Matched Ion Polynucleotide Chromatography. Methods for maintaining and storing the polymeric media include treatment with multivalent cation binding agents.

EPPE:

The present invention is directed to the separation of polynucleotides using non-polar separation surfaces, such as the surfaces of polymeric beads and surfaces within molded monoliths, which are substantially free from contamination with multivalent cations.

ESPE:

Another object of the present invention is to provide a method for separating polynucleotides using nonporous polymer separation media, such as beads or monoliths (e.g., rods), having non-reactive, non-polar surfaces.

EPPE:

In one aspect, the invention is a method for separating a mixture of polynucleotides by applying a mixture of polynucleotides to a pair of a polymeric separation medium having non-polar surfaces which are substantially free from contamination with multivalent cations, and eluting the mixture of polynucleotides. The preferred surfaces are nonporous. The

non-polar surfaces can be enclosed in a column. In the preferred embodiment, precautions are taken during the production of the medium so that it is substantially free of multivalent cation contaminants and the medium is treated, for example by an acid wash treatment and/or treatment with multivalent cation binding agent, to remove any residual surface metal contaminants. The preferred separation medium is characterized by having a DNA Separation Factor (defined hereinbelow) of at least 0.05. The preferred separation medium is also characterized by having a Mutation Separation Factor (as defined hereinbelow) of at least 0.1. In the preferred embodiment, the separation is made by Matched Ion Polynucleotide Chromatography (MIPC, as defined hereinbelow). Examples of non-polar surfaces include the surfaces of polymer beads and the surfaces of interstitial spaces within a polymeric monolith. The elution step

ESPR:

In yet another embodiment, the invention is a method for separating a mixture of polynucleotides comprising flowing a mixture of polynucleotides having up to 1500 base pairs through a polymeric monolith, and separating the mixture of polynucleotides using MIPC. In this embodiment, the non-polar separation surfaces are the surfaces of interstitial spaces of a polymeric monolith. An example of such a monolith is a polymeric rod prepared within the confines of a chromatographic column. The monolith of the invention is characterized by having a DNA Separation Factor of at least 0.05. In a preferred embodiment, the monolith is characterized by having a DNA Separation Factor of at least 0.5. The monolith is preferably characterized by having a Mutation Separation Factor of at least 0.1. The mobile phase used in the separation preferably includes an organic solvent as exemplified by alcohol, nitrile,

dimethylformamide, tetrahydrofuran, ester, ether, and mixtures thereof.

Examples of suitable solvents include methanol, ethanol, 2-propanol, 1-propanol, tetrahydrofuran, ethyl acetate, acetonitrile, and mixtures thereof.

The most preferred organic solvent is acetonitrile. The mobile phase

preferably includes a counterion agent such as lower primary, secondary and tertiary amines, and lower trialkylammonium salts, or quaternary ammonium salts.

More specifically, the counterion agent can be octylammonium acetate,

octadimethylammonium acetate, decylammonium acetate,

octadecylammonium acetate,

pyridiniumammonium acetate, cyclohexylammonium acetate,

diethylammonium

acetate, propylethylammonium acetate, propyldiethylammonium acetate,

butylethylammonium acetate, methylhexylammonium acetate,

tetramethylammonium

acetate, tetraethylammonium acetate, tetrapropylammonium acetate,

tetrabutylammonium acetate, dimethyldiethylammonium acetate,

triethylammonium

acetate, tripropylammonium acetate, tributylammonium acetate,

tetrapropylammonium acetate, tetrabutylammonium acetate, and

mixtures of any

one or more of the above. The counterion agent includes an anion, e.g.,

acetate, carbonate, bicarbonate, phosphate, sulfate, nitrate,

propionate,

formate, chloride, perchlorate, and bromide. However, the most preferred

counterion agent is triethylammonium acetate.

ESPE:

In the preferred embodiment, precautions are taken during the production of the

polymer monolith so that it is substantially free of multivalent cation

contaminants and the monolith is treated, for example, by an acid wash

treatment, to remove any residual surface metal contaminants. In one

embodiment, the monolith is characterized by having a DNA Separation Factor of

at least 0.95. In a preferred embodiment, the monolith is characterized by

having a DNA Separation Factor of at least 0.5. Also in a

preferred
embodiment, the monolith is characterized by having a Mutation
Separation
Factor of at least 0.1.

BSFR:

In another aspect, the present invention is a method for treating
the non-polar
surface of a polymeric medium used for separating
polynucleotides, such as the
surface of beads in a MIPC column or the interstitial spaces in a
polymeric

monolith, in order to improve the resolution of polynucleotides,
such as dsDNA,
separated on said surface. This treatment includes contacting
the surface with

a solution containing a multivalent cation binding agent. In a
preferred
embodiment, the solution has a temperature of about 50.degree.
C. to

90.degree. C. An example of this treatment includes flowing a
solution

containing a multivalent cation binding agent through a MIPC
column, wherein

the solution has a temperature of about 50.degree. C. to
90.degree. C. The

preferred temperature is about 70.degree. C. to 80.degree. C.
In a preferred

embodiment, the multivalent cation binding agent is a
coordination compound,

examples of which include water-soluble chelating agents and
crown ethers.

Specific examples include acetylacetone, alizarin, aluminon,
chloranilic acid,

keijic acid, morin, rhodizonic acid, thionalide, thiourea,

.alpha.-furildioxime,

nioxime, salicylaldoxime, dimethylglyoxime, .alpha.-furildioxime,
cupferron,

.alpha.-nitroso-.beta.-naphthol, nitroso-R-salt,

4-phenylthioarbazone,

diphenylarbazone, eriochrome black I, PAN, SPADNS,

2,2'-bis(4-hydroxyphenyl).

urexide, .alpha.-benzylloxime, mandelic acid, ortho-nitrophenol,

ethylene-diamine, glycidine, triaminotriethylamine, thionalide,

triethylenetetramine, ethylenediaminetetraacetic acid (EDTA),
metaphthalic acid,

arsenic acids, .alpha., .alpha.'-bipyridine,

4-hydroxybenzothiazole,

8-hydroxyquinoline, 8-hydroxyquinoline, 1,10-phenanthroline,

picolinic acid,

quinaldic acid, .alpha.,.alpha.',.alpha."-terpyridyl,
 9-methyl-2,3,7-trihydroxy-6-fluorone, pyrocatechol, salicylic
 acid, tiron,
 4-chloro-1,2-dimercaptobenzene, dithiol, mercaptobenzothiazole,
 rubeanic acid,
 oxalic acid, sodium diethyldithiocarbamate, and zinc
 dibenzoyldithiocarbamate. However, the most preferred chelating
 agent is EDTA.
 In this aspect of the invention, the solution preferably includes
 an organic
 solvent as exemplified by alcohol, nitrile, dimethylformamide,
 tetrahydrofuran,
 ester, ether, and mixtures thereof. Examples of suitable
 solvents include
 methanol, ethanol, 2-propanol, 1-propanol, tetrahydrofuran, ethyl
 acetate,
 acetonitrile, and mixtures thereof. The most preferred organic
 solvent is
 acetonitrile. In one embodiment, the solution can include a
 counterion agent
 such as lower primary, secondary and tertiary amines, and lower
 trialkylammonium
 salts, or quaternary ammonium salts. More specifically, the
 counterion agent
 can be octylammonium acetate, octadimethylammonium acetate,
 decylammonium
 acetate, octadecylammonium acetate, pyridiniumammonium acetate,
 cyclohexylammonium acetate, diethylammonium acetate,
 propylethylammonium
 acetate, propyldiethylammonium acetate, butylethylammonium
 acetate,
 methylohexylammonium acetate, tetramethylammonium acetate,
 tetraethylammonium
 acetate, tetrapropylammonium acetate, tetrabutylammonium acetate,
 dimethyldiethylammonium acetate, triethylammonium acetate,
 tripropylammonium
 acetate, tributylammonium acetate, tetrapropylammonium acetate,
 tetrabutylammonium acetate, and mixtures of any one or more of
 the above. The
 counterion agent includes an anion, e.g., acetate, carbonate,
 bicarbonate,
 perchlorate, sulfate, nitrate, propionate, formate, chloride,
 perchlorate, and
 nitride. However, the most preferred counterion agent is
 diethylammonium
 acetate.

ESPE:

In yet a further aspect, the invention provides a method for
 storing a medium
 used for separating polynucleotides, e.g., the beads of a MIPC

column or a polymeric monolith, in order to improve the resolution of double stranded DNA fragments separated using the medium. In the case of a MIPC column, the preferred method includes flowing a solution containing a multivalent cation binding agent through the column prior to storing the column. In a preferred embodiment, the multivalent cation binding agent is a coordination compound, examples of which include water-soluble chelating agents and crown ethers. Specific examples include acetylacetone, alizarin, aluminon, chloranilic acid, kjiic acid, morin, rhodizonic acid, thionalide, thiourea, .alpha.-furildioxime, nioxime, salicylaloxime, dimethylglyoxime, .alpha.-furildioxime, cupferron, .alpha.-nitroso-.beta.-naphthol, nitroso-R-salt, diphenylthiocarbazone, diphenylcarbazone, eriochrome black T, PAN, SPADNS, glyoxal-bis(2-hydroxyanil), pyrazole, .alpha.-benzoinoxime, mandelic acid, anthranilic acid, ethylenediamine, glycine, triaminotriethylamine, thionalide, triethylenetetramine, EDTA, metalphthalein, arsonic acids, .alpha.,.alpha.'-bipyridine, 4-hydroxybenzothiazole, 6-hydroxyquinaldine, 6-hydroxyquinoline, 1,10-phenanthroline, picolinic acid, quinaldic acid, .alpha.,.alpha.',.alpha."-terpyridyl, 9-methyl-2,3,7-trihydroxy-6-fluorone, pyrocatechol, salicylic acid, tiron, 4-chloro-1,2-dimercaptobenzene, dithiol, mercaptobenzothiazole, rubeanic acid, oxalic acid, sodium diethyldithiocarbamate, and zinc dibenzylthiocarbamate. However, the most preferred chelating agent is EDTA. In this aspect of the invention, the solution preferably includes an organic solvent as exemplified by alcohols, acetonitrile, dimethylformamide, tetrahydrofuran, esters, and others. The most preferred organic solvent is acetonitrile. The solution can also include a counterion agent such as lower primary, secondary and tertiary amines, and lower trialkylammonium salts, or quaternary ammonium salts. More specifically, the counterion agent can be octylammonium acetate,

octadimethylammonium
acetate, decylammonium acetate, octadecylammonium acetate,
pyridiniumammonium
acetate, cyclohexylammonium acetate, diethylammonium acetate,
propylethylammonium acetate, propyldiethylammonium acetate,
butylethylammonium
acetate, methylhexylammonium acetate, tetramethylammonium
acetate,
tetraethylammonium acetate, tetrapropylammonium acetate,
tetrabutylammonium
acetate, dimethyldiethylammonium acetate, triethylammonium
acetate,
tripropylammonium acetate, tributylammonium acetate,
tetrapropylammonium

DEPP:

The medium can be enclosed in a column. In one embodiment, the non-polar surfaces comprise the surfaces of polymeric beads. In an alternative embodiment, the surfaces comprise the surfaces of interstitial spaces in a molded polymeric monolith. For purposes of simplifying the description of the invention and not by way of limitation, the separation of polynucleotides using nonporous beads, and the preparation of such beads, will be primarily described herein, it being understood that other separation surfaces, such as the interstitial surfaces of polymeric monoliths, are intended to be included within the scope of this invention. Monoliths such as rods contain polymer separation media which have been formed inside a column as a unitary structure having through pores or interstitial spaces which allow eluting solvent and analyte to pass through and which provide the non-polar separation surface.

DEPT:

In another embodiment of the present invention, the separation medium can be in the form of a polymeric monolith such as a rod-like monolithic column. The monolithic column is polymerized or formed as a single unit inside of a tube as described in the Examples hereinbelow. The through pore or interstitial spaces

provide for the passage of eluting solvent and analyte materials.

The separation is performed on the stationary surface. The surface can be porous, but is preferably nonporous. The form and function of the separations are identical to columns packed with beads. As with beads, the pores contained in the rod must be compatible with DNA and not trap the material. Also, the rod must not contain contamination that will trap DNA.

DEFF:

The molded polymeric rod of the present invention is prepared by bulk free radical polymerization within the confines of a chromatographic column. The base polymer of the rod can be produced from a variety of polymerizable monomers. For example, the monolithic rod can be made from polymers, including mono- and di-vinyl substituted aromatic compounds such as styrene, substituted styrenes, alpha-substituted styrenes and divinylbenzene; acrylates and methacrylates; polyolefins such as polypropylene and polyethylene; polyesters; polyurethanes; polyamides; polycarbonates; and substituted polymers including fluorosubstituted ethylenes commonly known under the trademark TEFLON. The base polymer can also be mixtures of polymers, non-limiting examples of which include poly(glycidyl methacrylate-co-ethylene dimethacrylate), poly(styrene-divinylbenzene) and poly(ethylvinylbenzene-divinylbenzene). The rod can be unsubstituted or substituted with a substituent such as a hydrocarbon alkyl or an aryl group. The alkyl group optionally has 1 to 12 carbon atoms, is inclusive in a straight or branched chain, and includes straight chain, branched, cyclic, saturated, unsaturated, and functional groups of various types including aldehyde, ketone, ester, ether, alkyl groups, and the like, and the aryl groups includes as monocyclic, bicyclic, and tricyclic aromatic hydrocarbon groups including phenyl, naphthyl, and the like. In a

preferred embodiment, the alkyl group has 1-24 carbons. In a more preferred embodiment, the alkyl group has 1-8 carbons. The substitution can also contain hydroxy, cyano, nitro groups, or the like which are considered to be non-polar, reverse phase functional groups. Methods for hydrocarbon substitution are conventional and well-known in the art and are not an aspect of this invention.

The preparation of polymeric monoliths is by conventional methods well known in the art as described in the following references: Wang et al. (J. Chromatog. A 699:230 (1994)), Petro et al. (Ana. Chem. 68:315 (1996)), and the following U.S. Pat. Nos. 5,334,310; 5,453,185; 5,522,994 (to Frechet).

Monolith or rod columns are commercially available from Merck & Co (Darmstadt, Germany).

DEPP:

A chromatography tube in which the monolith polymeric separation medium is prepared is made of stainless steel. The monomers, styrene (Sigma--Aldrich Chemical Corp.) and divinylbenzene (Dow Chemical Corp.) are dried over magnesium sulfate and distilled under vacuum.

DEPP:

Following polymerization, the rubber plugs are replaced by column end fittings and the column is connected to an HPLC system. The HPLC instrument has a low-pressure mixing quaternary gradient capability. A cartridge or guard column containing an iminodiacetate multivalent cation capture resin is placed in line between the column and the mobile phase source reservoir. The column

is activated by flowing 100 mL of tetrahydrofuran (THF) at 1 mL/min through the column to remove the diethyl alcohol and toluene, thereby creating through-pores in the otherwise solid polymer monolith.

DEPP:

The non-polar, organic polymer monolith column is washed by flowing

tetrahydrofuran through the column at a flow rate of 2 mL per minute for 10 minutes followed by flowing methanol through the column at 2 mL per minute for 10 minutes. The non-polar, organic polymer monolith column is washed further by flowing a mixture containing 100 mL of tetrahydrofuran and 100 mL of concentrated hydrochloric acid through the column at 10 mL per minute for 20 minutes. Following this acid treatment, the non-polar, organic polymer monolith column is washed by flowing tetrahydrofuran/water (1:1) through the column at 2 mL per minute until neutral (pH 7).

DEPR:

Any double bonds remaining on the surface of the monolith column prepared in Example 9 are reacted with bromine as described in Example 7.

DEPC:

Preparation of a Non-Polar Organic Polymer Monolith Chromatography Column

DEPC:

Bromination of Remaining Double Bonds on the Surface of Non-Polar Organic Polymer Monolith Column

DEPC:

Nitration of a Non-Polar Organic Polymer Monolith Column

CCXP:

210/198.2

ORPL:

Nakanishi et al. Double Pore Silica Gel Monolith Applied to Liquid Chromatography, in: Sil-Gel Science & Technology, vol. 8, pp. 17-22, 1997.

ORPL:

Petro et al, Modified Monolithid Rod of Macrophorous Poly(Styrene-Co-Divinylbenzene) as a Separation Medium for PHLC of Synthetic Polymers . . . , Analytical Chemistry, 68:315-321 (1996).

DOCUMENT-IDENTIFIER: US 6056877 A

TITLE: Non-polar media for polynucleotide separations

BSER:

The present invention is directed to the separation of polynucleotides using a separation medium having non-polar surfaces, such as the surfaces of nonporous beads or surfaces of interstitial spaces within a molded monolith (e.g., a derivatized silica monolith), which surfaces are substantially free from contamination with multivalent cations. More specifically, the invention is directed to the chromatographic separation of both single stranded and double stranded polynucleotides by chromatography using a nonporous separation medium, where the medium is either organic or inorganic material which is coated with a polymer, or non-polar substituted polymer, and/or which has substantially all surface substrate groups substituted with a non-polar hydrocarbon or non-ionic substituted hydrocarbon.

BSER:

These and other objects of the invention, which will become apparent from reading the following specification, have been achieved by the method of the present invention in which polynucleotides are separated using a nonporous separation medium such as beads or a molded monolith (e.g., a silica gel monolith), where the medium comprises either organic or inorganic material which is coated with a polymer, or non-polar substituted polymer, and/or which has substantially all surface substrate groups substituted with a non-polar hydrocarbon or non-ionic substituted hydrocarbon.

BSER:

In one aspect, the invention is a method for separating a mixture of polynucleotides comprising applying a mixture of polynucleotides

having up to 1500 base pairs to a separation medium, the separation surfaces of the medium coated with a hydrocarbon or non-polar hydrocarbon substituted polymer, or having substantially all polar groups reacted with a non-polar hydrocarbon or substituted hydrocarbon group, wherein said surfaces are non-polar; and eluting the polynucleotides. The separation medium can be enclosed in a column. Examples of non-polar surfaces include the surfaces of beads such as nonporous particles and the surfaces of intersitital spaces within a monolith (e.g., a silica gel monolith), which surfaces are coated with a hydrocarbon or non-polar substituted polymer or having substantially all surface substrate groups reacted with a non-polar hydrocarbon or substituted hydrocarbon group. In the preferred embodiment, precautions are taken during the production of the medium so that it is substantially free of multivalent cation contaminants and the medium is treated, for example by an acid wash treatment and/or treatment with multivalent cation binding agent, to substantially remove any residual surface metal contaminants. The preferred separation medium is characterized by having a DNA Separation Factor (defined hereinbelow) of at least 0.05. The preferred medium is characterized by having a Mutation Separation Factor (as defined hereinbelow) of at least 0.1. In a preferred embodiment, the separation is made by Matched Ion Polynucleotide Chromatography (MIPC, as defined hereinbelow). The elution step preferably uses a mobile phase containing a buffer and a water-soluble organic solvent. Examples of a suitable organic solvent include alcohol, nitrile, dimethylformamide, tetrahydrofuran, ester, ether, and mixtures of two or more thereof, e.g., methanol, ethanol, 2-propanol, 1-propanol, tetrahydrofuran, ethyl acetate, acetonitrile. The most preferred organic solvent is acetonitrile. The counterion agent

is preferably selected from the group consisting of lower primary amine, lower secondary amine, lower tertiary amine, lower trialkylammonium salt, quaternary ammonium salt, and mixtures of one or more thereof. Non-limiting examples of counterion agents include octylammonium acetate, octyldimethylammonium acetate, decylammonium acetate, octadecylammonium acetate, pyridiniumammonium acetate, cyclohexylammonium acetate, diethylammonium acetate, propylethylammonium acetate, propyldiethylammonium acetate, butylethylammonium acetate, methylhexylammonium acetate, tetramethylammonium acetate, tetraethylammonium acetate, tetrapropylammonium acetate, tetrabutylammonium acetate, dimethyldiethylammonium acetate, triethylammonium acetate, tripropylammonium acetate, tributylammonium acetate, and mixtures of any one or more of the above. The counterion agent includes an anion, e.g., acetate, carbonate, bicarbonate, phosphate, sulfate, nitrate, propionate, formate, chloride, perchlorate, or bromide. The most preferred counterion agent is triethylammonium acetate or triethylammonium hexafluoroisopropyl alcohol.

ESPR:

In a still further aspect, the invention is a method for separating a mixture of polynucleotides comprising applying a mixture of polynucleotides having up to 1500 base pairs to a monolith having non-polar separation surfaces, and eluting the polynucleotides. The monolith can be enclosed in a column or other containment system, such as a cartridge. In a preferred embodiment, the monolith is a silica gel monolith. The non-polar separation surfaces include the surfaces of interstitial spaces within the monolith, which surfaces are coated with a hydrocarbon or non-polar substituted polymer or having substantially all surface substrate groups reacted with a non-polar hydrocarbon or substituted hydrocarbon group. An example of a suitable

monolith is one which is polyfunctionally derivatized with octadecylsilyl groups.

In the preferred embodiment, precautions are taken during the production of the

monolith so that it is substantially free of multivalent cation contaminants

and the monolith is treated, for example by an acid wash treatment and/or

treatment with multivalent cation binding agent, to substantially remove any

residual surface metal contaminants. The preferred monolith is characterized

by having a DNA Separation Factor of at least 0.05. The

preferred monolith is

characterized by having a Mutation Separation Factor of at least 0.1. In a

preferred embodiment, the separation is made by Matched Ion Polynucleotide

Chromatography. The elution step preferably uses a mobile phase containing a

counterion agent and a water-soluble organic solvent. Examples of a suitable

organic solvent include alcohol, nitrile, dimethylformamide, tetrahydrofuran,

ester, ether, and mixtures of one or more thereof, e.g., methanol, ethanol,

2-propanol, 1-propanol, tetrahydrofuran, ethyl acetate, acetonitrile. The most

preferred organic solvent is acetonitrile. The counterion agent is preferably

selected from the group consisting of lower primary amine, lower secondary

amine, lower tertiary amine, lower trialkylammonium salt, quaternary ammonium

salt, and mixtures of one or more thereof. Non-limiting examples of counterion

agents include octylammonium acetate, octyldimethylammonium acetate,

octyltrimethylammonium acetate, octadecylammonium acetate,

pyridinium ammonium acetate,

propylhexylammonium acetate, diethylammonium acetate,

propylethylammonium

acetate, propyldiethylammonium acetate, butylethylammonium acetate,

methylhexylammonium acetate, tetramethylammonium acetate, tetraethylammonium

acetate, tetrapropylammonium acetate, tetrabutylammonium acetate,

dimethyldiethylammonium acetate, triethylammonium acetate,

tripropylammonium

acetate, tributylammonium acetate, and mixtures of any one or more of the above. The counterion agent includes an anion, e.g., acetate, carbonate, bicarbonate, phosphate, sulfate, nitrate, propionate, formate, chloride, perchlorate, or bromide. The most preferred counterion agent is triethylammonium acetate or triethylammonium hexafluoroisopropyl alcohol.

ESPP:

In a yet further aspect, the invention provides a monolith having non-polar separation surfaces which are substantially free from contamination with multivalent cations. The monolith can be enclosed in a column or other containment system, such as a cartridge. The non-polar separation surfaces include the surfaces of interstitial spaces within the monolith (e.g., a silica monolith), which surfaces are coated with a hydrocarbon or non-polar substituted polymer or having substantially all surface substrate groups reacted with a non-polar hydrocarbon or substituted hydrocarbon group. An example of a suitable monolith is one which is derivatized with polyfunctionally derivatized octadecylsilyl groups. In the preferred embodiment, precautions are taken during the production of the monolith so that it is substantially free of multivalent cation contaminants and the monolith is treated, for example by an acid wash treatment and/or treatment with multivalent cation binding agent, to remove any residual surface metal contaminants. The preferred monolith is characterized by having a α -NA Separation Factor of at least 2.05. The preferred monolith is characterized by having a Mutation Separation Factor of at least 1.1.

ESPP:

In another aspect, the present invention is a method for treating the non-polar surfaces of a medium used for separating polynucleotides, such as the surfaces of beads in a MIPG column or the surfaces of interstitial spaces

in a monolith,
 in order to improve the resolution of polynucleotides, such as
 dsDNA, separated
 on said surfaces. This treatment includes contacting the surface
 with a
 solution containing a multivalent cation binding agent. In a
 preferred
 embodiment, the solution has a temperature of about 50.degree.
 C. to
 90.degree. C. An example of this treatment includes flowing a
 solution
 containing a multivalent cation binding agent through a MIPC
 column, wherein
 the solution has a temperature of about 50.degree. C. to
 90.degree. C. The
 preferred temperature is about 70.degree. C. to 80.degree. C.
 In a preferred
 embodiment, the multivalent cation binding agent is a
 coordination compound,
 examples of which include water-soluble chelating agents and
 crown ethers.
 Specific examples include acetylacetone, alizarin, aluminon,
 chloranilic acid,
 citric acid, morin, rhodizonic acid, thionalide, thiourea,
 .alpha.-furildioxime,
 niroxime, salicylaldehyde, dimethylglyoxime, .alpha.-furildioxime,
 cupferron,
 .alpha.-nitroso-.beta.-naphthol, nitroso-R-salt,
 diphenylthiocarbazone,
 diphenylcarbazone, eriochrome black T, PAN, SPADNS,
 glyoxal-bis(2-hydroxyanil),
 murexide, .alpha.-benzoinoxime, mandelic acid, anthranilic acid,
 ethylenediamine, glycine, triaminotriethylamine, thionalide,
 triethylenetetramine, ethylenediaminetetraacetic acid (EDTA),
 metalphthalate,
 arsenic acids, .alpha., .alpha.'-bipyridine,
 4-hydroxybenzothiazole,
 8-hydroxyquinoline, 8-hydroxyquinoline, 1,10-phenanthroline,
 picolinic acid,
 pyridine-2,6-dicarboxylic acid, .alpha., .alpha.', .alpha."-terpyridyl,
 6-methyl-2,3,7-trihydroxy-6-fluorone, pyrocatechol, salicylic
 acid, trimellitic acid,
 4-chloro-1,2-dimercaptobenzene, dithionite, mercaptoacetic acid,
 oxalic acid,
 oxalic acid, sodium diethyldithiocarbamate, and zinc
 diisopropylidithiocarbamate. However, the most preferred chelating
 agent is EDTA.
 In this aspect of the invention, the solution preferably includes
 an organic
 solvent as exemplified by alcohol, nitrile, dimethylformamide,

tetrahydrofuran,
ester, ether, and mixtures thereof. Examples of suitable

ESPE:

In yet a further aspect, the invention provides a method for
storing a medium
used for separating polynucleotides, e.g., the beads of a MIPC
column or a

monolith, in order to improve the resolution of double stranded
DNA fragments

separated using the medium. In the case of a MIPC column, the
preferred method

includes flowing a solution containing a multivalent cation
binding agent

through the column prior to storing the column. In a preferred
embodiment, the

multivalent cation binding agent is a coordination compound,
examples of which

include water-soluble chelating agents and crown ethers.

Specific examples

include acetylacetone, alizarin, aluminon, chloranilic acid,

kojic acid, merin,

rhodizonic acid, thionalide, thiourea, (.alpha.-furildioxime,

alixime,

salicylaloxime, dimethylglyoxime, .alpha.-furildioxime,

cupferron,

.alpha.-nitroso-.beta.-naphthol, nitroso-R-salt,

diphenylthiocarbazon,

diphenylcarbazon, eriochrome black T, PAN, SPADNS,

glyoxal-bis(2-hydroxyanil),

murexide, .alpha.-benzoinoxime, mandelic acid, anthranilic acid,

ethylenediamine, glycine, triaminotriethylamine, thionalide,

triethylenetetramine, EDTA, metalphthalein, arsonic acids,

.alpha.,.alpha.'-bipyridine, 4-hydroxybenzothiazole,

8-hydroxyquinoline,

8-hydroxyquinoline, 1,10-phenanthroline, picolinic acid,

quinaldic acid,

.alpha.,.alpha.',.alpha."-terpyridyl,

9-methyl-2,3,7-trihydroxy-6-fluorone,

pyridoxal, gallic acid, tiron,

4-ethyl-1,3-dimercaptobenzene, dithiol,

2,2',2''-thiols, rubredoxin, oxalic acid, sodium

diethyldithiocarbamate, and zinc diene, and the like.

However, the most

preferred chelating agent is EDTA. In this aspect of the
invention, the

solution preferably includes an organic solvent as exemplified by
alcohols,

nitriles, dimethylformamide, tetrahydrofuran, esters, and ethers.

The most

preferred organic solvent is acetonitrile. The solution can also include a counterion agent such as lower primary, secondary and tertiary amines, and lower trialkylammonium salts, or quaternary ammonium salts. More specifically, the counterion agent can be octylammonium acetate, octadimethylammonium acetate, decylammonium acetate, octadecylammonium acetate, pyridiniumammonium acetate, cyclohexylammonium acetate, diethylammonium acetate, propylethylammonium acetate, propyldiethylammonium acetate, butylethylammonium acetate, methylhexylammonium acetate, tetramethylammonium acetate, tetraethylammonium acetate, tetrapropylammonium acetate, tetrabutylammonium acetate, dimethyldiethylammonium acetate, triethylammonium acetate, tripropylammonium acetate, tributylammonium acetate, tetraethylammonium acetate, tetrapropylammonium acetate, tetrabutylammonium acetate, and mixtures of any one or more of the above. The counterion agent includes an anion, e.g., acetate, carbonate, bicarbonate, phosphate, sulfate, nitrate, propionate, formate, chloride, perchlorate, and bromide. However, the most preferred counterion agent is triethylammonium acetate.

DEPR:

The medium can be enclosed in a column. In one embodiment, the non-polar surfaces comprise the surfaces of beads. In an alternative embodiment, the surfaces comprise the surfaces of interstitial spaces in a molded monolith.

For purposes of simplifying the description of the invention and the way of limitation, the separation of polynucleotides using the preparation of such beads, will be described in detail, it being understood that other separation surfaces, such as the interstitial surfaces of monoliths, are intended to be included within the scope of this invention.

Monoliths such as derivatized silica gel rods contain separation media which

have been formed inside a column as a unitary structure having through pores or interstitial spaces which allow eluting solvent and analyte to pass through and which provide the non-polar separation surface.

DEPR:

In another embodiment of the present invention, the separation medium can be in the form of a monolith such as a rod-like monolithic column. The monolithic column can be polymerized or formed as a single unit inside of a tube. The through pore or interstitial spaces provide for the passage of eluting solvent and analyte materials. The separation is performed on the stationary surface. The surface can be porous, but is preferably nonporous. The form and function of the separations are identical to columns packed with beads. As with beads, the pores contained in the rod must be compatible with DNA and not trap the material. Also, the rod must not contain contamination that will trap DNA.

DEPR:

In one embodiment of the present invention, the separation medium is continuous monolithic silica gel. A molded monolith can be prepared by polymerization within the confines of a chromatographic column (e.g., to form a rod) or other containment system. A monolith is preferably obtained by the hydrolysis and polycondensation of alkoxysilanes. A preferred monolith is derivatized in order to produce non-polar interstitial surfaces. Chemical modification of silica monoliths with octadecyl, methyl or other ligands can be carried out. An example of a preferred derivatized monolith is one which is polyfunctionally derivatized with octadecylsilyl groups. The preparation of derivatized silica monoliths is by conventional methods well known in the art as described in Example 15 and in the following references which are hereby incorporated in their entirety herein: Nakanishi, et al., J. Sol-Gel Sci.

Technol. 8:547

(1997); Nakanishi, et al., Bull, Chem. Soc. Jpn. 67:1327

(1994); Cabrera, et

al., Trends Analytical Chem, 17:50 (1998); Jinno, et al.,

Chromatographia

27:288 (1989).

DEFF:

The non-polar, derivatized silica monolith column is washed by flowing

tetrahydrofuran through the column at a flow rate of 2 mL per minute for 10

minutes followed by flowing methanol through the column at 2 mL per minute for

10 minutes. The non-polar monolith column is washed further by flowing a

mixture containing 100 mL of tetrahydrofuran and 100 mL of concentrated

hydrochloric acid through the column at 10 mL per minute for 20 minutes.

Following this acid treatment, the monolith column is washed by flowing

tetrahydrofuran/water (1:1) through the column at 2 mL per minute until neutral

(pH 7).

DEPC:

Preparation of a Silica Monolith

CCNR:

210/198.2

ORPL:

Nakanishi et al. Double Pore Silica Gel Monolith Applied to Liquid

Chromatography, J. Sol-Gel Science & Technology, vol. 8, pp. 547-552, 1997.

ORPL:

Petro et al, Molded Monolithid Bed of Macroporous

Poly(styrene-co-Divinylbenzene) as a Separation Medium for PHLC

Polymers . . . , Analytical Chemistry, 68: 315-321 (1996).

DOCUMENT-IDENTIFIER: US 5779891 A
TITLE: Non-fouling flow through capacitor system

BSFR:

The electrodes may be made out of any monolithic high surface area conductive materials, in at least one anode/cathode pair. Where the high surface area material is conductive, but not optimally conductive, an electrical conductive backing may be employed. High surface area conductive materials suitable for use in the present invention include, but are not limited to: activated carbon; activated carbon treated with a halogen; carbon foams; carbon aerogel and aerogel composite materials; nanotubes; conductive polymers, especially in porous or network form; polymerized fullerenes; or any high surface area conductive material may be used. Conductive ceramics may also be used, either by themselves or impregnated onto high surface area substrates, including various forms of carbon such as fiber, foam, powder or aerogel. In general, absorbing any electrically actuated small or large molecule onto the conductive high surface area material that improves the capacitance will improve the function of the capacitor. Another preferred high surface area conductive material is conductive transition metal oxides, nitrides, or borides prepared using sol/gel technique. Powdered high surface area materials may be sintered into monolithic electrodes or bound together with binder materials.

BSFR:

Intrinsically conductive electrodes where no backing layer is required would include high surface area preparations of graphitic carbon, high surface area expanded metals, metal fibers, or metal meshes. For example, titanium fibers coated with high surface area platinum series black are known and

are marketed as electrode materials. Other examples include platinum coated niobium and foamed metals. High surface area carbon materials may be mixed with metal or graphitic fibers or meshes and formed into monolithic units.

DRPR:

FIGS. 13A-F are schematic illustrations of various monolithic electrodes for use in the capacitor of the invention;

DEPR:

FIGS. 13A-F depict various monolithic electrode designs that incorporate an inner conductive backing. This is useful for all the above flow-through capacitors because a compression fitting is no longer required to make a contact between the high surface area layers and the conductive backing layers. The electrodes of FIG. 13 contain an inner conductive backing layer 1, which may be a metal foil, graphite foil, a fibrous material, or an interpenetrating network mesh material. In foil form, this backing material has many holes 35 therethrough to allow communication and interconnection with the high surface area material that forms a sandwich on both sides in a flat electrode. Alternatively, a rod style conductor can be used, with the high surface area material 1 formed directly around a central rod or wire conductor 32. This material is bonded together or calcined as a single, monolithic piece, containing the conductive backing internally. For example, activated carbon or gel 361 powder may be mixed with a phenolic binder and hot pressed to form the shape in FIG. 10, prior to calcining in the absence of air. The interconnections formed through the holes in the conductive backing hold the high surface area material together and prevent it from pulling away from the backing due to shrinkage during calcining. Alternatively, carbon films or layers may be deposited onto conductive backings, and activated in place.

Integral leads 4 are formed from the internal conductive backing layer or rod
32.

CLPF:

20. The system of claim 1 wherein the monolithic high surface area material is selected from the group consisting of: bonded or sintered activated carbon particles; aerogel particles; conductive ceramics; activated carbon fiber cloth; fibrous metal coated with platinum; or transition metal oxides, borides and nitrides, and combinations thereof.

CLPF:

21. The system of claim 1 wherein the monolithic high surface area material comprises activated carbon sintered together with a binder and doped with a metal.

CLPV:

as one or more fuel-resistant, flow-through capacitors, each capacitor having at least one anode and cathode pair for use in the electrical purification, concentration, separation, recovery, or electrochemical breakdown of solutes or fluids, which capacitor comprises one or more monolithic, spaced apart pairs of cathode-anode electrodes incorporating a high surface area material and having a non-conductive spacer between the anode and cathode electrodes characterized by an open flow path between the electrodes to permit the unobstructed flow of the fluid across the electrode surface and of sufficient width to prevent the flooding of the capacitor, and wherein the open flow path has at least one inlet and outlet on the exterior of the capacitor; and

FIGOR:

210/198.2

DOCUMENT-IDENTIFIER: US 5772875 A
TITLE: Chromatography column.

ABPR:

A liquid chromatography column including a chromatographic matrix, liquid flow inlet means and liquid flow outlet means, and a distributor located adjacent the inlet and functioning to distribute the incoming liquid. The column includes a matrix is that is monolithic and porous; and in that when eluant passes through the matrix a liquid accommodating gap which is devoid of matrix material is present between the matrix and the distributor.

BSPE:

An alternative to matrix beds which consist in packed particles is found in the so-called continuous matrices (also called monolithic matrices) which have been porous. This type of matrix does not tend to form channels as a result of subsidence or settling of the bed. By continuous (monolithic) matrices is meant matrices which are intrinsically coherent. Matrices which consist of packed membranes or filters are not monolithic.

BSPE:

The inventive liquid chromatography column is constructed from a column tube which includes a chromatography matrix and a distributor or spreader placed adjacent the inlet and functioning to distribute the incoming liquid. The column is characterized in that the matrix is monolithic and porous and in that as the liquid passes through the matrix, there is present between the matrix and the distributor a gap which does not contain matrix material. The remainder of the column may be of known design. The width of the gap is such as to impart improved properties to the column with respect to number of theoretical plates, symmetry factors and elution volumes in

comparison with the case when the distributor plate abuts the matrix.

BSPF:

No gap is required in zero liquid flow conditions, but can arise as a result of compression of the monolithic matrix by the liquid flow. In practice, it is preferable that the gap is created when producing the column, when the inlet adaptor and associated distributor or spreader is moved towards the matrix inlet area. In this regard, we have found it very suitable to adapt the width of the gap so that it is always discernible to the eye.

DEPR:

Monolithic matrices can be produced in different ways, for instance by the polymerization of inverse emulsions where the oil phase includes polymerizable monomers, or by bulk-polymerization together with a so-called porogen (a solvent which can be washed out after polymerization).

DEPR:

The inventive column may include a number of monolithic chromatographic matrices stacked one upon the other.

CLPR:

1. A liquid chromatography column, comprising a column tube including a chromatographic matrix, the matrix being monolithic and porous, liquid flow inlet means and liquid flow outlet means, a distributor located adjacent the inlet for distributing incoming liquid, and, when eluant passes through the matrix, a liquid permeating gap which is devoid of matrix material, wherein the permeating gap is between the matrix and the distributor and is directly adjacent the matrix, the gap being effective to improve a theoretical plate number, an elution volume or a symmetry factor of the liquid chromatography column.

CCOR:

210/198.2

DOCUMENT-IDENTIFIER: US 5766460 A
TITLE: Liquid chromatographic system

DEPR:

FIGS. 5a and 5b illustrate separation modules which include a separation medium in the form of a matrix (32) and flow channels. The module illustrated in FIG. 5a has an inlet and an outlet through a common end-connecting means, and the module illustrated in FIG. 5b through separate ends. The end pieces of the separation medium have spreading and collecting functions respectively, which can be achieved with the aid of filter paper inserts, end-piece abutment surfaces with inlets combined with systems of channels, etc. (33 and 34 in FIG. 4a). The matrix (32) may be comprised of discrete, packed particles of cross-linked polysaccharide, polyacrylamide, and the like, or may be continuous (monolithic), i.e. have the form of a porous body. The matrix may exhibit substituents enabling the desired type of chromatography to be run (e.g. ion exchange groups, hydrophobic groups, affinity ligands). The width and the length of the matrix are determined by the separation performance desired. The matrix may be given the form of a membrane. It is believed that the future preferred embodiments of the invention will comprise matrixes in form of a continuous body with a cylindrical or frusto conical shape, narrowing in the flow direction of the column (see FIGS. 4a-b).

DEPR:

210/198.2

DOCUMENT-IDENTIFIER: US 5728457 A
TITLE: Porous polymeric material with gradients

ESPR:

Because the plug columns are essentially a single molded polymer monolith traversed by large channels and permeated by small pores, their hydrodynamic properties are excellent. They are unlike any existing separation medium based on packed polymer beads because flow through the plug column does not involve any interstitial space but results entirely from the existence of the through channels built into the porous polymer monolith. Therefore, high rates of mass transfer can be used. Despite the high flow rates and steep gradients, separations using these plug columns are remarkably effective. The continuous polymer plug media afford excellent resolution in the separation of proteins, peptides and small molecules.

CCMR:

210/198.2

DOCUMENT-IDENTIFIER: US 5707589 A

TITLE: Funnel-shaped sample-vial septum with membrane covered diffusion-barrier section

ABFL:

A funnel-shaped monolithic low-density polyethylene sample-vial septum

comprises a flange, a capture wall, a diffusion-barrier wall, and a membrane.

The capture wall is conical, extending from its truncated apex at the diffusion

barrier wall to its mouth about which the flange is disposed.

The membrane is

disposed at the end of the diffusion-barrier wall away from the capture wall.

This structure defines a septum aperture, including a conical capture section,

defined by the capture wall, and a cylindrical diffusion-barrier section,

defined by the diffusion-barrier wall. The 7.0 mm length of the diffusion-barrier section is 12/mm times the square of its diameter 0.76 mm.

The diameter of the diffusion-barrier section which is selected to be 0.05 mm

greater than the diameter of the largest needle to be used with the septum,

i.e., a 22 gauge needle typically used for liquid chromatography.

The minimum

thickness of the membrane is 0.05 mm so that a blunt 26 gauge needle will not

be damaged while piercing the membrane. The membrane has a curved surface

facing the capture section. The radius of curvature of this surface is 7.0 mm,

set equal to the barrier section length. This is a result of a

tip that is curved so that if misaligned, the desired minimum

thickness is still achieved. In fact, however, the membrane never is a full

barrier to evaporative sample loss. After piercing, the long diffusion-barrier

section serves as an effective barrier to evaporative sample loss.

BSPR:

The septum is preferably monolithic and preferably formed of polymer. By "monolithic" is meant that it is fabricated from a single piece of material, as by molding or machining, rather than formed by assembling or fusing separate components. The monolithic structure includes not only the walls of the aperture, but the flange and the membrane as well.

DEPR:

In accordance with the present invention, a monolithic funnel-shaped sample-vial septum A1 comprises a flange 12, a capture wall 14, a diffusion barrier wall 16, and a membrane 18, as shown in FIG. 1. Collectively, these elements define a septum aperture 20 comprising a capture section 22, defined by capture wall 14, and a diffusion-barrier section 24, defined by diffusion barrier wall 16. Capture wall 14 is generally conical with its apex end 30 truncated where it merges with one end 32 of cylindrical diffusion barrier wall 16. The other end 34 of the diffusion barrier wall 16 is sealed by membrane 18, as shown in FIG. 2. The wide end 36 of capture wall 14 is ringed by annular flange 12.

CUXE:

210/198.2

DOCUMENT-IDENTIFIER: US 5653875 A

TITLE: Nucleophilic bodies bonded to siloxane and use thereof for separations from sample matrices

BSPP:

The invention further contemplates a structure comprising discrete adsorbent bodies bonded to a monolithic substrate through a medium comprising a siloxane polymer.

DEPF:

Where the invention is embodied in a chromatographic apparatus or solid phase adsorption device, discrete adsorbent bodies are typically bonded to a monolithic or essentially monolithic substrate, such as an interior wall of a chromatographic column, or a fiber or contiguous network of fibers that support the adsorbent particles of a solid phase adsorption device. As used herein, the term "monolithic" includes essentially monolithic structures such as a weave or mat of contiguous fibers.

DEPE:

In certain applications known to the art, a particulate substrate may provide desirable functions or advantages. It will be understood that, in certain embodiments, the novel chromatographic apparatus or solid phase adsorption device of the invention may comprise a substrate which itself comprises discrete particles, to which discrete adsorbent bodies are bonded with a siloxane polymer. There may also be applications other than in chromatography, sample preparation, or catalysis wherein discrete bodies having a functional surface property are bonded to a monolithic or particulate substrate via the medium of a siloxane polymer.

CCOF:

210/198.2

DOCUMENT-IDENTIFIER: US 5647979 A
TITLE: One-step preparation of separation media for
reversed-phase
chromatography

BSPE:

These disadvantages are eliminated by the formation of a continuous bed in the capillary, i.e., a monolithic porous polymer used in place of the beads, the polymer having been formed by polymerization in the capillary itself, spanning the entire cross section of the capillary and bonded to the capillary wall. A description of this type of bed is found in granted European Patent Specification No. 0 407 560 of Bio-Rad Laboratories, Inc., and its United States counterpart, pending application Ser. No. 08/400,419, filed Mar. 2, 1995. The disclosures of both of these documents are incorporated herein by reference.

CODE:

210/198.2

DOCUMENT-IDENTIFIER: US 5630937 A

TITLE: Nucleophilic bodies bonded to siloxane and use thereof for separations from sample matrices

BSPP:

The invention further contemplates a structure comprising discrete adsorbent bodies bonded to a monolithic substrate through a medium comprising a siloxane polymer.

DEPP:

Where the invention is embodied in a chromatographic apparatus or solid phase adsorption device, discrete adsorbent bodies are typically bonded to a

monolithic or essentially monolithic substrate, such as an interior wall of a chromatographic column, or a fiber or contiguous network of fibers that support the adsorbent particles of a solid phase adsorption device. As used herein, the term "monolithic" includes essentially monolithic structures such as a weave or mat of contiguous fibers.

DEPP:

In certain applications known to the art, a particulate substrate may provide desirable functions or advantages. It will be understood that, in certain embodiments, the novel chromatographic apparatus or solid phase adsorption device of the invention may comprise a substrate which itself comprises discrete bodies, to which discrete adsorbent bodies are bonded with a siloxane polymer. There may also be applications other than in chromatography, separation, or catalysis wherein discrete bodies having a functional surface property are bonded to a monolithic or particulate substrate via the medium of a siloxane polymer.

CLPP:

1. A solid phase adsorption device comprising a monolithic substrate having adsorbent bodies bonded to the surfaces thereof through a medium comprising a siloxane polymer.

CCOR:

210/198.2

DOCUMENT-IDENTIFIER: US 5620603 A

TITLE: Nucleophilic bodies bonded to siloxane and use thereof for separations from sample matrices

BSFR:

The invention further contemplates a structure comprising discrete adsorbent bodies bonded to a monolithic substrate through a medium comprising a siloxane polymer.

DEFF:

Where the invention is embodied in a chromatographic apparatus or solid phase adsorption device, discrete adsorbent bodies are typically bonded to a monolithic or essentially monolithic substrate, such as an interior wall of a chromatographic column, or a fiber or contiguous network of fibers that support the adsorbent particles of a solid phase adsorption device. As used herein, the term "monolithic" includes essentially monolithic structures such as a weave or mat of contiguous fibers.

DEFF:

In certain applications known to the art, a particulate substrate may provide desirable functions or advantages. It will be understood that, in certain embodiments, the novel chromatographic apparatus or solid phase adsorption device of the invention may comprise a substrate which itself comprises discrete bodies, to which discrete adsorbent bodies are bonded with a siloxane polymer. There may also be applications other than in chromatography, sample separation, or catalysis wherein discrete bodies having a functional surface property are bonded to a monolithic or particulate substrate via the medium of a siloxane polymer.

CCXR:

210/198.2

DOCUMENT-IDENTIFIER: US 5620597 A
TITLE: Non-fouling flow-through capacitor

BSFR:

The electrodes may be made out of any monolithic high surface area conductive materials, in at least one anode/cathode pair. Where the high surface area material is conductive, but not optimally conductive, an electrical conductive backing may be employed. High surface area conductive materials suitable for use in the present invention include, but are not limited to: activated carbon; activated carbon treated with a halogen; carbon foams; carbon aerogel and aerogel composite materials; nanotubes; conductive polymers, especially in porous or network form; polymerized fullerenes; or any high surface area conductive material may be used. Conductive ceramics may also be used, either by themselves or impregnated onto high surface area substrates, including various forms of carbon such as fiber, foam, powder or aerogel. In general, absorbing any electrically actuated small or large molecule onto the conductive high surface area material that improves the capacitance will improve the function of the capacitor. Another preferred high surface area conductive material is conductive transition metal oxides, nitrides, or borides prepared using sol/gel technique. Powdered high surface area materials may be sintered into monolithic electrodes or bound together with binder materials.

BSFR:

Intrinsically conductive electrodes where no backing layer is required would include high surface area preparations of graphitic carbon, high surface area expanded metals, metal fibers, or metal meshes. For example, titanium fibers coated with high surface area platinum series black are known and

are marketed as electrode materials. Other examples include platinum coated niobium and foamed metals. High surface area carbon materials may be mixed with metal or graphitic fibers or meshes and formed into monolithic units.

DRFR:

FIGS. 13A-F are schematic illustrations of various monolithic electrodes for use in the capacitor of the invention;

DEPR:

FIGS. 13A-F depict various monolithic electrode designs that incorporate an inner conductive backing. This is useful for all the above flow-through capacitors because a compression fitting is no longer required to make a contact between the high surface area layers and the conductive backing layers. The electrodes of FIG. 13 contain an inner conductive backing layer 2, which may be a metal foil, graphite foil, a fibrous material, or an interpenetrating network mesh material. In foil form, this backing material has many holes 25 therethrough to allow communication and interconnection with the high surface area material that forms a sandwich on both sides in a flat electrode. Alternatively, a rod style conductor can be used, with the high surface area material 1 formed directly around a central rod or wire conductor 31. This material is bonded together or calcined as a single, monolithic piece, containing the conductive backing internally. For example, activated carbon or sugar powder may be mixed with a phenolic binder and hot pressed to form the structure of FIG. 13, prior to calcining in the absence of air. The interconnections formed through the holes in the conductive backing hold the high surface area material together and prevent it from pulling away from the backing due to shrinkage during calcining. Alternatively, carbon films or layers may be deposited onto conductive backings, and activated in place.

Integral leads 4 are formed from the internal conductive backing layer or rod

32.

CLPE:

1. A foul-resistant, flow-through capacitor having at least one anode and cathode pair for use in the electrical purification, concentration, separation, recovery, or electrochemical breakdown of solutes or fluids, which capacitor comprises one or more monolithic, spaced apart pairs of cathode-anode electrodes incorporating a high surface area material and having a non-conductive spacer between the anode and cathode electrodes characterized by an open flow path between the electrodes to permit the unobstructed flow of the fluid across the electrode surface and of sufficient width to prevent the fouling of the capacitor and wherein the open flow path has at least one dimension open to an exterior of the capacitor.

CLPE:

13. The capacitor of claim 1 wherein the monolithic high surface area material is selected from the group consisting of: bonded or sintered activated carbon particles; aerogel particles; conductive ceramics; activated carbon fiber cloth; fibrous metal coated with platinum; or transition metal oxides, borides and nitrides and combinations thereof.

CLPE:

14. The capacitor of claim 1 wherein the monolithic high surface area material comprises activated carbon sintered together with a binder and doped with a metal.

CLPE:

210/198.2

DOCUMENT-IDENTIFIER: US 5609756 A

TITLE: Nucleophilic bodies bonded to siloxane and use thereof for separations from sample matrices

BSPP:

The invention further contemplates a structure comprising discrete adsorbent bodies bonded to a monolithic substrate through a medium comprising a siloxane polymer.

DEPR:

Where the invention is embodied in a chromatographic apparatus or solid phase adsorption device, discrete adsorbent bodies are typically bonded to a

monolithic or essentially monolithic substrate, such as an interior wall of a chromatographic column, or a fiber or contiguous network of fibers that support the adsorbent particles of a solid phase adsorption device. As used herein, the term "monolithic" includes essentially monolithic structures such as a weave or mat of contiguous fibers.

DEPR:

In certain applications known to the art, a particulate substrate may provide desirable functions or advantages. It will be understood that, in certain embodiments, the novel chromatographic apparatus or solid phase adsorption device of the invention may comprise a substrate which itself comprises discrete bodies, to which discrete adsorbent bodies are bonded with a siloxane polymer. There may also be applications other than in chromatography, sample preparation, or catalysis wherein discrete bodies having a functional surface property are bonded to a monolithic or particulate substrate via the medium of a siloxane polymer.

CCOR:

210/198.2

DOCUMENT-IDENTIFIER: US 5607580 A

TITLE: Nucleophilic bodies bonded to siloxane and use thereof for separations from sample matrices

BSPP:

The invention further contemplates a structure comprising discrete adsorbent bodies bonded to a monolithic substrate through a medium comprising a siloxane polymer.

DEPR:

Where the invention is embodied in a chromatographic apparatus or solid phase adsorption device, discrete adsorbent bodies are typically bonded to a monolithic or essentially monolithic substrate, such as an interior wall of a chromatographic column, or a fiber or contiguous network of fibers that support the adsorbent particles of a solid phase adsorption device. As used herein, the term "monolithic" includes essentially monolithic structures such as a weave or mat of contiguous fibers.

DEPR:

In certain applications known to the art, a particulate substrate may provide desirable functions or advantages. It will be understood that, in certain embodiments, the novel chromatographic apparatus or solid phase adsorption device of the invention may comprise a substrate which itself comprises discrete bodies, to which discrete adsorbent bodies are bonded with a siloxane polymer. There may also be applications other than in chromatography, sample preparation, or catalysis wherein discrete bodies having a functional surface property are bonded to a monolithic or particulate substrate via the medium of a siloxane polymer.

CCOR:

210/198.2

DOCUMENT-IDENTIFIER: US 5599445 A

TITLE: Nucleophilic bodies bonded to siloxane and use thereof for separations from sample matrices

BSPP:

The invention further contemplates a structure comprising discrete adsorbent bodies bonded to a monolithic substrate through a medium comprising a siloxane polymer.

DEPR:

Where the invention is embodied in a chromatographic apparatus or solid phase adsorption device, discrete adsorbent bodies are typically bonded to a

monolithic or essentially monolithic substrate, such as an interior wall of a chromatographic column, or a fiber or contiguous network of fibers that support the adsorbent particles of a solid phase adsorption device. As used herein, the term "monolithic" includes essentially monolithic structures such as a weave or mat of contiguous fibers.

DEPR:

In certain applications known to the art, a particulate substrate may provide desirable functions or advantages. It will be understood that, in certain embodiments, the novel chromatographic apparatus or solid phase adsorption

device of the invention may comprise a substrate which itself comprises

discrete bodies, to which discrete adsorbent bodies are bonded with a siloxane

polymer. There may also be applications other than in chromatography, sample

preparation, or catalysis wherein discrete bodies having a functional surface

property are bonded to a monolithic or particulate substrate via the medium of a siloxane polymer.

CLPR:

1. A structure comprising discrete adsorbent bodies bonded to a monolithic substrate through a medium comprising a polysiloxane polymer.

CCOR:

210/198.2

DOCUMENT-IDENTIFIER: US 5308495 A

TITLE: Chromatography processes using doped sol gel glasses as chromatographic media

BSPP:

B. Chromatographic Applications: The selective interaction of doped sol-gel glass with the surrounding chemical compounds makes this type of glasses a promising chromatographic media. This possibility was specifically mentioned in the above mentioned patent application. In examples E-F we demonstrate planar and column chromatographies for liquid and gas applications. Monolithic doped sol-gel glasses can also be used for the same purpose.

SEPP:

FIG. 5 discloses monolithic glass detectors doped with ionic surface active agents to prevent cracking. Upper section: detectors before and after immersion in solution containing the analyzates; lower row: monolithic glass disks.

DEPP:

FIG. 5 depicts several typical photometric detectors containing doped surface active agents. The first upper section contains four examples of monolithic doped glasses before and after immersion in aqueous solution containing the analyzate (from left to right: Redox detector containing Diphenylaminesulfonate 1.1 mg salt (1 mg) and 5 mg cetylpyridinium bromide (CPB) before and after immersion in sodium hypochlorite solution; pH indicator containing phenolphthalein (2.5 mg) and 6.6 mg cetyltrimethylammonium bromide (CTAB) before and after immersion in basic aqueous solution; iron Detector containing 1.2 mg o-phenanthroline and 5.0 mg CPB before and after immersion in a solution containing ferrous ammonium sulfate; nitrite indicator containing

4 mg

1-Naphthylenediamine dihydrochloride, 10 mg sulphanilic acid and 6 mg CTAB).

The lower row depicts few detectors (from left to right; copper detector

containing 1 mg dithiooxamide and 6.25 mg CPB; lead detector containing 1 mg

galocyanine and 7.5 mg CPB; pH detector containing 5 mg bromophenol and 6.6 mg

CTAB; nickel detector containing 2 mg dimethyglyoxime and 6.25 mg CPB; aluminum

detector containing 5 mg alizarin and 12.5 mg CPB)

DEPR:

In all cases the described procedure prevented cracking of the monolithic glasses even after few cycles of wetting and drying of the glasses.

CCXR:

210/198.2

DOCUMENT-IDENTIFIER: US 5141609 A

TITLE: Method and device employing time-delayed integration for detecting sample components after separation

DEPR:

Charge-Coupled Device. A solid state device comprising an array of detectors which can be readily adapted for use in the present invention is the charge-coupled device (CCD). A CCD is a monolithic large-format silicon array detector. Characteristics that make it ideally suited for detection are extremely high quantum efficiency (.ltoreq.80%), virtually no dark current, and up to 10.sup.6 individual detector elements in the array. The CCD is conceptually similar to an electronic photographic film in that both integrate signal information. The integrating ability of the CCD and lack of dark current allow the CCD to perform exceptionally well in situations where several seconds are allowed for integration of the signal. In microcolumn separations, detection zones can be constructed such that analyte bands are viewed for many seconds--an ideal measurement task for a CCD.

CCKE:

210/198.2

DOCUMENT-IDENTIFIER: US 4954149 A
TITLE: Injection septum

DEPR:

In addition to the embodiments described above, the present invention provides for a monolithic septum with annular and duckbill ends of a unitary aperture. Such a septum differs from conventional and other known duckbill seals in that, when a needle is extended through the septum, the annular aperture and the duckbill aperture expand elastically to accommodate the needle and press against the needle under the pressure of their deformation. A wide variety of elastically deformable materials can be employed in the septum provided by the present invention. In addition, interlocking of components of a compound septum can be effected using a variety of means. Dimensions can be changed to accommodate devices and applications other than those illustrated.

CCXR:

210/198.2

DOCUMENT-IDENTIFIER: US 4792395 A

TITLE: High speed countercurrent centrifuge for removal attachment of chromatographic columns thereto, and chromatographic column for the same

DEPR:

FIG. 4 shows a variation of the column holder arrangement used in the present invention. Elements 5' 10' 11', 12', 13', 14', 14a', 16', 18', 19', 20', 21', 22' and 54' correspond, respectively, to the corresponding elements 10, 11, 12, 13, 14, 14a, 16, 18-22 and 54 in the embodiment of FIGS. 1-3 and will not be further described in detail. In this embodiment, a sleeve 62 is fitted around central shaft 5'. Thus, flow tubes 54' can extend upwardly through a central bore in shaft 5', and out through exits 60 and 61, finally connecting to top flange 20' and tubing adaptors 16'. Quick release nuts 12' and 13' are screw threaded about sleeve 62. Sleeve 62 is fixed to or is monolithic with column holder 10'. The embodiment of FIG. 4, which places the tubing connections 16' at the top of spool 19', permits facile inspection of the flow tubes 54' and their connections. Sleeve 62 is necessary to create space 62a so that the column 18' may be slid off shaft 5' without hindrance by the portion of tubes 54' extending through opening 60.

COOR:

210/198.2

DOCUMENT-IDENTIFIER: US 4636315 A
TITLE: Fluid separator apparatus and method

DEPR:

While the distributor body has been illustrated as comprising a pair of plates, with the channels conveniently formed at the plates' mating surfaces, it will be appreciated that the distributor body may be formed in other fashions. For example, the distributor body may be formed as a monolithic structure with the fluid passages formed therein to communicate between the fluid port and the uniformly spaced distribution openings. The fluid passages on such a monolithic structure may be formed by casting or etching techniques known in the art.

CCXR:

210/198.2

DOCUMENT-IDENTIFIER: US 4551251 A
TITLE: Monolithic integrated flow circuit

TTL:
Monolithic integrated flow circuit

ABPL:
A monolithic multi-channel integrated flow circuit comprising a support matrix sheet or plate impressed or embossed with the desired circuitry; the desired circuit elements such as transfer conduit and separation columns are integral with and defined by the support matrix, which conveniently comprises a first deformable support sheet embossed with the circuits elements by thermoforming techniques, and bonded to a support blank or correspondingly embossed second support sheet to complete and define the circuit.

BSPE:
The invention relates to monolithic integrated flow circuits useful in a variety of analytical and preparative applications, and particularly relates to chromatographic flow circuits adaptable to a broad spectrum of known chromatographic separation and concentration processes, including liquid partition chromatography, liquid-solid chromatography, ion-exchange chromatography, gas-liquid chromatography, and gas-solid chromatography.

BSPE:
The invention comprises a monolithic multi-channel integrated flow circuit MIFC for circulating fluids. For partitioning fluids according to conventional chromatographic principles, the flow circuit of the invention includes a plurality of separation columns or channels interconnected in series by integrated narrower-bore transfer conduits for delivering fluid from one column to a next successive column. The MIFC of the invention

comprises a support matrix or sheet impressed with the desired circuitry by appropriate molding or machining techniques; the circuitry is thus integral with and at least partially defined by the support matrix. Conveniently, the circuit is fabricated by embossing the circuit in a thin-gauge metal or plastic support sheet, followed by bonding of the embossed sheet to a support sheet blank to define the circuit, according to well-known thermoforming techniques, such as those used for blister packaging. Very complicated integrated flow circuits can thus readily be produced according to desired pattern in a compact sheet without prior art adaptors, transfer tubing, joints, and other discrete elements requiring assembly and structural integrity. Either flexible matrix sheets, or thicker, more rigid matrix sheets (herein referred to as matrix "plates") may be employed.

DEPR:

According to the present invention, many complex flow patterns are obtainable from plastic support sheets or similar matrix material molded to define the

monolithic integrated flow circuits (MIFC) herein described.

Characteristic

features of MIFC are as follows:

CLPR:

1. A monolithic integrated flow circuit consisting essentially of a deformable matrix plate having a flow circuit formed therein, said deformable matrix plate being of formed plastic or metallic thin-gauge sheets, at least one of said sheets having a pattern formed by embossing thereon, said sheets being bonded together to form said flow pattern, said pattern defining a plurality of locules, each of said locules being joined to an adjacent locule by an integral transfer conduit.

CLPR:

6. A plurality of monolithic flow circuits according to claim 1, wherein the flow circuits formed in said plates are interconnected in series.

CLPP:

8. A plurality of monolithic integrated flow circuits according to claim 1, wherein a plurality of said plates are formed into a block of circuits interconnected in series.

CLPP:

12. The monolithic integrated flow circuit of claim 1, wherein said support sheets are thin-gauge plastic.

CLPP:

13. The monolithic integrated flow circuit of claim 1, wherein said support sheets are thin-gauge metal.

CLPP:

14. The monolithic integrated flow circuit of claim 1, wherein the flow circuit is embossed in a thermoplastic matrix plate by thermoforming.

CLPP:

16. A method for counter-current chromatography comprising separating a first liquid from a second liquid with which said first liquid is mixed by passing a mixture of said first and second liquid through a monolithic integrated flow circuit consisting essentially of a deformable matrix plate having a flow circuit formed therein, said deformable matrix plate being of formed plastic or metallic thin-gauge sheets, at least one of said sheets having a pattern formed by embossing thereon, said sheets being bonded together at said pattern, said pattern defining a plurality of locules, each of said locules being joined to an adjacent locule by an integral transfer conduit and subjecting said mixture to counter-current chromatography in said column.

CCXR:

210/198.2

DOCUMENT-IDENTIFIER: US 4440638 A

TITLE: Surface field-effect device for manipulation of charged species

DEPR:

Dielectric film 96 is suitably a layer of glass. The glass layer serves to

hold the top portion of the capacitance device in juxtaposition with the lower

portion 100. The glass layer can be formed using conventional sintered infused

glass techniques employed in hybrid circuit processing. The capacitive device

and measurement system may suitably be in accordance with that disclosed in

Sander et al., "MONOLITHIC CAPACITANCE PRESSURE TRANSDUCER-IC WITH PULSE PERIOD

OUTPUT", IEEE/Engineering in Medicine and Biology Conference on Frontiers of

Engineering and Health Care, 1979, pages 189-192.

CCXR:

210/198.2

DOCUMENT-IDENTIFIER: US 4175037 A
TITLE: Process for packing chromatographic columns

DEPR:

FIG. 2 is a schematic diagram illustrating an apparatus for effecting the method of the present invention used in conjunction with a monolithic glass chromatographic column.

DEPR:

Referring to FIG. 2, another embodiment of the present invention is shown wherein a monolithic glass chromatographic column 32 is shown being packed in accordance with the method of the present invention. The monolithic glass chromatographic column has a continuous passageway formed therethrough which can be packed with chromatographic packing material in accordance with the present invention. Quick-connect and disconnect couplings 34 and 36 enable the column to be connected and disconnected from the packing system of the present invention. Moreover, these couplings enable the packed chromatographic column to be readily connected to gas feed and analytical devices.

CCXR:

210/198.2

DOCUMENT-IDENTIFIER: US 4116836 A
TITLE: Chromatographic column

ABPL:

A monolithic glass construction having a continuous passageway formed therethrough packed with chromatographic packing material is used as a chromatographic column. The monolithic construction enables the column to be formed of the longest practical length and smallest practical diameter to increase the efficiency of the column, while also providing a durable column. Quick-connect and disconnect couplings to a gas feed and analysis device are mounted on the monolithic construction.

BSPP:

In accordance with one form of the invention, the chromatographic column consists of a monolithic glass cylinder having a pair of opposed surfaces and a side wall in which a continuous helical passageway is formed. The passageway is packed with gas absorptive material for performing the chromatographic process. Gas can be injected directly into the helical passageway and a pair of metal flanges can be inserted over the opposed surface comprising the top and bottom edges of the glass cylinder.

BSPP:

The flanges can be formed from metal to preclude chipping and breaking of the monolithic glass construction and to further serve as a conductor to discharge static electricity which may adhere to the interior passageway in the monolithic structure. The couplings on the flange also provide a firm means of support for making the connections to the valve structure.

BSPP:

The monolithic glass construction may also take the form of a number of stacked

plates fused together which have continuous interconnected passages of sinusoidal or other configuration formed therethrough. The passages are also packed with gas absorptive or adsorptive material.

ESPR:

By using such monolithic plates or cylinders, the height of the resulting column can be constructed at will with any diameter passage within practical limits, serving to effect a chromatographic column of high efficiency. Further, the resultant column because of its monolithic (or solid one piece) construction is extremely durable.

BSPR:

The cylindrical column can be formed by wrapping an expendable, sacrificial tubing in a coil or helix of appropriate spacing and length around a section of glass tubing supported on a mandrel. It is considered preferable to first cut, grind, etch or mold the glass tubing in the desired helical configuration or other desired configuration and to wrap the sacrificial tubing in the resulting grooved path to thereby establish and retain the desired spacing between the coils of tubing during subsequent processing. A second tube is then placed over the coil so a concentric sandwich is formed on the mandrel. Sufficient heat is applied to one end of the inner and outer tubes to effect fusing of the tubes at said one end thereof. A vacuum is drawn between the inner and outer tubes and heat is applied to the tubes, causing said tubes to flow into the interspatial areas between the sacrificial tubing, thereby fusing the glass tubes together and forming a monolithic structure. When this operation is complete, the sacrificial tubing is removed by means of an etchant or other reactant chemical compound such as FeCl_3 , HCl , an acetic acid-nitric acid mixture and the like which creates a continuous passageway of the desired

configuration through the monolithic structure. The resultant column is monolithic (or solid one piece) in structure having the integrity of a single heavy wall glass tube. The continuous passageway is then loaded with chromatographic packing thereby forming a monolithic packed chromatographic column.

BSPR:

In the stacked plate construction, sacrificial tubing may be placed in grooves cut, ground, etched or molded in a desired continuous passageway configuration on one of the two opposing surfaces of a glass plate and a second plate fused thereto. After removal of the sacrificial tubing, a monolithic plate having a continuous passageway formed therein is obtained. A plurality of such monolithic plates may be stacked one on top of the other with the outlet of one joined to the inlet of the next adjacent plate such as with a glass or metal U tube. The resulting continuous passageway through the stack of monolithic plates can be packed with chromatographic packing material to create a packed chromatographic column of significant strength and resistance to breakage. Moreover, a column obtained in this fashion is surprisingly compact and efficient.

DEPR:

Column 10 includes a monolithic glass cylindrical tube 12 having a helical passageway 14 formed in a side wall 16 thereof. One end of passageway 14 opens at 18 in the bottom edge 18 of cylinder 12 and the other end of passageway 14 has an opening 20 in the top edge 20 of cylinder 12.

DEPR:

Because of the nature of the monolithic construction of column 10, durable columns as long as 200 feet or more may be manufactured having a small diameter passageway 14 whereby the efficiency of the column may be

substantially increased. For example, columns having more than 2,000 theoretical plates per foot have been fabricated with the construction illustrated in FIGS. 1 to 3 wherein heretofore it was only possible to fabricate columns of about 800 theoretical plates per foot.

DEPR:

Column 10 is fabricated in a monolithic construction by wrapping an expendable, sacrificial tubing in a helix of a length and width spacing between the coils of the helix as desired around an inner glass tubing section supported on a mandrel. It is considered preferable to cut, grind, etch or mold a grooved path on the outer surface of the inner glass tubing to serve as a guide and retainer for the sacrificial tubing. In this manner, the spacing between the wire can be preset and retained during the heating operation. A second outer section of glass tubing is placed over the coil and first tube to form a concentric sandwich on the supporting mandrel. Sufficient heat is applied to one end of the sandwich to effect fusing of the glass tubes at said one end thereof. A vacuum is then drawn between the inner and outer glass tubing to fuse and flow into the interspacial areas between the sacrificial tubing. When this operation is complete, the sacrificial tubing is removed by means of an acid etchant or other reactant compound which leaves the desired configuration for the column. A chromatographically chemically inert porous retainer is inserted into a terminal portion of the continuous passageway between the column. Such retainers can be formed of fiber glass, glass wool, glass frits, an inert metal filter or wool such as gold wool and the like. Such retainers can also be interposed in the terminal portion of the inlet and outlet lines leading to and from the column to further

protect against loss of packing material. The passageway thus formed by removal of the sacrificial tubing is packed with an appropriate adsorption-desorption material and a porous retainer as described hereinabove can be inserted into the initial portion of the continuous passageway of the column thereby sealing the packing within the passageway of the column. The resultant structure is monolithic in nature having the integrity of a single heavy wall glass tube of desired length. Moreover, the precision bore of the passageway obtained through use of the sacrificial tubing is believed to help in obtaining packing uniformity.

DEPR:

As shown in FIGS. 7 to 9, inclusive, an annular metal flange 24 may be seated on the bottom and top edges 18 and 22 of cylindrical column 12, respectively. The metal flanges 24 preclude chipping of the monolithic glass construction of column 12 and can also serve to aid in the elimination of static charges carried by the sample under analysis in passageway 14.

DEPR:

As shown in FIGS. 4 to 6, inclusive, the chromatographic column 10 can be formed with both the inlet and outlet for the gas sample in either the top or bottom edges 18 or 22 of the glass monolithic cylinder 12. As shown, the inlet 18 and outlet 20 can be disposed on opposite diametrical portions of the top edge 22 of cylinder 12. A flange 24 can be disposed over the top edge as well as the bottom edge, with a quick-disconnect and connect cylindrical coupling 26 at each of the inlet 18 and outlet 20. Accordingly, this monolithic construction of chromatographic tube 10 permits convenience for attachment for suitable analysis equipment.

CLPR:

1. A packed chromatographic column comprising a glass monolithic construction

having at least one pair of opposed surfaces and solid glass therebetween;

CLPF:

3. The packed chromatographic column of claim 2 wherein said glass monolithic construction is the side wall of a cylindrical tube, said side wall containing said continuous passageway.

CLPF:

11. The packed chromatographic column of claim 1 wherein said glass monolithic construction includes at least one substantially planar plate.

CLPF:

16. The packed chromatographic column of claim 15 wherein said glass monolithic construction is the side wall of a cylindrical tube, said side wall containing said continuous passageway.

CLPV:

A continuous passageway of essentially constant diameter adapted to receive a fluid therethrough contained wholly within said solid glass between said opposed surfaces of said monolithic construction, the passageway having an inlet and an outlet opening in at least one of said opposed surfaces, and

CCOR:

210/198.2

DOCUMENT-IDENTIFIER: US 5833861 A
TITLE: Perfusive chromatography

DEPR:

Broadly, in accordance with the invention, perfusion chromatography is practiced by passing fluids at velocities above a threshold level through a specially designed matrix characterized by a geometry which is bimodal or multimodal with respect to its porosity. Perhaps the most fundamental observation relevant to the new procedure is that it is possible to avoid both the loss of capacity characteristic of convection bound systems and the high plate height and bandspreading characteristics of diffusion bound systems. This can be accomplished by forcing chromatography fluids through a matrix having a set of larger pores, such as are defined by the interstices among a bed of particles, and which determine pressure drops and fluid flow velocities through the bed, and a set of pores of smaller diameter, e.g., anisotropic throughpores. The smaller pores permeate the individual particles and serve to deliver chromatography fluids by convection to surface regions within the particle interactive with the solutes in the chromatography fluid.

DEPS:

From the foregoing description many of the basic engineering goals to be pursued in the fabrication of matrix materials suitable for the practice of perfusive chromatography will be apparent to those skilled in the art. Thus, what is needed to practice perfusion chromatography is a matrix which will not crush under pressure having a bimodal or preferably multimodal pore structure and as large a surface area per unit volume as possible. The first and second pore sets which give the material its bimodal flow properties

must have mean diameters relative to each other so as to permit convective flow through both sets of pores at high V.sub.beds. The provision of subpores in the matrix is not required to conduct perfusion chromatography but is preferred because of the inherent increase in surface area per unit volume of matrix material such a construction provides.

DEPR:

In contrast to the PL 4,000 material, which, with respect to its pore structure, is multimodal, a more ideal perfusive particle might comprise a plurality of sets of throughpores and subpores of differing mean diameters. A bimodal pore size distribution can be achieved in such particle by mixing equal ratios of particles having two discrete pore sizes or by engineering this feature at the polymerization stage. Ideally, mean diameter ratio between throughpore subsets would be less than 10, the mean diameter ratio between the smallest throughpore sets and the subpores would be less than 20, and the mean diameter ratio between the first pore set, i.e., the interstices among the particles making up the matrix, and the largest throughpore subset would be less than 70, and preferably less than 50. A multimodal material might be produced by agglomerating 500 .ANG. porons to form approximately 1 .mu.m clusters, which in turn are agglomerated to form 10 .mu.m aggregates, which in turn may be aggregated to form 100 .mu.m particles. In such a design, the 1 .mu.m clusters would have interstices of a mean diameter in the vicinity of a few hundred .ANG.. These clusters would provide a large surface area. Diffusive transport within these pores would rarely have to exceed a distance of 0.5 .mu.m or 5,000 .ANG.. Interstices among the 1 .mu.m clusters making up the 10 .mu.m aggregates would permit convective flow to feed

the diffusive pores. These would be on the order of 0.3 μm in diameter.

These throughpores, in turn would be fed by larger pores defined by

interstices among the 10 μm particles making up the 100 μm particles.

These would have a mean diameter on the order of 35 μm .

CCXR:

210/198.2

DOCUMENT-IDENTIFIER: US 5605623 A
TITLE: Perfusive chromatography

DEPR:

Broadly, in accordance with the invention, perfusion chromatography is practiced by passing fluids at velocities above a threshold level through a specially designed matrix characterized by a geometry which is bimodal or multimodal with respect to its porosity. Perhaps the most fundamental observation relevant to the new procedure is that it is possible to avoid both the loss of capacity characteristic of convection bound systems and the high plate height and bandspreading characteristics of diffusion bound systems. This can be accomplished by forcing chromatography fluids through a matrix having a set of larger pores, such as are defined by the interstices among a bed of particles, and which determine pressure drops and fluid flow velocities through the bed, and a set of pores of smaller diameter, e.g., anisotropic throughpores. The smaller pores permeate the individual particles and serve to deliver chromatography fluids by convection to surface regions within the particle interactive with the solutes in the chromatography fluid.

DEPR:

From the foregoing description many of the basic engineering goals to be pursued in the fabrication of matrix materials suitable for the practice of perfusion chromatography will be apparent to those skilled in the art. Thus, what is needed to practice perfusion chromatography is a matrix which will not crush under pressure having a bimodal or preferably multimodal pore structure and as large a surface area per unit volume as possible. The first and second pore sets which give the material its bimodal flow properties

must have mean diameters relative to each other so as to permit convective flow through both sets of pores at high V.sub.beds. The provision of subpores in the matrix is not required to conduct perfusion chromatography but is preferred because of the inherent increase in surface area per unit volume of matrix material such a construction provides.

DEPR:

In contrast to the PL 4,000 material, which, with respect to its pore structure, is multimodal, a more ideal perfusive particle might comprise a plurality of sets of throughpores and subpores of differing mean diameters. A bimodal pore size distribution can be achieved in such particle by mixing equal ratios of particles having two discrete pore sizes or by engineering this feature at the polymerization stage. Ideally, mean diameter ratio between throughpore subsets would be less than 10, the mean diameter ratio between the smallest throughpore sets and the subpores would be less than 20, and the mean diameter ratio between the first pore set, i.e., the interstices among the particles making up the matrix, and the largest throughpore subset would be less than 70, and preferably less than 50. A multimodal material might be produced by agglomerating 500 .ANG. pores to form approximately 1 .mu.m clusters, which in turn are agglomerated to form 10 .mu.m aggregates, which in turn may be aggregated to form 100 .mu.m particles. In such a system, the 1 .mu.m clusters would have interstices of a mean diameter in the vicinity of a few hundred .ANG.. These clusters would be separated by a large surface area. Diffusive transport within these pores would rarely have to exceed a distance of 0.5 .mu.m or 5,000 .ANG.. Interstices among the 1 .mu.m clusters making up the 10 .mu.m aggregates would permit convective flow to feed

the diffusive pores. These would be on the order of 0.3 .mu.m in diameter.

These throughpores, in turn would be fed by larger pores defined by interstices among the 10 .mu.m particles making up the 100 .mu.m particles.

These would have a mean diameter on the order of 35 .mu.m.

CLPE:

8. The chromatography system of claim 7 wherein the packed particles define a bimodal or multimodal pore structure.

CCOR:

210/198.2

DOCUMENT-IDENTIFIER: US 5552041 A
TITLE: Perfusive chromatography

DEPR:

Broadly, in accordance with the invention, perfusion chromatography is practiced by passing fluids at velocities above a threshold level through a specially designed matrix characterized by a geometry which is bimodal or multimodal with respect to its porosity. Perhaps the most fundamental observation relevant to the new procedure is that it is possible to avoid both the loss of capacity characteristic of convection bound systems and the high plate height and bandspreading characteristics of diffusion bound systems. This can be accomplished by forcing chromatography fluids through a matrix having a set of larger pores, such as are defined by the interstices among a bed of particles, and which determine pressure drops and fluid flow velocities through the bed, and a set of pores of smaller diameter, e.g., anisotropic throughpores. The smaller pores permeate the individual particles and serve to deliver chromatography fluids by convection to surface regions within the particle interactive with the solutes in the chromatography fluid.

DEPR:

From the foregoing description many of the basic engineering goals to be achieved in the fabrication of matrix materials suitable for the practice of perfusion chromatography will be apparent to those skilled in the art. Thus, what is needed to practice perfusion chromatography is a matrix which will not crush under pressure having a bimodal or preferably multimodal pore structure and as large a surface area per unit volume as possible. The first and second pore sets which give the material its bimodal flow properties

must have mean diameters relative to each other so as to permit convective flow through both sets of pores at high V.sub.beds. The provision of subpores in the matrix is not required to conduct perfusion chromatography but is preferred because of the inherent increase in surface area per unit volume of matrix material such a construction provides.

DEPR:

In contrast to the PL 4,000 material, which, with respect to its pore structure, is multimodal, a more ideal perfusive particle might comprise a plurality of sets of throughpores and subpores of differing mean diameters. A bimodal pore size distribution can be achieved in such particle by mixing equal ratios of particles having two discrete pore sizes or by engineering this feature at the polymerization stage. Ideally, mean diameter ratio between throughpore subsets would be less than 10, the mean diameter ratio between the smallest throughpore sets and the subpores would be less than 20, and the mean diameter ratio between the first pore set, i.e., the interstices among the particles making up the matrix, and the largest throughpore subset would be less than 70, and preferably less than 50. A multimodal material might be produced by agglomerating 500 .ANG. porons to form approximately 1 .mu.m clusters, which in turn are agglomerated to form 10 .mu.m aggregates, which in turn may be aggregated to form 100 .mu.m particles. In such a system, the 1 .mu.m clusters would have interstices of a mean diameter in the vicinity of a few hundred .ANG.. These would afford the largest and provide a large surface area. Diffusive transport within these pores would rarely have to exceed a distance of 0.5 .mu.m or 5,000 .ANG.. Interstices among the 1 .mu.m clusters making up the 10 .mu.m aggregates would permit convective flow to feed

the diffusive pores. These would be on the order of 0.3 μm in diameter.

These throughpores, in turn would be fed by larger pores defined by interstices among the 10 μm particles making up the 100 μm particles.

These would have a mean diameter on the order of 35 μm .

CCOR:

210/198.2

DOCUMENT-IDENTIFIER: US 5540834 A
TITLE: Synthesis of porous inorganic particles by
polymerization-induced
colloid aggregation (FICA)

DEPR:

The specific surface area and porosity of the sintered particles measured by nitrogen adsorption were $13 \text{ m}^2/\text{g}$ and 29%, respectively, which are in reasonable agreement with close-packed, dense ZrO_2 spheres with nonuniform particle size. The pore size distributions (psd) obtained by nitrogen adsorption and desorption, and mercury porosimetry (intrusion) are displayed in FIGS. 4A, 4B and FIG. 5, respectively. From N_2 adsorption, the psd was determined to be rather narrow with a maximum near 400 Å and a small contribution of pores larger than 500 Å and smaller than 100 Å. From N_2 desorption, the psd was determined to be multimodal with nearly all pores between 100 Å and 200 Å in diameter and some pores below 50 Å. This discrepancy is mainly due to pore blocking or network effects, whereby desorption from a pore in a network is influenced by the state of the neighboring pores. Nitrogen adsorption probes the main channel size and can be considered free of pore blocking effects, while nitrogen desorption shows a disproportionately large amount of small pores due to "bottle necks." The psd obtained from mercury porosimetry (intrusion) are also influenced by pore interconnections; as shown in FIG. 5, it is broad with pores between approximately 100 Å and 300 Å in diameter and a maximum near 200 Å, in reasonable agreement with the psd from N_2 desorption.

DEPR:

The pore-size distributions (psd's) after sintering are shown in

FIG. 19. All samples exhibited multimodal psd's with pore diameters ranging between 100 .ANG. and 450 .ANG.. The sample synthesized at pH 1.2 contained some hollow particles but its psd seemed qualitatively similar to those of samples synthesized at higher pH (non-hollow particles). This is because N.sub.2 adsorption only probes the pores within the ZrO.sub.2 shells and not the large voids they encompass. Surface areas and porosities (.epsilon..sub.particle) for these samples are listed in Table 3. Note that there is qualitative agreement with FIG. 12E, but quantitative agreement is not expected since condensing N.sub.2 cannot be used to distinguish between hollow cores and interstitial volume between aggregates. The N.sub.2 psd's show that the higher the pH, the greater contribution of small pores to the total porosity.

CCOE:
210/198.2

DOCUMENT-IDENTIFIER: US 5522994 A
TITLE: Single column chromatographic determination of small
molecules in
mixtures with large molecules

CCXR:
210/198.2

ORPL:
Little, "Sequential Multimodal Elution for Pseudomultidimensional
Liquid
Chromatography on a Single Column," Anal. Chem., 63 (1991) pp.
33-34.

DOCUMENT-IDENTIFIER: US 5431807 A

TITLE: **Multimodal** chromatographic separation media and process for using same

TTL:

Multimodal chromatographic separation media and process for using same

ABPL:

A process for carrying out in a consecutive fashion different modes of chromatographic separation in a liquid chromatography column using a single separation medium is disclosed. Separation media for use in such **multimodal** separations are also disclosed.

BSPR:

It may be possible to use combinations of different separation media in different columns for **multimodal** separations. An example of this multiple column bimodal separation was described recently by Wheatley J. B., J. Chromatogr., 603 (1992) 273. The bimodal separation of small molecules in one column packed with one separation medium and based on sequential **multimodal** elution was described by Little E. L., Jeansonne M. S., Foley J. P.; Anal Chem., 63, 1991, 33. They combined ion-exchange and reversed phase chromatography for the separation of a complex sample containing two groups of compounds: charged and non-polar. The use of two different gradients, i.e. a pH gradient and a methanol gradient, resulted in the separation of the charged molecules first, followed by the separ

DOCUMENT-IDENTIFIER: US 5833861 A
TITLE: Perfusive chromatography

DEPR:

Broadly, in accordance with the invention, perfusion chromatography is practiced by passing fluids at velocities above a threshold level through a specially designed matrix characterized by a geometry which is bimodal or multimodal with respect to its porosity. Perhaps the most fundamental observation relevant to the new procedure is that it is possible to avoid both the loss of capacity characteristic of convection bound systems and the high plate height and bandspreading characteristics of diffusion bound systems. This can be accomplished by forcing chromatography fluids through a matrix having a set of larger pores, such as are defined by the interstices among a bed of particles, and which determine pressure drops and fluid flow velocities through the bed, and a set of pores of smaller diameter, e.g., anisotropic throughpores. The smaller pores permeate the individual particles and serve to deliver chromatography fluids by convection to surface regions within the particle interactive with the solutes in the chromatography fluid.

DEPR:

From the foregoing description many of the basic engineering goals to be pursued in the fabrication of matrix materials suitable for the practice of perfusion chromatography will be apparent to those skilled in the art. Thus, what is needed to practice perfusion chromatography is a matrix which will not crush under pressure having a bimodal or preferably multimodal pore structure and as large a surface area per unit volume as possible. The first and second pore sets which give the material its bimodal flow properties

must have mean diameters relative to each other so as to permit convective flow through both sets of pores at high V.sub.beds. The provision of subpores in the matrix is not required to conduct perfusion chromatography but is preferred because of the inherent increase in surface area per unit volume of matrix material such a construction provides.

DEPR:

In contrast to the PL 4,000 material, which, with respect to its pore structure, is multimodal, a more ideal perfusive particle might comprise a plurality of sets of throughpores and subpores of differing mean diameters. A bimodal pore size distribution can be achieved in such particle by mixing equal ratios of particles having two discrete pore sizes or by engineering this feature at the polymerization stage. Ideally, mean diameter ratio between throughpore subsets would be less than 10, the mean diameter ratio between the smallest throughpore sets and the subpores would be less than 20, and the mean diameter ratio between the first pore set, i.e., the interstices among the particles making up the matrix, and the largest throughpore subset would be less than 70, and preferably less than 50. A multimodal material might be produced by agglomerating 500 .ANG. perons to form approximately 1 .mu.m clusters, which in turn are agglomerated to form 10 .mu.m aggregates, which in turn may be aggregated to form 100 .mu.m particles. In such a design, the 1 .mu.m clusters would have interstices of a mean diameter in the vicinity of a few hundred .ANG.. These would serve the subpores and provide a very high surface area. Diffusive transport within these pores would rarely have to exceed a distance of 0.5 .mu.m or 5,000 .ANG.. Interstices among the 1 .mu.m clusters making up the 10 .mu.m aggregates would permit convective flow to feed

the diffusive pores. These would be on the order of 0.3 μm in diameter.

These throughpores, in turn would be fed by larger pores defined by interstices among the 10 μm particles making up the 100 μm particles.

These would have a mean diameter on the order of 35 μm .

CCKR:

210/198.2

DOCUMENT-IDENTIFIER: US 5605623 A
TITLE: Perfusive chromatography

DEPR:

Broadly, in accordance with the invention, perfusion chromatography is practiced by passing fluids at velocities above a threshold level through a specially designed matrix characterized by a geometry which is bimodal or multimodal with respect to its porosity. Perhaps the most fundamental observation relevant to the new procedure is that it is possible to avoid both the loss of capacity characteristic of convection bound systems and the high plate height and bandspreading characteristics of diffusion bound systems. This can be accomplished by forcing chromatography fluids through a matrix having a set of larger pores, such as are defined by the interstices among a bed of particles, and which determine pressure drops and fluid flow velocities through the bed, and a set of pores of smaller diameter, e.g., anisotropic throughpores. The smaller pores permeate the individual particles and serve to deliver chromatography fluids by convection to surface regions within the particle interactive with the solutes in the chromatography fluid.

DEPR:

From the foregoing description many of the basic engineering goals to be pursued in the fabrication of matrix materials suitable for the practice of perfusion chromatography will be apparent to those skilled in the art. Thus, what is needed to practice perfusion chromatography is a matrix which will not crush under pressure having a bimodal or preferably multimodal pore structure and as large a surface area per unit volume as possible. The first and second pore sets which give the material its bimodal flow properties

must have mean diameters relative to each other so as to permit convective flow through both sets of pores at high V.sub.beds. The provision of subpores in the matrix is not required to conduct perfusion chromatography but is preferred because of the inherent increase in surface area per unit volume of matrix material such a construction provides.

DEPR:

In contrast to the PL 4,000 material, which, with respect to its pore structure, is multimodal, a more ideal perfusive particle might comprise a plurality of sets of throughpores and subpores of differing mean diameters. A bimodal pore size distribution can be achieved in such particle by mixing equal ratios of particles having two discrete pore sizes or by engineering this feature at the polymerization stage. Ideally, mean diameter ratio between throughpore subsets would be less than 10, the mean diameter ratio between the smallest throughpore sets and the subpores would be less than 20, and the mean diameter ratio between the first pore set, i.e., the interstices among the particles making up the matrix, and the largest throughpore subset would be less than 70, and preferably less than 50. A multimodal material might be produced by agglomerating 500 .ANG. porons to form approximately 1 .mu.m clusters, which in turn are agglomerated to form 10 .mu.m aggregates, which in turn may be aggregated to form 100 .mu.m particles. In such a material, the 1 .mu.m clusters would have interstices of a mean diameter in the order of a few hundred .ANG.. These would define the capillary wall area, which is very high. Diffusive transport within these pores would rarely have to exceed a distance of 0.5 .mu.m or 5,000 .ANG.. Interstices among the 1 .mu.m clusters making up the 10 .mu.m aggregates would permit convective flow to feed

the diffusive pores. These would be on the order of 0.3 .mu.m in diameter.

These throughpores, in turn would be fed by larger pores defined by interstices among the 10 .mu.m particles making up the 100 .mu.m particles.

These would have a mean diameter on the order of 35 .mu.m.

CLPR:

8. The chromatography system of claim 7 wherein the packed particles define a bimodal or multimodal pore structure.

CCOR:

210/198.2

DOCUMENT-IDENTIFIER: US 5552041 A
TITLE: Perfusive chromatography

DEPR:

Broadly, in accordance with the invention, perfusion chromatography is practiced by passing fluids at velocities above a threshold level through a specially designed matrix characterized by a geometry which is bimodal or multimodal with respect to its porosity. Perhaps the most fundamental observation relevant to the new procedure is that it is possible to avoid both the loss of capacity characteristic of convection bound systems and the high plate height and bandspreading characteristics of diffusion bound systems. This can be accomplished by forcing chromatography fluids through a matrix having a set of larger pores, such as are defined by the interstices among a bed of particles, and which determine pressure drops and fluid flow velocities through the bed, and a set of pores of smaller diameter, e.g., anisotropic throughpores. The smaller pores permeate the individual particles and serve to deliver chromatography fluids by convection to surface regions within the particle interactive with the solutes in the chromatography fluid.

DEPR:

From the foregoing description many of the basic engineering goals to be practiced in the fabrication of matrix materials suitable for the practice of perfusive chromatography will be apparent to those skilled in the art. Thus, what is needed to practice perfusion chromatography is a matrix which will not crush under pressure having a bimodal or preferably multimodal pore structure and as large a surface area per unit volume as possible. The first and second pore sets which give the material its bimodal flow properties

must have mean diameters relative to each other so as to permit convective flow through both sets of pores at high V.sub.beds. The provision of subpores in the matrix is not required to conduct perfusion chromatography but is preferred because of the inherent increase in surface area per unit volume of matrix material such a construction provides.

DEFF:

In contrast to the PL 4,000 material, which, with respect to its pore structure, is multimodal, a more ideal perfusive particle might comprise a plurality of sets of throughpores and subpores of differing mean diameters. A bimodal pore size distribution can be achieved in such particle by mixing equal ratios of particles having two discrete pore sizes or by engineering this feature at the polymerization stage. Ideally, mean diameter ratio between throughpore subsets would be less than 10, the mean diameter ratio between the smallest throughpore sets and the subpores would be less than 20, and the mean diameter ratio between the first pore set, i.e., the interstices among the particles making up the matrix, and the largest throughpore subset would be less than 70, and preferably less than 50. A multimodal material might be produced by agglomerating 500 .ANG. porons to form approximately 1 .mu.m clusters, which in turn are agglomerated to form 10 .mu.m aggregates, which in turn may be aggregated to form 100 .mu.m particles. In such a design, the 1 .mu.m clusters would have interstices of a mean diameter in the vicinity of a few hundred .ANG.. These would define the surface area, which is very high. Diffusive transport within these pores would rarely have to exceed a distance of 0.5 .mu.m or 5,000 .ANG.. Interstices among the 1 .mu.m clusters making up the 10 .mu.m aggregates would permit convective flow to feed

the diffusive pores. These would be on the order of 0.3 μm in diameter.

These throughpores, in turn would be fed by larger pores defined by

interstices among the 10 μm particles making up the 100 μm particles.

These would have a mean diameter on the order of 35 μm .

CCOR:

210/198.2

DOCUMENT-IDENTIFIER: US 5540834 A
TITLE: Synthesis of porous inorganic particles by
polymerization-induced
colloid aggregation (PICA)

DEFR:

The specific surface area and porosity of the sintered particles measured by nitrogen adsorption were $13 \text{ m}^2/\text{g}$ and 29%, respectively, which are in reasonable agreement with close-packed, dense ZrO_2 spheres with nonuniform particle size. The pore size distributions (psd) obtained by nitrogen adsorption and desorption, and mercury porosimetry (intrusion) are displayed in FIGS. 4A, 4B and FIG. 5, respectively. From N_2 adsorption, the psd was determined to be rather narrow with a maximum near 400 Å and a small contribution of pores larger than 500 Å and smaller than 100 Å. From N_2 desorption, the psd was determined to be multimodal with nearly all pores between 100 Å and 200 Å in diameter and some pores below 50 Å. This discrepancy is mainly due to pore blocking or network effects, whereby desorption from a pore in a network is influenced by the state of the neighboring pores. Nitrogen adsorption probes the main channel size and can be considered free of pore blocking effects, while nitrogen desorption shows a disproportionately large amount of small pores due to "bottle necks." The psd obtained from mercury porosimetry (intrusion) are also influenced by pore interconnections; as shown in FIG. 5, it is broad with pores between approximately 125 Å and 300 Å in diameter and a maximum near 200 Å, in reasonable agreement with the psd from N_2 desorption.

DEPR:

The pore-size distributions (psd's) after sintering are shown in

FIG. 19. All samples exhibited multimodal psd's with pore diameters ranging between 100 .ANG. and 450 .ANG.. The sample synthesized at pH 1.2 contained some hollow particles but its psd seemed qualitatively similar to those of samples synthesized at higher pH (non-hollow particles). This is because N.sub.2 adsorption only probes the pores within the ZrO.sub.2 shells and not the large voids they encompass. Surface areas and porosities ($\epsilon_{\text{particle}}$) for these samples are listed in Table 3. Note that there is qualitative agreement with FIG. 12E, but quantitative agreement is not expected since condensing N.sub.2 cannot be used to distinguish between hollow cores and interstitial volume between aggregates. The N.sub.2 psd's show that the higher the pH, the greater contribution of small pores to the total porosity.

CCOR:
210/198.2

DOCUMENT-IDENTIFIER: US 5522994 A
TITLE: Single column chromatographic determination of small
molecules in
mixtures with large molecules

CCXR:
210/198.2

ORPL:
Little, "Sequential Multimodal Elution for Pseudomultidimensional
Liquid
Chromatography on a Single Column," Anal. Chem., 63 (1991) pp.
33-34.

DOCUMENT-IDENTIFIER: US 5431807 A

TITLE: Multimodal chromatographic separation media and process for using same

TTL:

Multimodal chromatographic separation media and process for using same

ABPL:

A process for carrying out in a consecutive fashion different modes of chromatographic separation in a liquid chromatography column using a single separation medium is disclosed. Separation media for use in such multimodal separations are also disclosed.

ESPR:

It may be possible to use combinations of different separation media in different columns for multimodal separations. An example of this multiple column bimodal separation was described recently by Wheatley J. E., J. Chromatogr., 603 (1992) 273. The bimodal separation of small molecules in one column packed with one separation medium and based on sequential multimodal elution was described by Little E. L., Jeansonne M. S., Foley J. E.; Anal Chem., 63, 1991, 33. They combined ion-exchange and reversed phase chromatography for the separation of a complex sample containing two groups of compounds: charged and non-polar. The use of two different gradients, i.e. a pH gradient and a methanol gradient, resulted in the separation of the charged compounds first, followed by the separation of the neutral molecules after switching to the second mobile phase. This approach makes use of imperfect surface functionalization of porous silica beads which contained C.sub.1, C.sub.8 or C.sub.18 groups together with the original acidic surface silanol groups. Similarly, the DIONEX OmniPack PAX-500 column is packed

with
non-porous poly[styrene-divinylbenzene] beads coated on the bead
surface with
attached ion-exchange latex particles (as described by the DIONEX
booklet).
Here again, the coating of the bead surface is imperfect and it
is the
non-covered hydrophobic areas of the original non-porous beads
that are used
for separation in the second mode. This approach excludes
combinations not
involving the reversed phase mode (the original ST-DVB surface
remains
non-polar even after attachment of latex particles) as well as
any size
exclusion separation.

BSPR:

This multimodal separation process is able to achieve separation
in a single
column in a consecutive operation because of the properties of
the separation
medium. The separation medium generally comprises a porous
material which has
been pretreated so that it has at least two different types of
surface groups
which have different functionalities. These different surface
groups are
disposed in different size range pores within the porous
material. Pore size
as used herein can mean a single measured average size, for
example, 25 nm, but
in most cases it means a particular range of sizes, for example,
50-500 nm. An
example of such a porous material of the present invention is one
wherein there
are hydrophilic surface groups in pores having a size of from
about 5-25 nm and
hydrophobic surface groups in pores ranging in a size of from
about 50-500 nm.
Another example is a material having hydrophilic groups in pores
below 25 nm in
size and hydrophobic groups in pores above 50 nm in size. As
discussed above,
the different functionalities of the surface groups, molecules
that have
affinities to such different surface groups may be separated
during different
modes of separation, which may be carried out in a consecutive
fashion. As

used herein, different molecules means molecules of different sizes, different chemical affinities, different structures, compositions, polarities, chiralities, activities, etc.

DRPF:

The key in selecting a mobile phase for a multimodal separation is the consecutive use of mobile phases, in each individual mode, that do not interfere with the absorption of compounds to be separated in any of the subsequent modes. Otherwise, the separation will not be multimodal and one group of compounds will leave the column without any separation, as documented in Example 6 and FIG. 7. Under this assumption, even a mobile phase for trimodal separation is easily designed by a person skilled in the art of liquid chromatography.

DRPF:

Reaction schemes 1-4 not only describe the particular sets of reactions leading to multimodal separation media but they also show the concepts of making such media in general. The starting polymer must be porous with relatively broad pore size distribution and possess reactive groups on the surface of the pores. Typically, the pore-size selectivity of the modification reactions are controlled by the molecular weight of the catalyst or reagent used in the particular modifying reaction and by the solvent. The number of modes accommodated in a separation medium is theoretically not limited but practically will rarely exceed three. The most important part in designing the reaction strategy for preparation of a multimodal medium is the right choice of the path. The product of a given reaction affecting pores of a given size should not affect the groups already built up in the previous reaction step within pores of a different size.

DRFR:

The multimodal separation process of the present invention may even use very tiny differences between the separation modes as is the case with reversed phase and hydrophobic interaction chromatography. The separation medium can be prepared by a set of reactions shown in Reaction Scheme 5.

CLPR:

1. A multimodal separation medium for use in liquid chromatography comprising a porous separation medium having at least two different pore size ranges with each pore size range containing a different surface group, having a different functionality compared to the surface groups in the other pore size range, said porous separation medium being capable of separating molecules in a sample added to a chromatography column containing said separation medium during different modes of separation which are carried out in a consecutive fashion using a single separation medium.

CLPR:

2. A multimodal separation medium for use in chromatography comprising a porous separation medium having at least two ranges of pore size, with each range of pore sizes having surface groups of a chemical composition different from that of other pore size ranges.

CCOR:

210/198.2

ORPL:

Title, "Sequential Multimodal Elution for Pseudomultidimensional Liquid Chromatography in a Single Column." Anal. Chem., 63 (1991), pp. 33-44.

DOCUMENT-IDENTIFIER: US 5384042 A
TITLE: Perfusive chromatography

DEFR:

Broadly, in accordance with the invention, perfusion chromatography is practiced by passing fluids at velocities above a threshold level through a specially designed matrix characterized by a geometry which is bimodal or multimodal with respect to its porosity. Perhaps the most fundamental observation relevant to the new procedure is that it is possible to avoid both the loss of capacity characteristic of convection bound systems and the high plate height and bandspreading characteristics of diffusion bound systems. This can be accomplished by forcing chromatography fluids through a matrix having a set of larger pores, such as are defined by the interstices among a bed of particles, and which determine pressure drops and fluid flow velocities through the bed, and a set of pores of smaller diameter, e.g., anisotropic throughpores. The smaller pores permeate the individual particles and serve to deliver chromatography fluids by convection to surface regions within the particle interactive with the solutes in the chromatography fluid.

DEPR:

From the foregoing description many of the basic engineering goals to be pursued in the fabrication of matrix materials suitable for the practice of perfusion chromatography will be apparent to those skilled in the art. That, what is needed to practice perfusion chromatography is a matrix which will not crush under pressure having a bimodal or preferably multimodal pore structure and as large a surface area per unit volume as possible. The first and second pore sets which give the material its bimodal flow properties

must have mean diameters relative to each other so as to permit convective flow through both sets of pores at high V.sub.beds. The provision of subpores in the matrix is not required to conduct perfusion chromatography but is preferred because of the inherent increase in surface area per unit volume of matrix material such a construction provides.

DEPR:

In contrast to the PL 4,000 material, which, with respect to its pore structure, is multimodal, a more ideal perfusive particle might comprise a plurality of sets of throughpores and subpores of differing mean diameters. A bimodal pore size distribution can be achieved in such particle by mixing equal ratios of particles having two discrete pore sizes or by engineering this feature at the polymerization stage. Ideally, mean diameter ratio between throughpore subsets would be less than 10, the mean diameter ratio between the smallest throughpore sets and the subpores would be less than 20, and the mean diameter ratio between the first pore set, i.e., the interstices among the particles making up the matrix, and the largest throughpore subset would be less than 70, and preferably less than 50. A multimodal material might be produced by agglomerating 500 .ANG. pores to form approximately 1 .mu.m clusters, which in turn are agglomerated to form 10 .mu.m aggregates, which in turn may be aggregated to form 100 .mu.m particles. In such a design, the 1 .mu.m clusters would have interstices of mean diameter in the vicinity of 0.5 .mu.m or 5,000 .ANG.. This would define the edges of a very high surface area. Diffusive transport within these pores would rarely have to exceed a distance of 0.5 .mu.m or 5,000 .ANG.. Interstices among the 1 .mu.m clusters making up the 10 .mu.m aggregates would permit convective flow to feed

the diffusive pores. These would be on the order of 0.3 μm in diameter.

These throughpores, in turn would be fed by larger pores defined by interstices among the 10 μm particles making up the 100 μm particles.

These would have a mean diameter on the order of 35 μm .

CLPR:

22. A matrix for conducting high efficiency adsorption chromatography of biological molecules, the matrix comprising a packed bed of rigid, polystyrene divinylbenzene particles having a mean diameter between 10 and 20 micrometers, defining a bimodal or multimodal pore structure and comprising chemically active regions linked to the surface of the particles for reversibly sorbing biological molecules, said matrix being characterized in that

CLPV:

a packed bed of rigid particles comprising an inorganic material, the particles having a mean diameter within the range of 20 micrometers to 100 micrometers and defining a bimodal or multimodal pore structure, one set of pores being particle transecting throughpores having a mean diameter greater than at least 4000 \AA , another set of pores being subpores in fluid communication with the throughpores, and, disposed within at least said subpores, reactive groups comprising one of anionic sulfonate groups, cationic quaternary ammonium groups, immunoglobulines, or hydrocarbons, the ratio of the means diameter of the particles to the mean diameter of a throughpores passing through the particle being sufficient to permit convective transport of biological molecules into the throughpores, the rate of chromatographic separation so that, at flow rates greater than 100 cm/hr, the rate of biological molecule transport into the throughpores is dependent on the velocity of liquid passing through the bed, and sorption capacity remains

substantially constant over a range of flowrates.

CCOR:

210/198.2

DOCUMENT-IDENTIFIER: US 5316680 A

TITLE: Multimodal chromatographic separation media and process for using same

TTL:

Multimodal chromatographic separation media and process for using same

ABPL:

A process for carrying out in a consecutive fashion different modes of chromatographic separation in a liquid chromatography column using a single separation medium is disclosed. Separation media for use in such multimodal separations are also disclosed.

ESPR:

It may be possible to use combinations of different separation media in different columns for multimodal separations. An example of this multiple column bimodal separation was described recently by Wheatley J. B., J. Chromatogr., 603 (1992) 273. The bimodal separation of small molecules in one column packed with one separation medium and based on sequential multimodal elution was described by Little E. L., Jeanson M. S., Foley J. F.; Anal Chem., 63, 1991, 23. They combined ion-exchange and reversed phase chromatography for the separation of a complex sample containing two groups of compounds: charged and non-polar. The use of two different gradients, i.e. a pH gradient and a methanol gradient, resulted in the separation of the charged molecules followed by the separation of the neutral molecules after switching to the second mobile phase. This approach makes use of imperfect surface functionalization of porous silica beads which contained C.sub.1, C.sub.8 or C.sub.18 groups together with the original acidic surface silanol groups. Similarly, the DIONEX OmniPack PAX-500 column is packed

with non-porous poly[styrene-divinylbenzene] beads coated on the bead surface with attached ion-exchange latex particles (as described by the DIONEX booklet). Here again, the coating of the bead surface is imperfect and it is the non-covered hydrophobic areas of the original non-porous beads that are used for separation in the second mode. This approach excludes combinations not involving the reversed phase mode (the original ST-DVB surface remains non-polar even after attachment of latex particles) as well as any size exclusion separation.

BSPR:

This multimodal separation process is able to achieve separation in a single column in a consecutive operation because of the properties of the separation medium. The separation medium generally comprises a porous material which has been pretreated so that it has at least two different types of surface groups which have different functionalities. These different surface groups are disposed in different size range pores within the porous material. Pore size as used herein can mean a single measured average size, for example, 25 nm, but in most cases it means a particular range of sizes, for example, 50-500 nm. An example of such a porous material of the present invention is one wherein there are hydrophilic surface groups in pores having a size of from about 5-25 nm and hydrophobic surface groups in pores ranging in a size of from about 50-500 nm. Another example is a material having hydrophilic groups in pores of a size of from about 5-25 nm and hydrophobic groups in pores above 25 nm in size. As a result of the different functionalities of the surface groups, molecules that have affinities to such different surface groups may be separated during different modes of separation, which may be carried out in a consecutive fashion. As

used herein, different molecules means molecules of different sizes, different chemical affinities, different structures, compositions, polarities, chiralities, activities, etc.

DEPR:

The key in selecting a mobile phase for a multimodal separation is the consecutive use of mobile phases, in each individual mode, that do not interfere with the absorption of compounds to be separated in any of the subsequent modes. Otherwise, the separation will not be multimodal and one group of compounds will leave the column without any separation, as documented in Example 6 and FIG. 7. Under this assumption, even a mobile phase for trimodal separation is easily designed by a person skilled in the art of liquid chromatography.

OEPR:

Reaction Schemes 1-4 not only describe the particular sets of reactions leading to multimodal separation media but they also show the concepts of making such media in general. The starting polymer must be porous with relatively broad pore size distribution and possess reactive groups on the surface of the pores. Typically, the pore-size selectivity of the modification reactions are controlled by the molecular weight of the catalyst or reagent used in the particular modifying reaction and by the solvent. The number of modes accommodated in a separation medium is theoretically not limited but practically will rarely exceed three. The most important part in building the reaction strategy for preparation of a multimodal media is the choice of the path. The product of a given reaction affecting pores of a given size should not affect the groups already built up in the previous reaction step within pores of a different size.

DEPR:

The multimodal separation process of the present invention may even use very tiny differences between the separation modes as is the case with reversed phase and hydrophobic interaction chromatography. The separation medium can be prepared by a set of reactions shown in Reaction Scheme 5.

CCXR:

210/198.2

ORPL:

Little, "Sequential Multimodal Elution for Pseudomultidimensional Liquid Chromatography on a Single Column," Anal. Chem., 63, (1991) pp. 33-44.

DOCUMENT-IDENTIFIER: US 5228989 A
TITLE: Perfusive chromatography

DEPR:

Broadly, in accordance with the invention, perfusion chromatography is practiced by passing fluids at velocities above a threshold level through a specially designed matrix characterized by a geometry which is bimodal or multimodal with respect to its porosity. Perhaps the most fundamental observation relevant to the new procedure is that it is possible to avoid both the loss of capacity characteristic of convection bound systems and the high plate height and bandspreading characteristics of diffusion bound systems. This can be accomplished by forcing chromatography fluids through a matrix having a set of larger pores, such as are defined by the interstices among a bed of particles, and which determine pressure drops and fluid flow velocities through the bed, and a set of pores of smaller diameter, e.g., anisotropic throughpores. The smaller pores permeate the individual particles and serve to deliver chromatography fluids by convection to surface regions within the particle interactive with the solutes in the chromatography fluid.

DEPR:

From the foregoing description many of the basic engineering goals to be pursued in the fabrication of matrix materials suitable for the practice of perfusive chromatography will be apparent to those skilled in the art. Thus, what is needed to practice perfusion chromatography is a matrix which will not crush under pressure having a bimodal or preferably multimodal pore structure and as large a surface area per unit volume as possible. The first and second pore sets which give the material its bimodal flow properties

must have mean diameters relative to each other so as to permit convective flow through both sets of pores at high V.sub.beds. The provision of subpores in the matrix is not required to conduct perfusion chromatography but is preferred because of the inherent increase in surface area per unit volume of matrix material such a construction provides.

DEFF:

In contrast to the PL 4,000 material, which, with respect to its pore structure, is multimodal, a more ideal perfusive particle might comprise a plurality of sets of throughpores and subpores of differing mean diameters. A bimodal pore size distribution can be achieved in such particle by mixing equal ratios of particles having two discrete pore sizes or by engineering this feature at the polymerization stage. Ideally, mean diameter ratio between throughpore subsets would be less than 10, the mean diameter ratio between the smallest throughpore sets and the subpores would be less than 20, and the mean diameter ratio between the first pore set, i.e., the interstices among the particles making up the matrix, and the largest throughpore subset would be less than 70, and preferably less than 50. A multimodal material might be produced by agglomerating 500 .ANG. porons to form approximately 1 .mu.m clusters, which in turn are agglomerated to form 10 .mu.m aggregates, which in turn may be aggregated to form 100 .mu.m particles. In such a design, the 1 .mu.m clusters would have interstices of a mean diameter in the order of a few hundred .ANG.. These would define the only pore size of a fairly high surface area. Diffusive transport within these pores would rarely have to exceed a distance of 0.5 .mu.m or 5,000 .ANG.. Interstices among the 1 .mu.m clusters making up the 10 .mu.m aggregates would permit convective flow to feed

the diffusive pores. These would be on the order of 0.3 μm in diameter.

These throughpores, in turn would be fed by larger pores defined by interstices among the 10 μm particles making up the 100 μm particles.

These would have a mean diameter on the order of 35 μm .

CCOR:

210/198.2

DOCUMENT-IDENTIFIER: US 5104530 A

TITLE: Chromatography column with carbonaceous adsorbents from
pyrolyzed
polysulfonated polymers

ABPL:

Carbonaceous adsorbent particles having multimodal pore size, including micropores and macropores, with improved adsorptive and separative properties, are prepared by partial pyrolysis of polysulfonated macroporous precursor resins, said resins being in turn derived from macroporous poly(vinylaromatic) resins. The particles may be further treated by activating with reactive gases or by functionalization.

ESPR:

British Patent No 1,525,420, in a broad description of method for rendering infusible various porous high molecular weight compounds (including macroporous resins), and then calcining them, relates techniques for polysulfonation earlier described by Corte et al. among those suitable for creating infusibility. No characterization data are given for the polymer prior to calcination. Preferred infusibility reactants are sulfur trioxide, sulfuric acid, or chlorosulfonic acid. This reference discloses pyrolysis of macroporous resins treated with 15% fuming sulfuric acid and pyrolyzed, and describes an experimental method for determining the porosity of the pyrolyzed material down to 1-5 nm. The results described in the patent show the absence of any porosity development below 100 nm, and multimodal porosity is not taught. In contrast, Neely in the cited references fully shows the development of microporosity for monosulfonated macroporous resins. Further, the British patent is silent about the processing advantages observed

in pyrolysis of polysulfonated resins.

CLPR:

1. A chromatographic column packed with the carbonaceous adsorbent particles which comprise the product of controlled pyrolysis of a polysulfonated macroporous crosslinked, vinylaromatic polymer, the particles having multimodal pore-size distribution and a minimum micropore volume of about 0.02 cm.sup.3 /g.

CLPR:

4. A chromatographic column packed with the carbonaceous adsorbent particles which comprise the product of controlled pyrolysis of a polysulfonated macroporous crosslinked, vinylaromatic polymer, the particles having multimodal pore-size distribution and a minimum micropore volume of about 0.02 cm.sup.3 /g, wherein the particles are treated, subsequent to pyrolysis, with adsorbable reactive agent.

CCOR:

210/198.2

DOCUMENT-IDENTIFIER: US 5071547 A

TITLE: Column chromatographic column apparatus with switching capability

DEFF:

The fluid conduit assembly or means is comprised of conduit piping connecting the dual columns, detector, multi-valve arrangement in a similar manner to that of FIG. 2 with three exceptions. First, the fluid conduit assembly in addition to conduits 11A through 11L as in FIG. 2, also has conduits 11X through 11Z and 11Z'. Second, there are two valves present in what conceptually can be referred to as each valve set 12A and 12B of FIG. 2. The third exception is the presence of an additional valve 42 that isolates the detector from the pressure of the DCC Apparatus. Although valves 12iv and 12v can be indicated as being in the valve set 12A of FIG. 2 and valves 12vi and 12vii can be indicated as being in the valve set 12B of FIG. 2, these valves may exist with independent identity from these valve sets. In other words these valves may be just a plurality of valves without valves 12iv and 12v as well as the two valves 12vi and 12vii perform the same function as valve sets 12A and 12B, respectively. Valve 42 is any multi-port and multimodal valve known to those skilled in the art as are valves 12iv through 12vii, and valve 42 can be considered a part of the multi-valve arrangement 12. As with the DCC Apparatus of FIG. 2, that of FIG. 3 has each valve with an actuating connection 14 to the controller 12. So valve 42 is again connected to the controller 12 by an actuating connection 14A as valves 12iv-12vii and 42 are connected by connections 14J, 14K, 14L, 14M, and 14N, respectively. Valves 12iv and 12v are connected together for pressurized fluid passage between them by conduit 11X,

and in a similar manner valves 12vi and 12vii are connected by conduit 11Y.

CCOR:

210/198.2

DOCUMENT-IDENTIFIER: US 5019270 A
TITLE: Perfusive chromatography

DEPR:

Broadly, in accordance with the invention, perfusion chromatography is practiced by passing fluids at velocities above a threshold level through a specially designed matrix characterized by a geometry which is bimodal or multimodal with respect to its porosity. Perhaps the most fundamental observation relevant to the new procedure is that it is possible to avoid both the loss of capacity characteristic of convection bound systems and the high plate height and bandspreading characteristics of diffusion bound systems. This can be accomplished by forcing chromatography fluids through a matrix having a set of larger pores, such as are defined by the interstices among a bed of particles, and which determine pressure drops and fluid flow velocities through the bed, and a set of pores of smaller diameter, e.g., anisotropic throughpores. The smaller pores permeate the individual particles and serve to deliver chromatography fluids by convection to surface regions within the particle interactive with the solutes in the chromatography fluid.

DEPR:

From the foregoing description many of the basic engineering goals to be pursued in the fabrication of matrix materials suitable for the practice of perfusion chromatography will be apparent to those skilled in the art. Thus, what is needed to practice perfusion chromatography is a matrix which will not crush under pressure having a bimodal or preferably multimodal pore structure and as large a surface area per unit volume as possible. The first and second pore sets which give the material its bimodal flow properties

must have mean diameters relative to each other so as to permit convective flow through both sets of pores at high V.sub.beds. The provision of subpores in the matrix is not required to conduct perfusion chromatography but is preferred because of the inherent increase in surface area per unit volume of matrix material such a construction provides.

DEPR:

In contrast to the PL 4,000 material, which, with respect to its pore structure, is multimodal, a more ideal perfusive particle might comprise a plurality of sets of throughpores and subpores of differing mean diameters. A bimodal pore size distribution can be achieved in such particle by mixing equal ratios of particles having two discrete pore sizes or by engineering this feature at the polymerization stage. Ideally, mean diameter ratio between throughpore subsets would be less than 10, the mean diameter ratio between the smallest throughpore sets and the subpores would be less than 20, and the mean diameter ratio between the first pore set, i.e., the interstices among the particles making up the matrix, and the largest throughpore subset would be less than 70, and preferably less than 50. A multimodal material might be produced by agglomerating 500 .ANG. pores to form approximately 1 .mu.m clusters, which in turn are agglomerated to form 10 .mu.m aggregates, which in turn may be aggregated to form 100 .mu.m particles. In such a system, the 1 .mu.m clusters would have interstices of a mean diameter in the range of a few hundred .ANG.. These would define the subpores and provide a very high surface area. Diffusive transport within these pores would rarely have to exceed a distance of 0.5 .mu.m or 5,000 .ANG.. Interstices among the 1 .mu.m clusters making up the 10 .mu.m aggregates would permit convective flow to feed

the diffusive pores. These would be on the order of 0.3 μm in diameter.

These throughpores, in turn would be fed by larger pores defined by

interstices among the 10 μm particles making up the 100 μm particles.

These would have a mean diameter on the order of 35 μm .

CCXR:

210/198.2

DOCUMENT-IDENTIFIER: US 5833861 A
TITLE: Perfusive chromatography

DEPR:

Broadly, in accordance with the invention, perfusion chromatography is practiced by passing fluids at velocities above a threshold level through a specially designed matrix characterized by a geometry which is bimodal or multimodal with respect to its porosity. Perhaps the most fundamental observation relevant to the new procedure is that it is possible to avoid both the loss of capacity characteristic of convection bound systems and the high plate height and bandspreading characteristics of diffusion bound systems. This can be accomplished by forcing chromatography fluids through a matrix having a set of larger pores, such as are defined by the interstices among a bed of particles, and which determine pressure drops and fluid flow velocities through the bed, and a set of pores of smaller diameter, e.g., anisotropic throughpores. The smaller pores permeate the individual particles and serve to deliver chromatography fluids by convection to surface regions within the particle interactive with the solutes in the chromatography fluid.

DEPR:

From the foregoing description many of the basic engineering goals to be pursued in the fabrication of matrix materials suitable for the practice of perfusive chromatography will be apparent to those skilled in the art. Thus, what is needed to practice perfusion chromatography is a matrix which will not crush under pressure having a bimodal or preferably multimodal pore structure and as large a surface area per unit volume as possible. The first and second pore sets which give the material its bimodal flow properties

must have mean diameters relative to each other so as to permit convective flow through both sets of pores at high V.sub.beds. The provision of subpores in the matrix is not required to conduct perfusion chromatography but is preferred because of the inherent increase in surface area per unit volume of matrix material such a construction provides.

DEPR:

In contrast to the PL 4,000 material, which, with respect to its pore structure, is multimodal, a more ideal perfusive particle might comprise a plurality of sets of throughpores and subpores of differing mean diameters. A bimodal pore size distribution can be achieved in such particle by mixing equal ratios of particles having two discrete pore sizes or by engineering this feature at the polymerization stage. Ideally, mean diameter ratio between throughpore subsets would be less than 10, the mean diameter ratio between the smallest throughpore sets and the subpores would be less than 20, and the mean diameter ratio between the first pore set, i.e., the interstices among the particles making up the matrix, and the largest throughpore subset would be less than 70, and preferably less than 50. A multimodal material might be produced by agglomerating 500 .ANG. porons to form approximately 1 .mu.m clusters, which in turn are agglomerated to form 10 .mu.m aggregates, which in turn may be aggregated to form 100 .mu.m particles. In such a design, the 1 .mu.m clusters would have interstices of a mean diameter in the order of a few hundred .ANG.. These would define the subpores and provide a very high surface area. Diffusive transport within these pores would rarely have to exceed a distance of 0.5 .mu.m or 5,000 .ANG.. Interstices among the 1 .mu.m clusters making up the 10 .mu.m aggregates would permit convective flow to feed

the diffusive pores. These would be on the order of 0.3 μm in diameter.

These throughpores, in turn would be fed by larger pores defined by

interstices among the 10 μm particles making up the 100 μm particles.

These would have a mean diameter on the order of 35 μm .

CCKR:

210/198.2

DOCUMENT-IDENTIFIER: US 5605623 A
TITLE: Perfusive chromatography

DEPR:

Broadly, in accordance with the invention, perfusion chromatography is practiced by passing fluids at velocities above a threshold level through a specially designed matrix characterized by a geometry which is bimodal or multimodal with respect to its porosity. Perhaps the most fundamental observation relevant to the new procedure is that it is possible to avoid both the loss of capacity characteristic of convection bound systems and the high plate height and bandspreading characteristics of diffusion bound systems. This can be accomplished by forcing chromatography fluids through a matrix having a set of larger pores, such as are defined by the interstices among a bed of particles, and which determine pressure drops and fluid flow velocities through the bed, and a set of pores of smaller diameter, e.g., anisotropic throughpores. The smaller pores permeate the individual particles and serve to deliver chromatography fluids by convection to surface regions within the particle interactive with the solutes in the chromatography fluid.

DEPR:

From the foregoing description many of the basic engineering goals to be pursued in the fabrication of matrix materials suitable for the practice of perfusion chromatography will be apparent to those skilled in the art. Namely, what is needed to practice perfusion chromatography is a matrix which will not crush under pressure having a bimodal or preferably multimodal pore structure and as large a surface area per unit volume as possible. The first and second pore sets which give the material its bimodal flow properties

must have mean diameters relative to each other so as to permit convective flow through both sets of pores at high V.sub.beds. The provision of subpores in the matrix is not required to conduct perfusion chromatography but is preferred because of the inherent increase in surface area per unit volume of matrix material such a construction provides.

DEPR:

In contrast to the PL 4,000 material, which, with respect to its pore structure, is multimodal, a more ideal perfusive particle might comprise a plurality of sets of throughpores and subpores of differing mean diameters. A bimodal pore size distribution can be achieved in such particle by mixing equal ratios of particles having two discrete pore sizes or by engineering this feature at the polymerization stage. Ideally, mean diameter ratio between throughpore subsets would be less than 10, the mean diameter ratio between the smallest throughpore sets and the subpores would be less than 20, and the mean diameter ratio between the first pore set, i.e., the interstices among the particles making up the matrix, and the largest throughpore subset would be less than 70, and preferably less than 50. A multimodal material might be produced by agglomerating 500 .ANG. porons to form approximately 1 .mu.m clusters, which in turn are agglomerated to form 10 .mu.m aggregates, which in turn may be aggregated to form 100 .mu.m particles. In such a design, the 1 .mu.m clusters would have interstices of a mean diameter in the vicinity of a few hundred .ANG.. These would define the surface area and thus a very high surface area. Diffusive transport within these pores would rarely have to exceed a distance of 0.5 .mu.m or 5,000 .ANG.. Interstices among the 1 .mu.m clusters making up the 10 .mu.m aggregates would permit convective flow to feed

the diffusive pores. These would be on the order of 0.3 .mu.m in diameter.

These throughpores, in turn would be fed by larger pores defined by interstices among the 10 .mu.m particles making up the 100 .mu.m particles.

These would have a mean diameter on the order of 35 .mu.m.

CLPR:

8. The chromatography system of claim 7 wherein the packed particles define a bimodal or multimodal pore structure.

CCOR:

210/198.2

DOCUMENT-IDENTIFIER: US 5552041 A
TITLE: Perfusive chromatography

DEPR:

Broadly, in accordance with the invention, perfusion chromatography is practiced by passing fluids at velocities above a threshold level through a specially designed matrix characterized by a geometry which is bimodal or multimodal with respect to its porosity. Perhaps the most fundamental observation relevant to the new procedure is that it is possible to avoid both the loss of capacity characteristic of convection bound systems and the high plate height and bandspreading characteristics of diffusion bound systems. This can be accomplished by forcing chromatography fluids through a matrix having a set of larger pores, such as are defined by the interstices among a bed of particles, and which determine pressure drops and fluid flow velocities through the bed, and a set of pores of smaller diameter, e.g., anisotropic throughpores. The smaller pores permeate the individual particles and serve to deliver chromatography fluids by convection to surface regions within the particle interactive with the solutes in the chromatography fluid.

DEPR:

From the foregoing description many of the basic engineering goals to be pursued in the fabrication of matrix materials suitable for the practice of perfusive chromatography will be apparent to those skilled in the art. Thus, what is needed for practice perfusion chromatography is a matrix which will not crush under pressures having a bimodal or preferably multimodal pore structure and as large a surface area per unit volume as possible. The first and second pore sets which give the material its bimodal flow properties

must have mean diameters relative to each other so as to permit convective flow through both sets of pores at high V.sub.beds. The provision of subpores in the matrix is not required to conduct perfusion chromatography but is preferred because of the inherent increase in surface area per unit volume of matrix material such a construction provides.

DEFF:

In contrast to the PL 4,000 material, which, with respect to its pore structure, is multimodal, a more ideal perfusive particle might comprise a plurality of sets of throughpores and subpores of differing mean diameters. A bimodal pore size distribution can be achieved in such particle by mixing equal ratios of particles having two discrete pore sizes or by engineering this feature at the polymerization stage. Ideally, mean diameter ratio between throughpore subsets would be less than 10, the mean diameter ratio between the smallest throughpore sets and the subpores would be less than 20, and the mean diameter ratio between the first pore set, i.e., the interstices among the particles making up the matrix, and the largest throughpore subset would be less than 70, and preferably less than 50. A multimodal material might be produced by agglomerating 500 .ANG. porons to form approximately 1 .mu.m clusters, which in turn are agglomerated to form 10 .mu.m aggregates, which in turn may be aggregated to form 100 .mu.m particles. In such a design, the 1 .mu.m clusters would have interstices of a mean diameter in the order of a few hundred .ANG.. These would define the largest and provide very high surface area. Diffusive transport within these pores would rarely have to exceed a distance of 0.5 .mu.m or 5,000 .ANG.. Interstices among the 1 .mu.m clusters making up the 10 .mu.m aggregates would permit convective flow to feed

the diffusive pores. These would be on the order of 0.3 μm in diameter.

These throughpores, in turn would be fed by larger pores defined by

interstices among the 10 μm particles making up the 100 μm particles.

These would have a mean diameter on the order of 35 μm .

CCOR:

210/198.2

DOCUMENT-IDENTIFIER: US 5540834 A
TITLE: Synthesis of porous inorganic particles by
polymerization-induced
colloid aggregation (PICA)

DEPR:

The specific surface area and porosity of the sintered particles measured by nitrogen adsorption were $13 \text{ m}^2/\text{g}$ and 29%, respectively, which are in reasonable agreement with close-packed, dense ZrO_2 spheres with nonuniform particle size. The pore size distributions (psd) obtained by nitrogen adsorption and desorption, and mercury porosimetry (intrusion) are displayed in FIGS. 4A, 4B and FIG. 5, respectively. From N_2 adsorption, the psd was determined to be rather narrow with a maximum near 400 Å and a small contribution of pores larger than 500 Å and smaller than 100 Å. From N_2 desorption, the psd was determined to be multimodal with nearly all pores between 100 Å and 200 Å in diameter and some pores below 50 Å. This discrepancy is mainly due to pore blocking or network effects, whereby desorption from a pore in a network is influenced by the state of the neighboring pores. Nitrogen adsorption probes the main channel size and can be considered free of pore blocking effects, while nitrogen desorption shows a disproportionately large amount of small pores due to "bottle necks." The psd obtained from mercury porosimetry (intrusion) are also influenced by pore interconnections; as shown in FIG. 5, it is broad with pores between approximately 125 Å and 300 Å in diameter and a maximum near 200 Å, in reasonable agreement with the psd from N_2 desorption.

DEPR:

The pore-size distributions (psd's) after sintering are shown in

FIG. 19. All samples exhibited multimodal psd's with pore diameters ranging between 100 .ANG. and 450 .ANG.. The sample synthesized at pH 1.2 contained some hollow particles but its psd seemed qualitatively similar to those of samples synthesized at higher pH (non-hollow particles). This is because N.sub.2 adsorption only probes the pores within the ZrO.sub.2 shells and not the large voids they encompass. Surface areas and porosities ($\epsilon_{\text{particle}}$) for these samples are listed in Table 3. Note that there is qualitative agreement with FIG. 12E, but quantitative agreement is not expected since condensing N.sub.2 cannot be used to distinguish between hollow cores and interstitial volume between aggregates. The N.sub.2 psd's show that the higher the pH, the greater contribution of small pores to the total porosity.

CCOR:
210/198.2

DOCUMENT-IDENTIFIER: US 5522994 A
TITLE: Single column chromatographic determination of small
molecules in
mixtures with large molecules

CCKR:
210/198.2

ORPL:
Little, "Sequential Multimodal Elution for Pseudomultidimensional
Liquid
Chromatography on a Single Column," Anal. Chem., 63 (1991) pp.
33-34.

DOCUMENT-IDENTIFIER: US 5431807 A

TITLE: Multimodal chromatographic separation media and process for using same

TTL:

Multimodal chromatographic separation media and process for using same

ABPL:

A process for carrying out in a consecutive fashion different modes of chromatographic separation in a liquid chromatography column using a single separation medium is disclosed. Separation media for use in such multimodal separations are also disclosed.

BSPP:

It may be possible to use combinations of different separation media in different columns for multimodal separations. An example of this multiple column bimodal separation was described recently by Wheatley J. E., J. Chromatogr., 603 (1992) 273. The bimodal separation of small molecules in one column packed with one separation medium and based on sequential multimodal elution was described by Little E. L., Jeansonne M. S., Foley J. E.; Anal Chem., 63, 1991, 13. They combined ion-exchange and reversed phase chromatography for the separation of a complex sample containing two groups of compounds: charged and non-polar. The use of two different gradients, i.e. a pH gradient and a methanol gradient, resulted in the separation of the charged molecules followed by the separation of the neutral molecules after switching to the second mobile phase. This approach makes use of imperfect surface functionalization of porous silica beads which contained C.sub.1, C.sub.8 or C.sub.18 groups together with the original acidic surface silanol groups. Similarly, the BIONEX OmniPack PAX-500 column is packed

with non-porous poly[styrene-divinylbenzene] beads coated on the bead surface with attached ion-exchange latex particles (as described by the DIONEX booklet). Here again, the coating of the bead surface is imperfect and it is the non-covered hydrophobic areas of the original non-porous beads that are used for separation in the second mode. This approach excludes combinations not involving the reversed phase mode (the original ST-DVB surface remains non-polar even after attachment of latex particles) as well as any size exclusion separation.

BSPR:

This multimodal separation process is able to achieve separation in a single column in a consecutive operation because of the properties of the separation medium. The separation medium generally comprises a porous material which has been pretreated so that it has at least two different types of surface groups which have different functionalities. These different surface groups are disposed in different size range pores within the porous material. Pore size as used herein can mean a single measured average size, for example, 25 nm, but in most cases it means a particular range of sizes, for example, 50-500 nm. An example of such a porous material of the present invention is one wherein there are hydrophilic surface groups in pores having a size of from about 5-25 nm and hydrophobic surface groups in pores ranging in a size of from about 50-500 nm. Another example is a material having hydrophilic groups in pores of 5-25 nm in size and hydrophobic groups in pores of 50-500 nm in size. Because of the different functionalities of the surface groups, molecules that have affinities to such different surface groups may be separated during different modes of separation, which may be carried out in a consecutive fashion. As

used herein, different molecules means molecules of different sizes, different chemical affinities, different structures, compositions, polarities, chiralities, activities, etc.

DRPR:

The key in selecting a mobile phase for a multimodal separation is the consecutive use of mobile phases, in each individual mode, that do not interfere with the absorption of compounds to be separated in any of the subsequent modes. Otherwise, the separation will not be multimodal and one group of compounds will leave the column without any separation, as documented in Example 6 and FIG. 7. Under this assumption, even a mobile phase for trimodal separation is easily designed by a person skilled in the art of liquid chromatography.

DRPR:

Reaction Schemes 1-4 not only describe the particular sets of reactions leading to multimodal separation media but they also show the concepts of making such media in general. The starting polymer must be porous with relatively broad pore size distribution and possess reactive groups on the surface of the pores. Typically, the pore-size selectivity of the modification reactions are controlled by the molecular weight of the catalyst or reagent used in the particular modifying reaction and by the solvent. The number of modes accommodated in a separation medium is theoretically not limited but practically will rarely exceed three. The most important part in selecting the reaction strategy for preparation of a multimodal medium is the right choice of the path. The product of a given reaction affecting pores of a given size should not affect the groups already built up in the previous reaction step within pores of a different size.

DRPR:

The multimodal separation process of the present invention may even use very tiny differences between the separation modes as is the case with reversed phase and hydrophobic interaction chromatography. The separation medium can be prepared by a set of reactions shown in Reaction Scheme 5.

CLPF:

1. A multimodal separation medium for use in liquid chromatography comprising a porous separation medium having at least two different pore size ranges with each pore size range containing a different surface group, having a different functionality compared to the surface groups in the other pore size range, said porous separation medium being capable of separating molecules in a sample added to a chromatography column containing said separation medium during different modes of separation which are carried out in a consecutive fashion using a single separation medium.

CLPF:

2. A multimodal separation medium for use in chromatography comprising a porous separation medium having at least two ranges of pore size, with each range of pore sizes having surface groups of a chemical composition different from that of other pore size ranges.

CLPF:

210/198.2

OEPL:

Title, "Sequential Multimodal Elution for Pseudomultidimensional Liquid Chromatography in a Single Column," Anal. Chem., 63 (1991) pp. 33-44.

DOCUMENT-IDENTIFIER: US 5384042 A
TITLE: Perfusive chromatography

DEPR:

Broadly, in accordance with the invention, perfusion chromatography is practiced by passing fluids at velocities above a threshold level through a specially designed matrix characterized by a geometry which is bimodal or multimodal with respect to its porosity. Perhaps the most fundamental observation relevant to the new procedure is that it is possible to avoid both the loss of capacity characteristic of convection bound systems and the high plate height and bandspreading characteristics of diffusion bound systems. This can be accomplished by forcing chromatography fluids through a matrix having a set of larger pores, such as are defined by the interstices among a bed of particles, and which determine pressure drops and fluid flow velocities through the bed, and a set of pores of smaller diameter, e.g., anisotropic throughpores. The smaller pores permeate the individual particles and serve to deliver chromatography fluids by convection to surface regions within the particle interactive with the solutes in the chromatography fluid.

DEPR:

From the foregoing description many of the basic engineering goals to be pursued in the fabrication of matrix materials suitable for the practice of perfusion chromatography will be apparent to those skilled in the art. Thus, what is needed to practice perfusion chromatography is a matrix which will not crush under pressure having a bimodal or preferably multimodal pore structure and as large a surface area per unit volume as possible. The first and second pore sets which give the material its bimodal flow properties

must have mean diameters relative to each other so as to permit convective flow through both sets of pores at high V.sub.beds. The provision of subpores in the matrix is not required to conduct perfusion chromatography but is preferred because of the inherent increase in surface area per unit volume of matrix material such a construction provides.

DEPR:

In contrast to the PL 4,000 material, which, with respect to its pore structure, is multimodal, a more ideal perfusive particle might comprise a plurality of sets of throughpores and subpores of differing mean diameters. A bimodal pore size distribution can be achieved in such particle by mixing equal ratios of particles having two discrete pore sizes or by engineering this feature at the polymerization stage. Ideally, mean diameter ratio between throughpore subsets would be less than 10, the mean diameter ratio between the smallest throughpore sets and the subpores would be less than 20, and the mean diameter ratio between the first pore set, i.e., the interstices among the particles making up the matrix, and the largest throughpore subset would be less than 70, and preferably less than 50. A multimodal material might be produced by agglomerating 500 .ANG. porons to form approximately 1 .mu.m clusters, which in turn are agglomerated to form 10 .mu.m aggregates, which in turn may be aggregated to form 100 .mu.m particles. In such a way, the 1 .mu.m clusters would have interstices of a mean diameter of a few hundred .ANG.. These would define the support surface area. Diffusive transport within these pores would rarely have to exceed a distance of 0.5 .mu.m or 5,000 .ANG.. Interstices among the 1 .mu.m clusters making up the 10 .mu.m aggregates would permit convective flow to feed

the diffusive pores. These would be on the order of 0.3 .mu.m in diameter.

These throughpores, in turn would be fed by larger pores defined by interstices among the 10 .mu.m particles making up the 100 .mu.m particles.

These would have a mean diameter on the order of 35 .mu.m.

CLPF:

22. A matrix for conducting high efficiency adsorption chromatography of biological molecules, the matrix comprising a packed bed of rigid, polystyrene divinylbenzene particles having a mean diameter between 10 and 20 micrometers, defining a bimodal or multimodal pore structure and comprising chemically active regions linked to the surface of the particles for reversibly sorbing biological molecules, said matrix being characterized in that

CLPV:

a packed bed of rigid particles comprising an inorganic material, the particles having a mean diameter within the range of 10 micrometers to 100 micrometers and defining a bimodal or multimodal pore structure, one set of pores being particle transecting throughpores having a mean diameter greater than at least 4000 .ANG., another set of pores being subpores in fluid communication with the throughpores, and, disposed within at least said subpores, reactive groups comprising one of anionic sulfonate groups, cationic quaternary ammonium groups, immunoglobulines, or hydrocarbons, the ratio of the means diameter of the particles to the mean diameter of a throughpores passing through the particle being sufficient to permit convective transport of biological molecules into the throughpores and the subpores for chromatographic separation so that, at flow rates greater than 100 cm./hr, the rate of biological molecule transport into the throughpores is dependent on the velocity of liquid passing through the bed, and sorption capacity remains

substantially constant over a range of flowrates.

CCOR:

210/198.2

DOCUMENT-IDENTIFIER: US 5316680 A

TITLE: Multimodal chromatographic separation media and process for using same

TTL:

Multimodal chromatographic separation media and process for using same

ABPL:

A process for carrying out in a consecutive fashion different modes of chromatographic separation in a liquid chromatography column using a single separation medium is disclosed. Separation media for use in such multimodal separations are also disclosed.

BSPP:

It may be possible to use combinations of different separation media in different columns for multimodal separations. An example of this multiple column bimodal separation was described recently by Wheatley J. B., J. Chromatogr., 603 (1992) 273. The bimodal separation of small molecules in one column packed with one separation medium and based on sequential multimodal elution was described by Little E. L., Jeanson M. S., Foley J. P.; Anal Chem., 63, 1991, 33. They combined ion-exchange and reversed phase chromatography for the separation of a complex sample containing two groups of compounds: charged and non-polar. The use of two different gradients, i.e. a pH gradient and a methanol gradient, resulted in the separation of the charged molecules, followed by the separation of the neutral molecules after switching to the second mobile phase. This approach makes use of imperfect surface functionalization of porous silica beads which contained C.sub.1, C.sub.8 or C.sub.18 groups together with the original acidic surface silanol groups. Similarly, the DIONEX OmniPack PAX-500 column is packed

with non-porous poly[styrene-divinylbenzene] beads coated on the bead surface with attached ion-exchange latex particles (as described by the DIONEX booklet). Here again, the coating of the bead surface is imperfect and it is the non-covered hydrophobic areas of the original non-porous beads that are used for separation in the second mode. This approach excludes combinations not involving the reversed phase mode (the original ST-DVB surface remains non-polar even after attachment of latex particles) as well as any size exclusion separation.

BSPP:

This multimodal separation process is able to achieve separation in a single column in a consecutive operation because of the properties of the separation medium. The separation medium generally comprises a porous material which has been pretreated so that it has at least two different types of surface groups which have different functionalities. These different surface groups are disposed in different size range pores within the porous material. Pore size as used herein can mean a single measured average size, for example, 25 nm, but in most cases it means a particular range of sizes, for example, 50-500 nm. An example of such a porous material of the present invention is one wherein there are hydrophilic surface groups in pores having a size of from about 5-25 nm and hydrophobic surface groups in pores ranging in a size of from about 50-500 nm. Another example is a material having hydrophilic groups in pores of about 5-25 nm and hydrophobic groups in pores of about 50-500 nm. As a result of the different functionalities of the surface groups, molecules that have different affinities to such different surface groups may be separated during different modes of separation, which may be carried out in a consecutive fashion. As

used herein, different molecules means molecules of different sizes, different chemical affinities, different structures, compositions, polarities, chiralities, activities, etc.

DEFF:

The key in selecting a mobile phase for a multimodal separation is the consecutive use of mobile phases, in each individual mode, that do not interfere with the absorption of compounds to be separated in any of the subsequent modes. Otherwise, the separation will not be multimodal and one group of compounds will leave the column without any separation, as documented in Example 6 and FIG. 7. Under this assumption, even a mobile phase for trimodal separation is easily designed by a person skilled in the art of liquid chromatography.

DEFF:

Reaction Schemes 1-4 not only describe the particular sets of reactions leading to multimodal separation media but they also show the concepts of making such media in general. The starting polymer must be porous with relatively broad pore size distribution and possess reactive groups on the surface of the pores. Typically, the pore-size selectivity of the modification reactions are controlled by the molecular weight of the catalyst or reagent used in the particular modifying reaction and by the solvent. The number of modes accommodated in a separation medium is theoretically not limited but practically will rarely exceed three. The most important part in the reaction strategy for preparation of a multimodal medium is the right choice of the path. The product of a given reaction affecting pores of a given size should not affect the groups already built up in the previous reaction step within pores of a different size.

DEPR:

The multimodal separation process of the present invention may even use very tiny differences between the separation modes as is the case with reversed phase and hydrophobic interaction chromatography. The separation medium can be prepared by a set of reactions shown in Reaction Scheme 5.

CCKE:

210/198.2

ORPL:

Little, "Sequential Multimodal Elution for Pseudomultidimensional Liquid Chromatography on a Single Column," Anal. Chem., 63, (1991) pp. 33-44.

DOCUMENT-IDENTIFIER: US 5228989 A
TITLE: Perfusive chromatography

DEPR:

Broadly, in accordance with the invention, perfusion chromatography is practiced by passing fluids at velocities above a threshold level through a specially designed matrix characterized by a geometry which is bimodal or multimodal with respect to its porosity. Perhaps the most fundamental observation relevant to the new procedure is that it is possible to avoid both the loss of capacity characteristic of convection bound systems and the high plate height and bandspreading characteristics of diffusion bound systems. This can be accomplished by forcing chromatography fluids through a matrix having a set of larger pores, such as are defined by the interstices among a bed of particles, and which determine pressure drops and fluid flow velocities through the bed, and a set of pores of smaller diameter, e.g., anisotropic throughpores. The smaller pores permeate the individual particles and serve to deliver chromatography fluids by convection to surface regions within the particle interactive with the solutes in the chromatography fluid.

DEPR:

From the foregoing description many of the basic engineering goals to be pursued in the fabrication of matrix materials suitable for the practice of perfusion chromatography will be apparent to those skilled in the art. Thus, what is needed in practice perfusion chromatography is a matrix which will not crush under pressure having a bimodal or preferably multimodal pore structure and as large a surface area per unit volume as possible. The first and second pore sets which give the material its bimodal flow properties

must have mean diameters relative to each other so as to permit convective flow through both sets of pores at high V.sub.beds. The provision of subpores in the matrix is not required to conduct perfusion chromatography but is preferred because of the inherent increase in surface area per unit volume of matrix material such a construction provides.

DEPP:

In contrast to the PL 4,000 material, which, with respect to its pore structure, is multimodal, a more ideal perfusive particle might comprise a plurality of sets of throughpores and subpores of differing mean diameters. A bimodal pore size distribution can be achieved in such particle by mixing equal ratios of particles having two discrete pore sizes or by engineering this feature at the polymerization stage. Ideally, mean diameter ratio between throughpore subsets would be less than 10, the mean diameter ratio between the smallest throughpore sets and the subpores would be less than 20, and the mean diameter ratio between the first pore set, i.e., the interstices among the particles making up the matrix, and the largest throughpore subset would be less than 70, and preferably less than 50. A multimodal material might be produced by agglomerating 500 .ANG. porons to form approximately 1 .mu.m clusters, which in turn are agglomerated to form 10 .mu.m aggregates, which in turn may be aggregated to form 100 .mu.m particles. In such a design, the 1 .mu.m clusters would have interstices of a mean diameter in the order of a few hundred .ANG.. These would define the surface area, provide a very high surface area. Diffusive transport within these pores would rarely have to exceed a distance of 0.5 .mu.m or 5,000 .ANG.. Interstices among the 1 .mu.m clusters making up the 10 .mu.m aggregates would permit convective flow to feed

the diffusive pores. These would be on the order of 0.3 .mu.m in diameter.

These throughpores, in turn would be fed by larger pores defined by

interstices among the 10 .mu.m particles making up the 100 .mu.m particles.

These would have a mean diameter on the order of 35 .mu.m.

CCOR:

210/198.2

DOCUMENT-IDENTIFIER: US 5104530 A
TITLE: Chromatography column with carbonaceous adsorbents from
pyrolyzed
polysulfonated polymers

ABFL:

Carbonaceous adsorbent particles having multimodal pore size, including micropores and macropores, with improved adsorptive and separative properties, are prepared by partial pyrolysis of polysulfonated macroporous precursor resins, said resins being in turn derived from macroporous poly(vinylaromatic) resins. The particles may be further treated by activating with reactive gases or by functionalization.

ESPE:

British Patent No 1,525,420, in a broad description of method for rendering infusible various porous high molecular weight compounds (including macroporous resins), and then calcining them, relates techniques for polysulfonation earlier described by Corte et al. among those suitable for creating infusibility. No characterization data are given for the polymer prior to calcination. Preferred infusibility reactants are sulfur trioxide, sulfuric acid, or chlorosulfonic acid. This reference discloses pyrolysis of macroporous resins treated with 15% fuming sulfuric acid and pyrolyzed, and describes an experimental method for determining the porosity of the pyrolyzed material down to 2-5 nm. The results described in the tables of the reference show the absence of any porosity development below 2 nm, and multimodal porosity is not taught. In contrast, Neely in the cited references fully shows the development of microporosity for monosulfonated macroporous resins. Further, the British patent is silent about the processing advantages observed

in pyrolysis of polysulfonated resins.

CLPE:

1. A chromatographic column packed with the carbonaceous adsorbent particles which comprise the product of controlled pyrolysis of a polysulfonated macroporous crosslinked, vinylaromatic polymer, the particles having multimodal pore-size distribution and a minimum micropore volume of about 0.02 cm.sup.3 /g.

CLPE:

4. A chromatographic column packed with the carbonaceous adsorbent particles which comprise the product of controlled pyrolysis of a polysulfonated macroporous crosslinked, vinylaromatic polymer, the particles having multimodal pore-size distribution and a minimum micropore volume of about 0.02 cm.sup.3 /g, wherein the particles are treated, subsequent to pyrolysis, with adsorbable reactive agent.

COOR:

210/198.2

DOCUMENT-IDENTIFIER: US 5071547 A

TITLE: Column chromatographic column apparatus with switching capability

DEPR:

The fluid conduit assembly or means is comprised of conduit piping connecting the dual columns, detector, multi-valve arrangement in a similar manner to that of FIG. 2 with three exceptions. First, the fluid conduit assembly in addition to conduits 11A through 11L as in FIG. 2, also has conduits 11X through 11Z and 11Z'. Second, there are two valves present in what conceptually can be referred to as each valve set 12A and 12B of FIG. 2. The third exception is the presence of an additional valve 42 that isolates the detector from the pressure of the DCC Apparatus. Although valves 12iv and 12v can be indicated as being in the valve set 12A of FIG. 2 and valves 12vi and 12vii can be indicated as being in the valve set 12B of FIG. 2, these valves may exist with independent identity from these valve sets. In other words these valves may be just a plurality of valves without valves 12iv and 12v as well as the two valves 12vi and 12vii perform the same function as valves sets 12A and 12B, respectively. Valve 42 is any multi-port and multimodal valve known to those skilled in the art as are valves 12iv through 12vii, and valve 42 can be considered a part of the multi-valve arrangement 12. As with the DCC Apparatus of FIG. 2, that of FIG. 3 has each valve with an actuating connection with a control line. As valve 42 is a multi-port valve, it has an actuating connection 14A as valves 12iv-12vii and 42 are connected by connections 14J, 14K, 14L, 14M, and 14N, respectively. Valves 12iv and 12v are connected together for pressurized fluid passage between them by conduit 11X,

and in a similar manner valves 12vi and 12vii are connected by conduit 11Y.

CCOR:

210/198.2

DOCUMENT-IDENTIFIER: US 5019270 A
TITLE: Perfusive chromatography

DEPR:

Broadly, in accordance with the invention, perfusion chromatography is practiced by passing fluids at velocities above a threshold level through a specially designed matrix characterized by a geometry which is bimodal or multimodal with respect to its porosity. Perhaps the most fundamental observation relevant to the new procedure is that it is possible to avoid both the loss of capacity characteristic of convection bound systems and the high plate height and bandspreading characteristics of diffusion bound systems. This can be accomplished by forcing chromatography fluids through a matrix having a set of larger pores, such as are defined by the interstices among a bed of particles, and which determine pressure drops and fluid flow velocities through the bed, and a set of pores of smaller diameter, e.g., anisotropic throughpores. The smaller pores permeate the individual particles and serve to deliver chromatography fluids by convection to surface regions within the particle interactive with the solutes in the chromatography fluid.

IEPR:

From the foregoing description many of the basic engineering goals to be pursued in the fabrication of matrix materials suitable for the practice of perfusion chromatography will be apparent to those skilled in the art. Thus, what is needed to practice perfusion chromatography is a matrix which will not crush under pressure having a bimodal or preferably multimodal pore structure and as large a surface area per unit volume as possible. The first and second pore sets which give the material its bimodal flow properties

must have mean diameters relative to each other so as to permit convective flow through both sets of pores at high V.sub.beds. The provision of subpores in the matrix is not required to conduct perfusion chromatography but is preferred because of the inherent increase in surface area per unit volume of matrix material such a construction provides.

DEPR:

In contrast to the PL 4,000 material, which, with respect to its pore structure, is multimodal, a more ideal perfusive particle might comprise a plurality of sets of throughpores and subpores of differing mean diameters. A bimodal pore size distribution can be achieved in such particle by mixing equal ratios of particles having two discrete pore sizes or by engineering this feature at the polymerization stage. Ideally, mean diameter ratio between throughpore subsets would be less than 10, the mean diameter ratio between the smallest throughpore sets and the subpores would be less than 20, and the mean diameter ratio between the first pore set, i.e., the interstices among the particles making up the matrix, and the largest throughpore subset would be less than 70, and preferably less than 50. A multimodal material might be produced by agglomerating 500 .ANG. porons to form approximately 1 .mu.m clusters, which in turn are agglomerated to form 10 .mu.m aggregates, which in turn may be aggregated to form 100 .mu.m particles. In such a design, the 1 .mu.m clusters would have interstices of a mean diameter in the vicinity of 0.5 .mu.m or 5,000 .ANG.. These would define the surface area. Diffusive transport within these pores would rarely have to exceed a distance of 0.5 .mu.m or 5,000 .ANG.. Interstices among the 1 .mu.m clusters making up the 10 .mu.m aggregates would permit convective flow to feed

the diffusive pores. These would be on the order of 0.3 .mu.m in diameter.

These throughpores, in turn would be fed by larger pores defined by

interstices among the 10 .mu.m particles making up the 100 .mu.m particles.

These would have a mean diameter on the order of 35 .mu.m.

CCKR:

210/198.2

	Type	L #	Hits	Search Text	DBs	Time Stamp
1	BRS	L1	37759	monolith\$3	USPAT	2001/12/18 15:45
2	BRS	L2	1551	210/198.2.ccls.	USPAT	2001/12/18 15:46
3	BRS	L3	40	1 and 2	USPAT	2001/12/18 15:59
4	BRS	L4	942	multimodal	USPAT	2001/12/18 15:59
5	BRS	L5	12	2 and 4	USPAT	2001/12/18 15:59

	Comments	Error Definition	Errors
1			0
2			0
3			0
4			0
5			0

L Number	Hits	Search Text	DB	Time stamp
1	37759	monolith\$3	USPAT	2001/12/18 15:45
2	1551	210/198.2.ccls.	USPAT	2001/12/18 15:46
3	40	monolith\$3 and 210/198.2.ccls.	USPAT	2001/12/18 15:59
4	942	multimodal	USPAT	2001/12/18 15:59
5	12	210/198.2.ccls. and multimodal	USPAT	2001/12/18 15:59

L Number	Hits	Search Text	DB	Time stamp
1	37759	monolith\$3	USPAT	2001/12/18 15:45
2	1551	210/198.2.ccls.	USPAT	2001/12/18 15:46
3	40	monolith\$3 and 210/198.2.ccls.	USPAT	2001/12/18 15:59
4	942	multimodal	USPAT	2001/12/18 15:59
5	12	210/198.2.ccls. and multimodal	USPAT	2001/12/18 15:59